

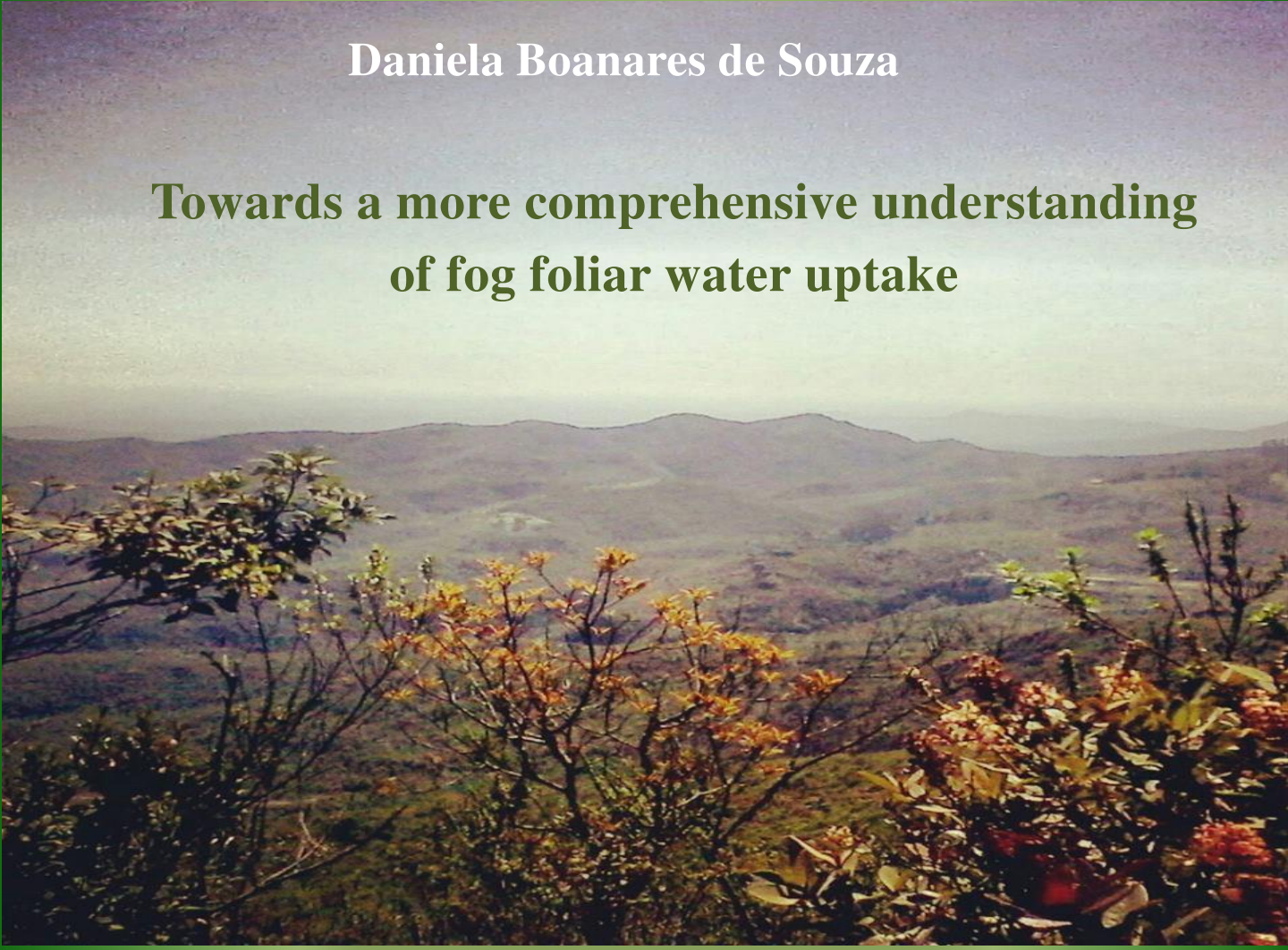


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Programa de Pós Graduação em Biologia Vegetal

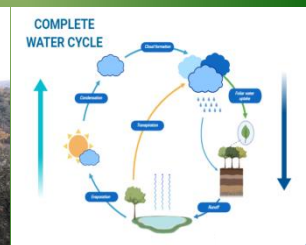
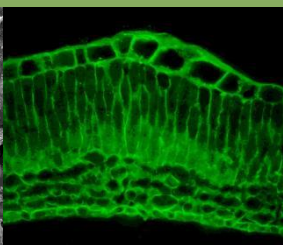
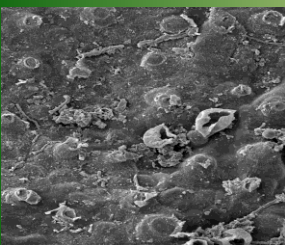


**Daniela Boaneres de Souza**

# **Towards a more comprehensive understanding of fog foliar water uptake**



**Orientador: Prof. Dr. Marcel Giovanni Costa França**  
**Coorientadora: Prof<sup>a</sup> Dr<sup>a</sup> Rosy Mary dos Santos Isaias**



**Belo Horizonte, MG**

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**UNIVERSIDADE FEDERAL DE MINAS GERAIS**  
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Departamento de Botânica

**Programa de Pós-Graduação em Biologia Vegetal**



**UFMG**

**DANIELA BOANARES DE SOUZA**

**TOWARDS A MORE COMPREHENSIVE  
UNDERSTANDING OF FOG FOLIAR WATER UPTAKE**

**Tese apresentada ao Programa de Pós  
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Departamento de Botânica do Instituto de  
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**Área de concentração: Fisiologia e Ecologia Vegetal**

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## RESUMO

A absorção foliar de água tem um papel importante na fisiologia de algumas espécies, principalmente aquelas que ocorrem em ambientes com presença de neblina. A neblina pode ser a única fonte de água para uma determinada vegetação, principalmente na estação seca, tornando essa condição climática uma fonte alternativa de água para a vegetação. Assim, o objetivo desta tese foi avaliar atributos ultraestruturais, fisiológicos, bioquímicos e fenológicos de espécies com diferentes capacidades de absorção foliar de água que ocorrem em ambientes sujeitos à neblina. Foi verificado que a composição química da cutícula e a proporção de componentes da parede celular interferem na absorção foliar de água. A assimilação de carbono é, momentaneamente, afetada pela absorção foliar de água, mas com diminuição na perda de água pela transpiração. O status hídrico, avaliado a partir do potencial hídrico, foi beneficiado pela absorção foliar de água, demonstrando valores similares de potencial hídrico na estação chuvosa e na estação seca com neblina. A absorção foliar de água aumentou a termotolerância, diminuiu a possibilidade de fotoinibição e melhorou o sistema antioxidante, principalmente nas espécies que absorvem grande quantidade de água. A presença de neblina, juntamente com a absorção de água pelas folhas interfere na fenologia de uma espécie decídua, retardando a queda de suas folhas. Assim, concluímos que a absorção foliar de água, principalmente de neblina tem um papel fundamental para a manutenção de várias plantas em seus habitats naturais; além disso, a estratégia de absorver maior quantidade de água, apesar de lentamente seria a melhor estratégia para as plantas lidarem, a longo prazo, com as mudanças climáticas previsíveis.

Palavras-chave: neblina, absorção foliar, fotossíntese, cutícula, fenologia

## ABSTRACT

Foliar water uptake plays an important role in the physiology of some species, especially those that occur in foggy environments. Fog may be the only source of water for some particular vegetation, especially in the dry season, making the occurrence of fog an alternative source of water for the vegetation. Thus, the objective of this thesis was to evaluate ultrastructural, physiological, biochemical and phenological attributes of plant species with different leaf absorption capacities that occur in environments subjected to fog. The chemical composition of the cuticle and the proportion of cell wall components interfere with leaf water uptake. Carbon assimilation is momentarily affected by leaf water absorption, but there has been a decrease in water loss through transpiration. The water status, evaluated through water potential, was benefited by leaf water uptake, demonstrating similar values of water potential in rainy season and in dry season with fog. Foliar water uptake increased thermotolerance, reduced the possibility of photoinhibition and improved antioxidant system, especially in species that absorb a large amount of water. The presence of fog, along with the absorption of water by the leaves, interferes with the phenology of a deciduous plant species, delaying the fall of its leaves. Thus, we conclude that leaf water uptake, especially fog, plays a key role in maintaining various plants in their habitats. The strategy of absorbing more water, although slowly, would be the best strategy for plants in the long term to cope with possible climate changes.

Keywords: fog, leaf absorption, photosynthesis, cuticle, phenology

## INTRODUÇÃO GERAL

Diferentes estratégias moleculares, fisiológicas e bioquímicas são acionadas, rapidamente ou mais tardiamente, quando as plantas enfrentam estresse por falta de água. Algumas das estratégias podem ser facilmente observadas, quando, p.ex., plantas perdem água pelos estômatos na busca pelo equilíbrio do status hídrico foliar. Analogamente, vem sendo demonstrado que existem diferentes estratégias de captura de água pelas folhas, o que caracteriza o movimento contrário da água ao fluxo no sistema solo-planta-atmosfera. Esta absorção foliar acontece quando a água ultrapassa a superfície da folha em resposta ao gradiente de potencial hídrico difundindo-se para os tecidos internos da folha (Gouvra & Grammatikopoulos 2003). A absorção foliar de água pode ocorrer durante e logo após as diversas formas de precipitação, como a neblina.

Um exemplo de ecossistema tropical no qual a neblina é um evento atmosférico frequente é o campo rupestre ferruginoso, que ocupa nos topos de morros, em geral com altitudes acima de 1000 metros, no Quadrilátero Ferrífero. Devido à marcada sazonalidade de precipitação de chuvas e pela presença de fisionomias campestres ocorrendo sobre afloramentos rochosos ricos em ferro como itabiritos ou cangas, ou seja, predominantemente sobre solos rasos e descontínuos, acredita-se que a vegetação sobre campos rupestres ferruginosos experimente períodos de déficit hídrico durante a estação seca (Benites et al. 2003). Já foi visto que nos períodos sem neblina, ambientes tropicais de altitudes podem ser considerados áridos, caracterizados por uma alta demanda evaporativa (Leuschner, 2000).

A absorção foliar de água, a partir de agora referida como FWU (em inglês “foliar water uptake”), pode proporcionar uma vantagem ecológica para as plantas que ocorrem em locais que apresentam déficit hídrico, pelo menos sazonalmente, desempenhando

papel importante na sua sobrevivência e crescimento (Limm et al. 2009; Eller et al. 2013). Das 1100 espécies estimadas para os campos rupestres ferruginosos (Jacobi e Carmo, 2012), foi verificado que pelo menos onze espécies têm capacidade de absorver água pelas folhas, mas com diferentes estratégias para esta entrada de água. Algumas espécies absorvem água rapidamente em quantidades menores enquanto outras espécies absorvem água lentamente em quantidades maiores (Boanares et al. 2018 a, b). Como existe um consenso de que FWU pode ser a norma e não a exceção em plantas (Berry et al. 2019), é de extrema importância o conhecimento mais aprofundado sobre como ocorre e o que influencia este mecanismo nas diferentes espécies de plantas.

Desta forma, o objetivo desta tese foi responder se diferenças nas características ultraestruturais da folha interferem na capacidade de FWU e se essas diferentes capacidades influenciam parâmetros ecofisiológico e ecossistêmicos das espécies estudadas. A tese é apresentada na forma de 7 capítulos conforme os objetivos propostos a seguir:

Como a cutícula é a primeira barreira física para a entrada de água na folha, o **Capítulo 1** aborda a relação da composição química da cera cuticular e das ornamentações da cera epicuticular com as diferentes capacidades de FWU. Considerando que a parede celular é a próxima barreira física à FWU, o **Capítulo 2** verifica a relação entre a composição química da parede celular, quanto aos seus componentes estruturais, pectinas e celulose, com as estratégias de FWU. Uma vez avaliada a interferência da química da cutícula e dos componentes da parede celular nas diferentes estratégias de FWU, o objetivo seguinte foi responder como estas estratégias podem interferir na fisiologia destas espécies. Assim, o **Capítulo 3** avalia se as diferentes estratégias de FWU podem interferir no balanço hídrico e de carbono das espécies ao longo de um ano, contemplando as estações climáticas seca e chuvosa. A

premissa central deste capítulo é de que as mudanças climáticas globais podem interferir no regime hídrico e térmico nos diferentes ecossistemas. Por isso, o **Capítulo 4** aborda diferentes estratégias de FWU e sua relação com as respostas de termotolerância e de fotoinibição. No **Capítulo 5**, verifica-se se as mesmas estratégias de FWU podem minimizar o metabolismo oxidativo induzido pelos respectivos estresses ambientais, regime hídrico e amplitude térmica.

Uma vez entendido como parâmetros ultraestruturais e fisiológicos podem interferir na FWU de espécies de um ambiente tropical de altitude, partimos para verificar alguns desses parâmetros em plantas com diferentes respostas à presença de neblina em um ecótono Mediterrâneo - Temperado. Neste ambiente, já foi verificado uma redução dos eventos de neblina, impactando de forma distinta o crescimento destas espécies.

Desta forma, outro objetivo da tese foi verificar a importância da FWU e da neblina na ecofisiologia de espécies ocorrentes neste ecótono tendo como cenário questões sobre Mudanças Climáticas Globais. Assim, no **Capítulo 6** verificamos se a ocorrência de neblina e, conseqüentemente, sua absorção pelas folhas interfere na fisiologia e fenologia de duas espécies que coexistem no ecótono Mediterrâneo-Temperado.

Por fim, o **Capítulo 7** propõe uma abordagem multidisciplinar focando na genética, ultraestrutura, anatomia, fisiologia e ecologia buscando o maior entendimento do fluxo reverso de água atmosfera-planta-solo, contrário ao que se observa na literatura clássica considerando o sistema solo-planta-atmosfera.

Assim, os objetivos específicos foram:

- Avaliar se a composição química e a ornamentação da cutícula interferem na FWU
- Avaliar se as diferentes proporções de celulose e pectina na parede celular interferem na FWU

- Verificar se as diferentes capacidades de FWU influenciam no balanço hídrico e de carbono e na termotolerância das espécies.
- Analisar se o metabolismo oxidativo tem relação com as distintas capacidades de FWU.
- Verificar o comportamento fisiológico e fenológico de espécies com distintas respostas à presença de neblina.

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## Lista de abreviaturas

FWU - foliar water uptake	PD ds - Predawn dry season
LWU - leaf water uptake	PD ds fog - Predawn dry season with fog
CW - cuticular wax	MD ds - Midday dry season with fog
EW - epicuticular wax	MD ds fog – Midday dry season with fog
$C_{max}$ - maximum leaf water absorption	PD rs - Predawn rainy season
$k$ - foliar water absorption speed	MD rs - midday rainy season
$C_{max}^L$ - total amount of water leaf lost	SPAC - soil-plant-atmosphere continuum
$k^L$ - water leaf lost speed	Fv/Fm - potential quantum yield of photosystem II
GLM - General Linear model	$T_{50}$ - temperature of Fv/Fm 50% decline
LMM - linear mixed model	PSII - photosystem II
HGA - Homogalacturonans	ROS - reactive oxygen species
RG - rhamnogalacturonans	CAT - catalase
LEM – leaf specific mass	SOD - superoxide dismutase
LSA - leaf specific area	APX - ascorbate peroxidase
RWC - relative water content	LQ - species that absorb less water more quickly
WSD - water saturation deficit	MS - species that absorb more water more slowly
$A$ - net CO <sub>2</sub> assimilation	PRI - photochemical reflectance index
$E$ - transpiration	NDVI - normalized difference vegetation index
$g_s$ - stomatal conductance	APSC – atmosphere-plant-soil continuum
$C_i / C_a$ - internal carbon and ambient carbon ratio	
WUE - water-use efficiency	
iWUE – intrinsic water-use efficiency	
$\Psi_w$ – water potential	

*Cuticular wax composition and wax crystals reveal different capacities to foliar water uptake*

1 **Cuticular wax composition and wax crystals reveal different capacities**  
2 **to foliar water uptake**

3  
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21  
22 **ABSTRACT**

23  
24 Leaf cuticle is the first physical barrier for foliar water uptake (FWU), since its main  
25 function is to prevent excessive water loss through transpiration. We assume that not

26 only the cuticle itself but the chemical composition of the cuticular wax and the  
27 ornamentation of the epicuticular wax must influence the different capacities of FWU.  
28 To address this hypothesis, the chemical analysis of cuticular wax (CW) was performed  
29 by gas chromatography-mass spectrometry/flame ionization detection, and the  
30 epicuticular wax ornamentation was investigated by scanning microscopy on six species  
31 with different FWU capacities. The total chemical composition of CW related to the  
32 speed of foliar water absorption but did not relate to the amount of water absorbed.  
33 However, the classes of the chemical compounds analyzed separately had no relation to  
34 FWU. Current chemical data suggest that the permeability of CW does not regard to the  
35 amount of CW or to a specific class of chemical compound but to the presence and  
36 proportion of all classes together. The greater or lesser FWU capacity is related to a  
37 three-dimensional network of the chemical molecules. Also, the three-dimensional  
38 ornamentation of EW indicates the relation of the higher ornamentation complexity with  
39 the slowest water absorption capacity, due to the higher hydrophobicity of the leaf  
40 surface. Together, the composition of the CW and the crystals wax of the EW  
41 differentially affect the FWU of the studied species.

42

43 **Key words:** Foliar water uptake; ultrastructural traits; epicuticular ornamentation; gas  
44 chromatography; *Campo rupestre*

45

46 **List of abbreviation:**

47  $C_{\max}$  - maximum water content in leaf lamina

48  $k$  - water leaf uptake speed

49 CW - cuticular wax

50 EW – epicuticular wax

51

52 **Author contributions**

53

54 D.B., R.M.S.I. and M.G.C.F. designed the research. D.B., H.C.S, A.R.K., A.B. and  
55 A.X.S collected the data for the research, and D.B., L.P.S.P and D.R.M.N analyzed the  
56 data. D.B., M.R., R.M.S.I. and M.G.C.F wrote the manuscript.

57

58 **HIGHLIGHTS**

59

- 60 • Cuticular and crystals waxes was related to foliar water uptake (FWU)
- 61 • Chemical compounds analyzed separately were not related to FWU
- 62 • Complexity in the ornamentation gives greater hydrophobicity to the leaf surface

63

64 **INTRODUCTION**

65

66 Foliar water absorption (FWU) occurs when there is a difference of vapor pressure in  
67 atmosphere and inside the leaf, inducing a difference in water potential, which promote  
68 water o overtake the boundary layer and into the epidermal cells, causing hydration of  
69 leaf tissues (Todd and Dowson, 2004). The FWU can play an important role in the  
70 water relations of many terrestrial plants, especially in humid environments and in  
71 ecosystems where, in addition to rainfall, water is provided by mist, fog and dew  
72 causing frequent leaf wetting (Breshears et al., 2008; Limm et al., 2009; Limm and  
73 Dawson, 2010; Goldsmith et al., 2013; Mayr et al., 2014; Boanares et al., 2019).

74 Due to the emergence of new investigation methods, researches into the water  
75 pathways inside the leaf have increased in recent years (Eller et al, 2016; Goldsmith et

76 al., 2017, Boanares et al, 2018; Brinks et al, 2019). The mechanism of leaf water entry  
77 is of great interest and several pathways have been proposed: water movement directly  
78 through the cuticle (Goldsmith et al., 2013; Ritpitakphong et al., 2016), through the  
79 pores (Schonherr, 2006), trichomes (Mencuccini and Ball, 2016; Boanares et al., 2018),  
80 hydathodes (Martin and von Willert, 2000), and also by stomata (Burkhardt et al., 2012;  
81 Berry et al., 2014; Fernandez et al., 2017).

82 Such observations about the pathway of water raises critical leaf surface issues  
83 associated to the FWU capacity, such as morphological (leaf angle and size), anatomical  
84 (stomatal location, leaf tissue thickness) and cuticle traits (Goldsmith et al., 2017;  
85 Boanares et al., 2018). The cuticle is the first physical barrier that water faces to enter  
86 leaf tissues. The plant cuticle is most typically associated to a fixed barrier to excessive  
87 water loss from transpiration. However, several secondary functions have evolved, such  
88 as the defense against pests and pathogens, the avoidance of dust and other debris due to  
89 many epicuticular crystals that promote a self-cleaning surface, and may also play the  
90 role of filtering excessive light radiation (Yeast and Rose, 2013).

91 Plant cuticles contain, beyond the cutin, wax as the second largest chemical  
92 component (Riederer and Schreiber, 2001). Wax composition may vary substantially  
93 with species, ontogeny, and environmental growth conditions (Jenks and Ashworth,  
94 1999). This set of compounds is characterized by unbranched and fully saturated  
95 hydrocarbon backbones, which may have a primary oxygen-containing functionality,  
96 that is, a hydroxyl, carbonyl or carboxyl terminal group. Therefore, plant cuticle waxes  
97 are typically mixtures of primary n-alcohols, n-aldehydes and fatty acids, as well as n-  
98 alkanes and various lipophilic secondary metabolites such as pentacyclic triterpenoids  
99 and flavonoids, which may also be important components (Jetter et al., 2006; Yeast and  
100 Rose, 2013). Each of these classes of compounds comprises a homologous series of

101 chemical compounds with chain lengths ranging from 20 to nearly 40 carbons, and a  
102 species-specific matrix that may comprise more than 50 chemical compounds (Riederer  
103 and Schreiber, 2001).

104 When considering cuticular permeability (Kerstiens, 1996; Riederer and Schreiber,  
105 2001), interspecific and intraspecific differences in water entry capacity may partly  
106 result from differences in chemical composition, epicuticular wax morphology among  
107 species, and tissue age (Kerstiens, 2006). Some studies have evidenced that cuticular  
108 permeability to water is mainly determined by genetic control, and only slightly subject  
109 to environmental influence. Therefore, cuticular water permeability may be related to  
110 the plant life strategy to deal with different environmental constraints (Bueno et al.,  
111 2019).

112 As there is a consensus that FWU may be the norm rather than the exception in plants  
113 (Berry et al., 2019), in-depth knowledge of how it occurs and what influences FWU at  
114 the cuticular level is of utmost importance. Differences in the FWU capacity of species  
115 that occur in a Brazilian altitude ecosystem revealed species with low and fast water  
116 absorption by leaves (Boanares et al, 2018a,b) but the influence of cuticular  
117 morphology and cuticular wax chemical composition differences on leaf absorption  
118 speed has not been explored, yet. Taking for granted that the different water absorption  
119 speeds relate both to the chemical composition of cuticular wax (CW) and to the  
120 morphology of epicuticular wax, gas chromatography coupled to mass spectrometry or  
121 flame ionization detection and scanning electron micrography (SEM) were used to  
122 evaluate the composition of CW and the epicuticular wax (EW) ornamentation of six  
123 species of a Brazilian altitude ecosystem.

124

## 125 **MATERIAL AND METHODS**

126

127 *Plant material*

128

129 The studied species are present in the ferruginous *Campo Rupestre*. This ecosystem is a  
130 Brazilian OCBIL, characterized by herbaceous and shrub species growing on rock  
131 outcrops of itabirites or *cangas* of shallow soils (Hopper 2009; Valim *et al.*, 2013;  
132 Silveira *et al.*, 2016). These plant species were selected according to their different  
133 water absorption capacity from calculated by a differential equation, which provides  
134 data on maximum leaf water absorption ( $C_{\max}$ ) and leaf uptake velocity ( $k$ ) ( $\text{g g}^{-1}$  dry  
135 mass) (Liang *et al.*, 2009; Boanares *et al.*, 2018b). The species *Leandra australis*  
136 (Melastomataceae), *Byrsonima variabilis* (Malpighiaceae) and *Ocotea pulchella*  
137 (Lauraceae) have a higher speed of leaf absorption and uptake a smaller amount of  
138 water ( $k=0.250\pm 0.02$ ;  $C_{\max}=0.140\pm 0.02$ ). However, the species *Trembleya laniflora*  
139 (Melastomataceae), *Pleroma heteromallum* (Melastomataceae) and *Senna reniformis*  
140 (Fabaceae) are slow in absorption, but they uptake a greater amount of water through  
141 the leaves than the species of the first strategy ( $k=0.067\pm 0.06$ ;  $C_{\max}=0.443\pm 0.19$ ).

142

143 *Chemical analyses of cuticular waxes*

144

145 Cuticular waxes were extracted by dipping the whole leaf (except the wounds of cut  
146 petioles) twice into chloroform ( $\geq 99.8\%$ , Roth) at room temperature for 1.5 min. 10  $\mu\text{g}$   
147 of n-tetracosane (C24;  $\geq 99.5\%$ , Sigma-Aldrich) was added as an internal standard and  
148 the solutions were reduced to dryness under a gentle flow of nitrogen. Dry cuticular wax  
149 samples were derivatized with N,O bis(trimethylsilyl)trifluoroacetamide (BSTFA,  
150 Marchery-Nagel) in dry pyridine ( $\geq 99.5\%$ , Roth) at 70°C for 30 min. Quantification of

151 cuticular wax compounds was performed with a gas chromatograph equipped with a  
152 flame ionization detector and an on-column injector (7890A, Agilent Technologies).  
153 Separation of compounds was carried out on a fused-silica capillary column (DB1-ms,  
154 30 m length × 0.32 mm inner diameter, 0.1 µm film thickness, Agilent Technologies)  
155 with hydrogen as a carrier gas. The temperature program consisted of injection at 50°C  
156 for 2 min, raised by 40°C min<sup>-1</sup> to 200°C, held at 200°C for 2 min, and then raised by  
157 3°C min<sup>-1</sup> to 320°C, and held at 320°C for 30 min. Qualitative analysis was carried out  
158 using a gas chromatograph equipped with a mass spectrometric detector (5975 iMSD,  
159 Agilent Technologies) following the same gas chromatographic conditions but using  
160 helium as the carrier gas. Cuticular wax compounds were identified comparing a query  
161 mass spectrum with reference mass spectra in a library via spectrum matching and  
162 quantitated against the internal standards (Bueno et al., 2019).

163

#### 164 *Scanning electron microscopy*

165

166 For scanning electron microscopy (SEM), leaves collected and fixed in Karnovsky (pH  
167 7.2 in 0.1 M phosphate buffer; modified from Karnovsky 1965) were used to remove  
168 one fragment of 0.5 cm from the middle part of leaves of 4 individuals of each species.  
169 The material was dehydrated in increasing ethanol solutions (70%, 90% and 100%) and  
170 then the fragments were mounted in stubs and brought to dryness to the critical point  
171 with liquid CO<sub>2</sub> (Balzers CPD 020). Subsequently, the surfaces of the fragments were  
172 covered by a thin layer of metallic gold (Edwards Scancoat Six) and examined by  
173 scanning electron microscope (SEM) - DSM 950 Carl Zeiss.

174

#### 175 *Data analysis*

176

177 We first explored the patterns of chemical differentiation between species by calculating  
178 their pairwise Euclidean Distance in chemical composition, and performing non-metric  
179 multidimensional scaling (NMDS) (McCune & Grace, 2002). We explored the results  
180 visually by plotting the species and their ecological strategies (fast or slow) in  
181 ordination space (NMDS) space, and then fitting the values of FWU parameters: leaf  
182 total water absorbed ( $C_{\max}$ ) and leaf water absorption speed ( $k$ ), via generalized additive  
183 models (GAM). The NMDS and GAM analyses were conducted in the statistical  
184 package *vegan* (Oksanen *et al.*, 2016), in the R Statistical Environment (R Development  
185 Core Team, 2019). We further determined whether the chemic compound classes were  
186 related to FWU parameters ( $C_{\max}$  and  $k$ ) via generalised linear models (using Gaussian  
187 error distributions) with the compound class as the response variables and the species as  
188 explanatory variable.

189

## 190 **RESULTS**

191

### 192 *Chemical composition of leaf cuticular waxes*

193

194 The chemical composition of the cuticular waxes influences, qualitatively and  
195 quantitatively, the FWU of the six studied species. All species presented primary  
196 alkanols, *n*-alkanes, alkanolic acid, steroids and triterpenoids. Alkyl esters was present in  
197 all species except *P. heteromallum*. The simplest cuticular wax with only 4 aliphatic  
198 compound classes was found in *P. heteromallum* (Table 1). *Leandra australis* and *T.*  
199 *laniflora* had most complex cuticular wax with 7 compound classes (Table 1). Alfa-  
200 amyirin was detected in five of the six species studied (Supplementary data).

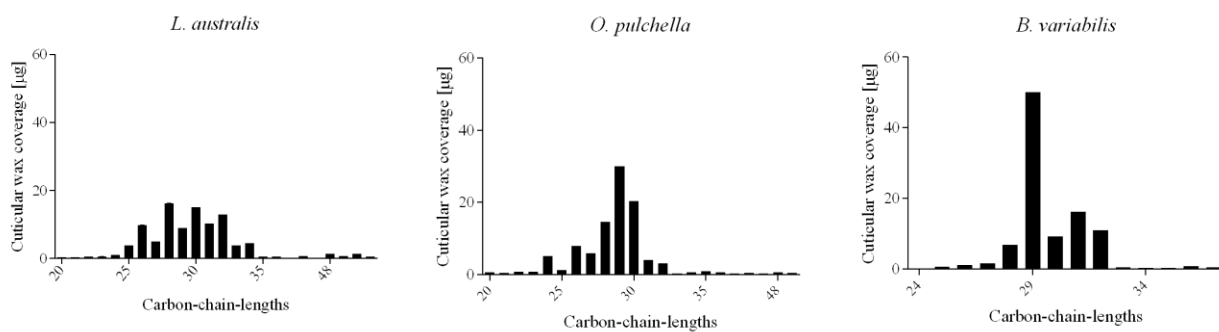
201 Among the species with fast FWU, the most abundant compound classes in *L.*  
202 *australis* and *O. pulchella* was the primary alkanols ( $0.418 \pm 0.1 \mu\text{g cm}^{-2}$ , 29.6%, and  
203  $0.827 \pm 0.111 \mu\text{g cm}^{-2}$ , 26.2%, respectively), and the steroids and triterpenoids in *B.*  
204 *variabilis* ( $9.094 \pm 1.66 \mu\text{g cm}^{-2}$ , 35.9%). For the species with the lowest absorption  
205 speed strategy, *T. laniflora* ( $1.777 \pm 0.186 \mu\text{g cm}^{-2}$ , 37.8%) and *P. heteromallum* ( $2.703$   
206  $\pm 0.668 \mu\text{g cm}^{-2}$ , 50.7%), the n-alkanes as the main class of compounds. Although the  
207 n-alkanes was the second largest class detected in *S. reniformis* ( $1.005 \pm 0.165 \mu\text{g cm}^{-2}$ ,  
208 15.4%), the steroids and triterpenoids were the most abundant classes of compounds  
209 ( $3.144 \pm 0.534 \mu\text{g cm}^{-2}$ , 47.8%), demonstrating a chemical composition similar to those  
210 of the fast water absorption species. The species with the highest amount of cuticular  
211 wax was *B. variabilis* ( $25.320 \pm 7.50 \mu\text{g cm}^{-2}$ ). The other species showed wax amounts  
212 between 1.41 to  $6.51 \mu\text{g cm}^{-2}$  (Table 1).

213 The very long chain aliphatic compounds with around 20 to 34 carbons varied among  
214 species. *B. variabilis* and *O. pulchella* but the highest percentage were molecules with  
215 29 C ( $6.207 \pm 2.82 \mu\text{g cm}^{-2}$ , 50%) and 29 C ( $0.770 \pm 0.14 \mu\text{g cm}^{-2}$ , 30%), respectively.  
216 *Leandra australis* was the exception, with molecules up to 36 carbons (Fig. 1) but the  
217 highest percentage were molecules with 28 C ( $0.125 \pm 0.06 \mu\text{g cm}^{-2}$ , 16%). *P.*  
218 *heteromallum*, *S. reniformis* and *T. laniflora* presented chains around 20 to 37 carbons, in  
219 which the highest percentage was represented by molecules with 33 C ( $1.103 \pm 0.395$   
220  $\mu\text{g cm}^{-2}$ , 34%), 29 C ( $0.606 \pm 0.15 \mu\text{g cm}^{-2}$ , 29%) and 26 C ( $0.541 \pm 0.16 \mu\text{g cm}^{-2}$ ,  
221 14%), respectively (Fig. 2). The amount of CW and the amount of each compound of  
222 the CW analyzed separately were not related to the different FWU strategies ( $C_{\text{max}}$  and  
223  $k$ ). For all major classes of compounds no species separation occurred according to the  
224 FWU strategies (Fig. 3).

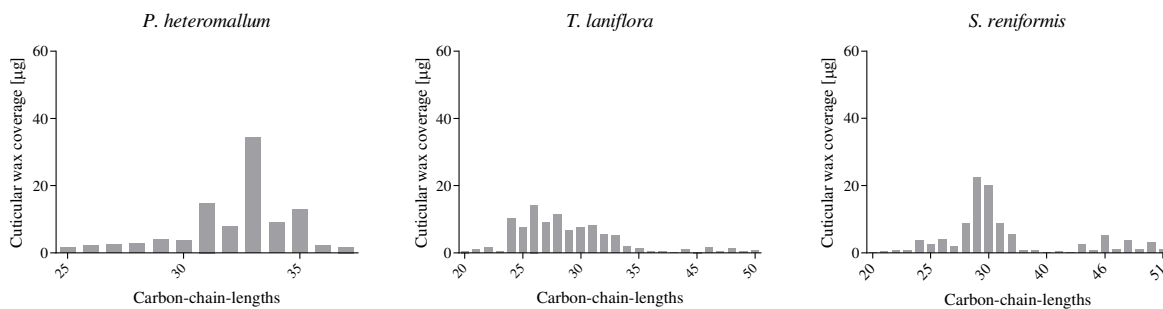
**Table 1.** Simplified chemical composition of leaf cuticular waxes of six species with different capacities of foliar water uptake. Each value represents the mean value  $\pm$  SD (n = 3).

Compound class	Coverage ( $\mu\text{g cm}^{-2}$ )					
	<i>L.australis</i>	<i>O. pulchella</i>	<i>B. variabilis</i>	<i>P. heteromallum</i>	<i>T. laniflora</i>	<i>S. reniformis</i>
primary alkanols	0.41 $\pm$ 0.10	0.82 $\pm$ 0.11	2.492 $\pm$ 0.765	0.321 $\pm$ 0.092	0.970 $\pm$ 0.605	0.798 $\pm$ 0.074
branched alkanols	0.04 $\pm$ 0.01					
n-alkanals	0.02 $\pm$ 0.00	0.17 $\pm$ 0.00			0.023 $\pm$ 0.014	0.130 $\pm$ 0.030
n-alkanes	0.24 $\pm$ 0.07	0.72 $\pm$ 0.20	8.584 $\pm$ 3.601	2.703 $\pm$ 0.668	1.777 $\pm$ 0.186	1.005 $\pm$ 0.165
branched alkanes					0.412 $\pm$ 0.130	
alkanol acetates						
alkyl esters	0.04 $\pm$ 0.01	0.110 $\pm$ 0.028	0.239 $\pm$ 0.064		0.243 $\pm$ 0.158	0.512 $\pm$ 0.008
alkanoic acids	0.02 $\pm$ 0.00	0.716 $\pm$ 0.075	1.093 $\pm$ 0.588	0.183 $\pm$ 0.052	0.368 $\pm$ 0.207	0.240 $\pm$ 0.036
flavonoids						
steroids and triterpenoids	0.38 $\pm$ 0.06	0.078 $\pm$ 0.038	9.094 $\pm$ 1.667	1.800 $\pm$ 0.865	0.436 $\pm$ 0.294	3.113 $\pm$ 0.534
not identified	0.25 $\pm$ 0.00	0.506 $\pm$ 0.023	3.816 $\pm$ 0.913	0.314 $\pm$ 0.091	0.462 $\pm$ 0.266	0.713 $\pm$ 0.160
Cuticular wax quantity total *	1.41 $\pm$ 0.21	3.14 $\pm$ 0.30	25.32 $\pm$ 7.50	5.32 $\pm$ 1.64	4.69 $\pm$ 1.75	6.51 $\pm$ 0.96

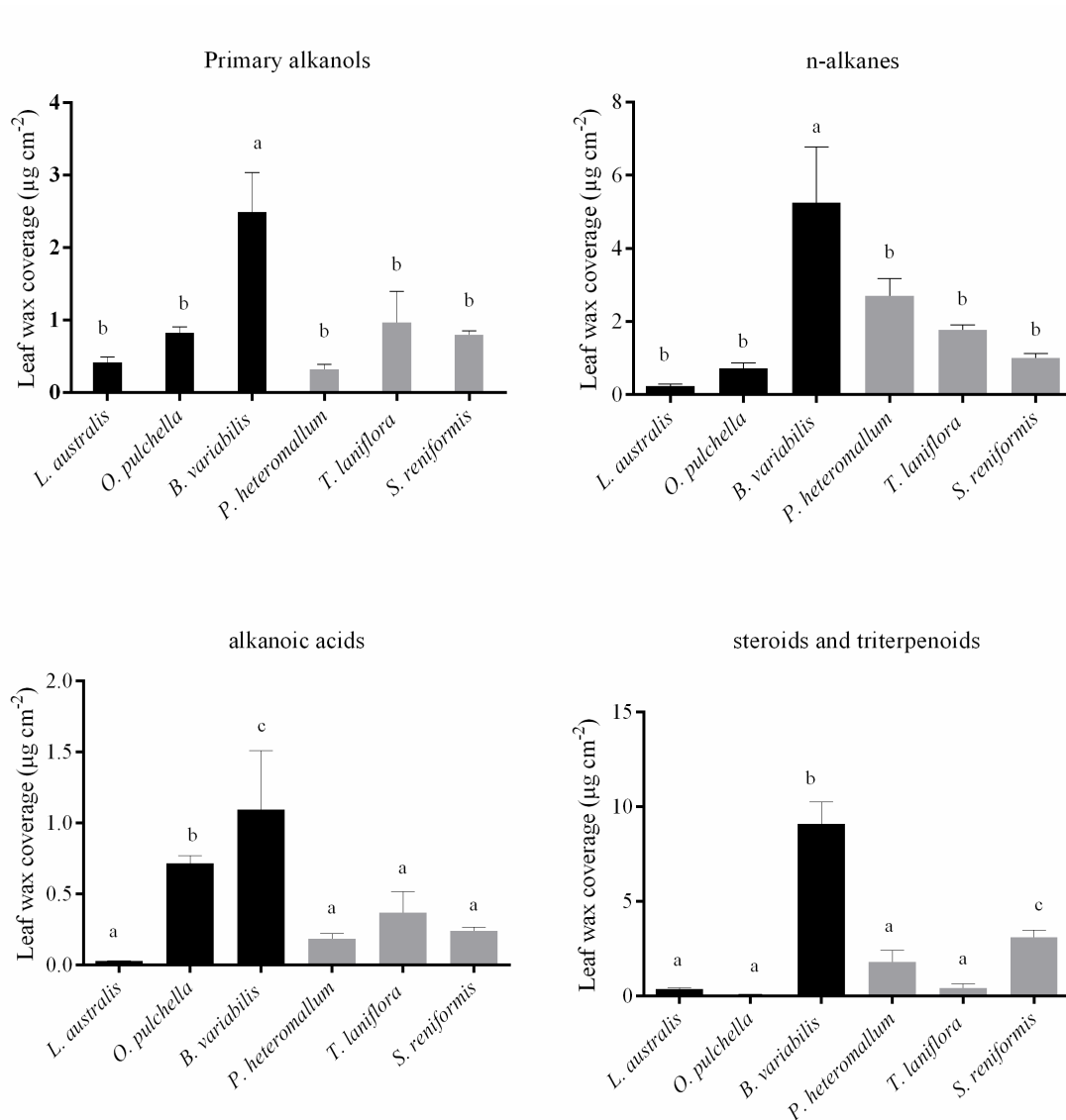
(\*) means the total amount of cuticular wax also including the other non-aliphatic compounds described in this table.



**Fig. 1.** Chain-length distributions of the very-long-chain aliphatic fraction of cuticular waxes of species with speed water absorption. Bars stand for the mole-based contribution of a single chain-length to the total very-long-chain aliphatic wax coverage.



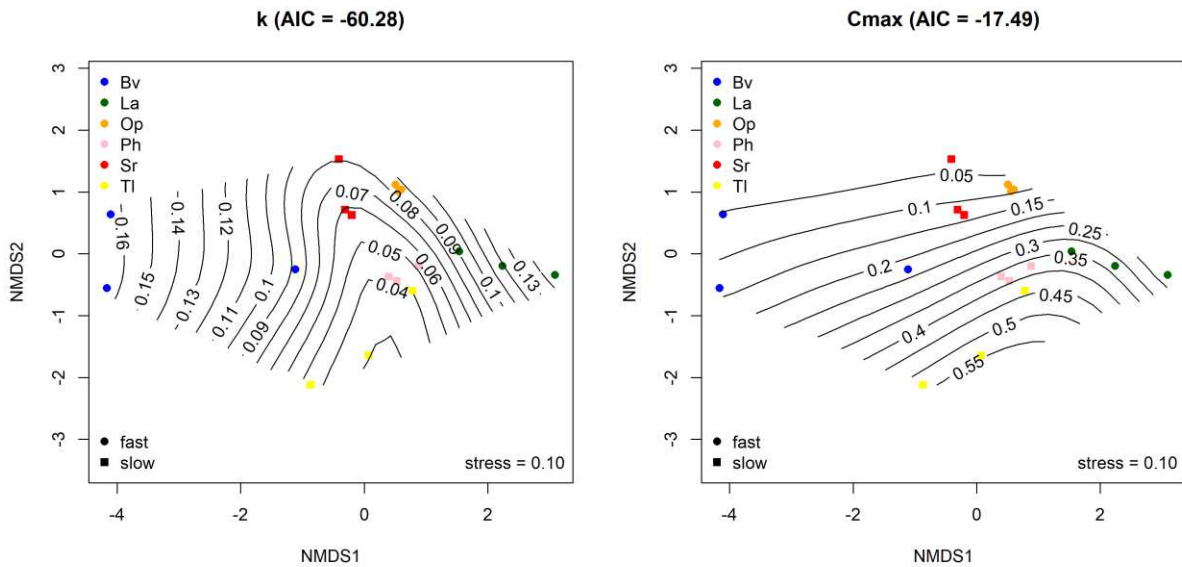
**Fig. 2.** Chain-length distributions of the very-long-chain aliphatic fraction of cuticular waxes of species with slow water absorption. Bars stand for the mole-based contribution of a single chain-length to the total very-long-chain aliphatic wax coverage.



**Fig. 3.** Cuticular wax coverage of principal's compound class of six species leaves with different speed of FWU (*k*). Each value represents the mean value  $\pm$  SD. Black and grey bars indicate species with higher *k* and lower *k*, respectively. Different letters indicate differences between species ( $P \leq 0.05$ ). Error bars represent  $\pm$  SE.

225 The first two NMDS axes (there was a negligible decrease in stress values by adding  
 226 a third axis) were effective in segregating the two ecological strategies. Only species  
 227 with a slow strategy (*T. laniflora*, *P. heteromallum* and *S. reniformis*) are placed in both  
 228 negative values of the first NMDS axis and positive values of the second NMDS axis

229 (Fig. 4). We also found that the speed of water uptake (parameter  $k$ ) represented a better  
 230 fit to the relationship between species chemical composition and their water uptake  
 231 parameter (AIC = -60.28) than the amount of water uptake (parameter  $C_{\max}$ ; AIC = -  
 232 17.49) (Fig. 4).  
 233

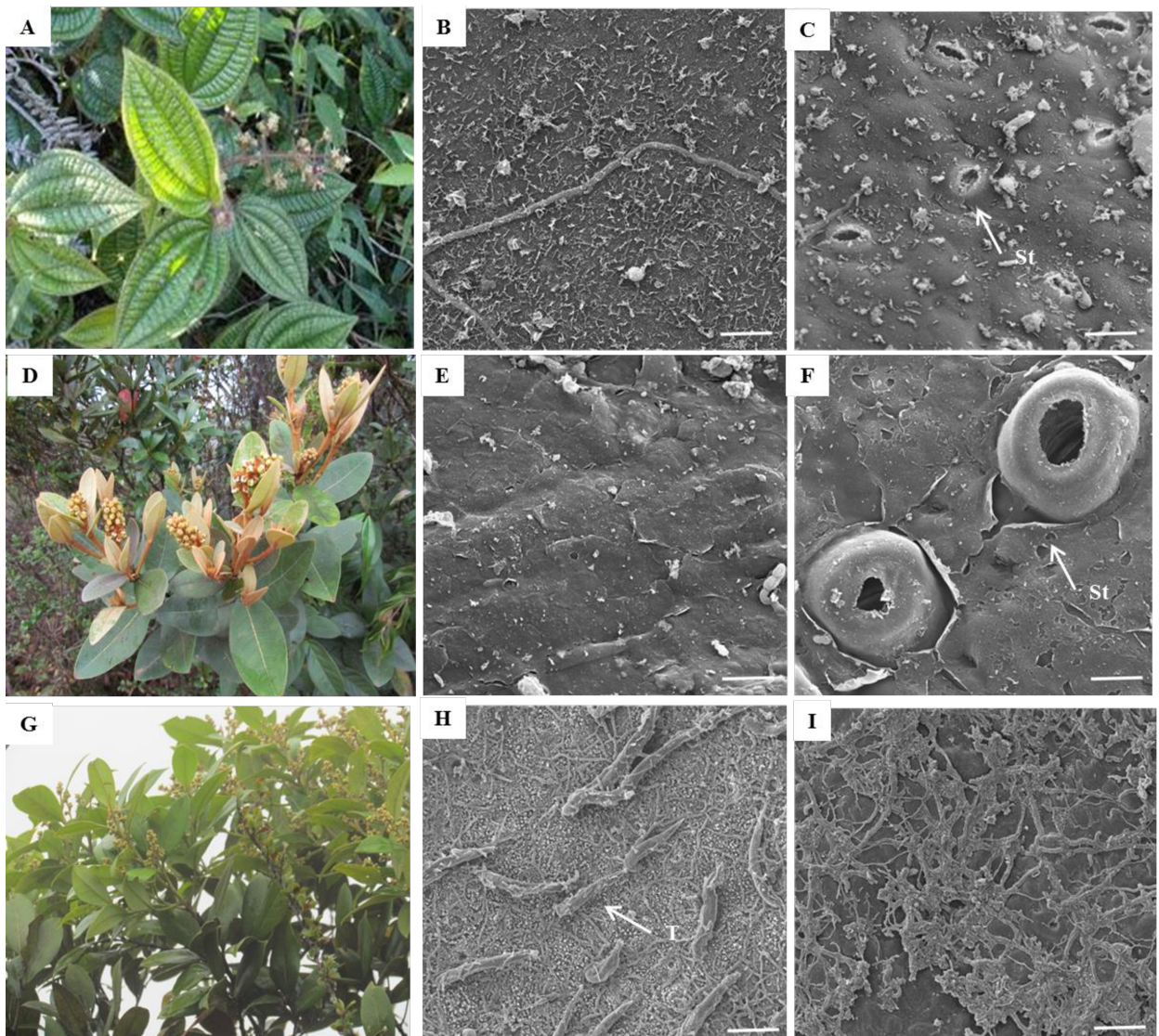


234  
 235  
 236  
 237 **Fig. 4.** NMDS plot of cuticular waxes composition of six species (a) maximum leaf water  
 238 absorption ( $C_{\max}$ ) between species and (b) leaf water uptake speed ( $k$ ) among species. BV: *B.*  
 239 *variabilis*, La: *L. australis*, Op: *O. pulchella*, Ph: *P. heteromallum*, Sr: *S. reniformis* and Tl: *T.*  
 240 *laniflora*.

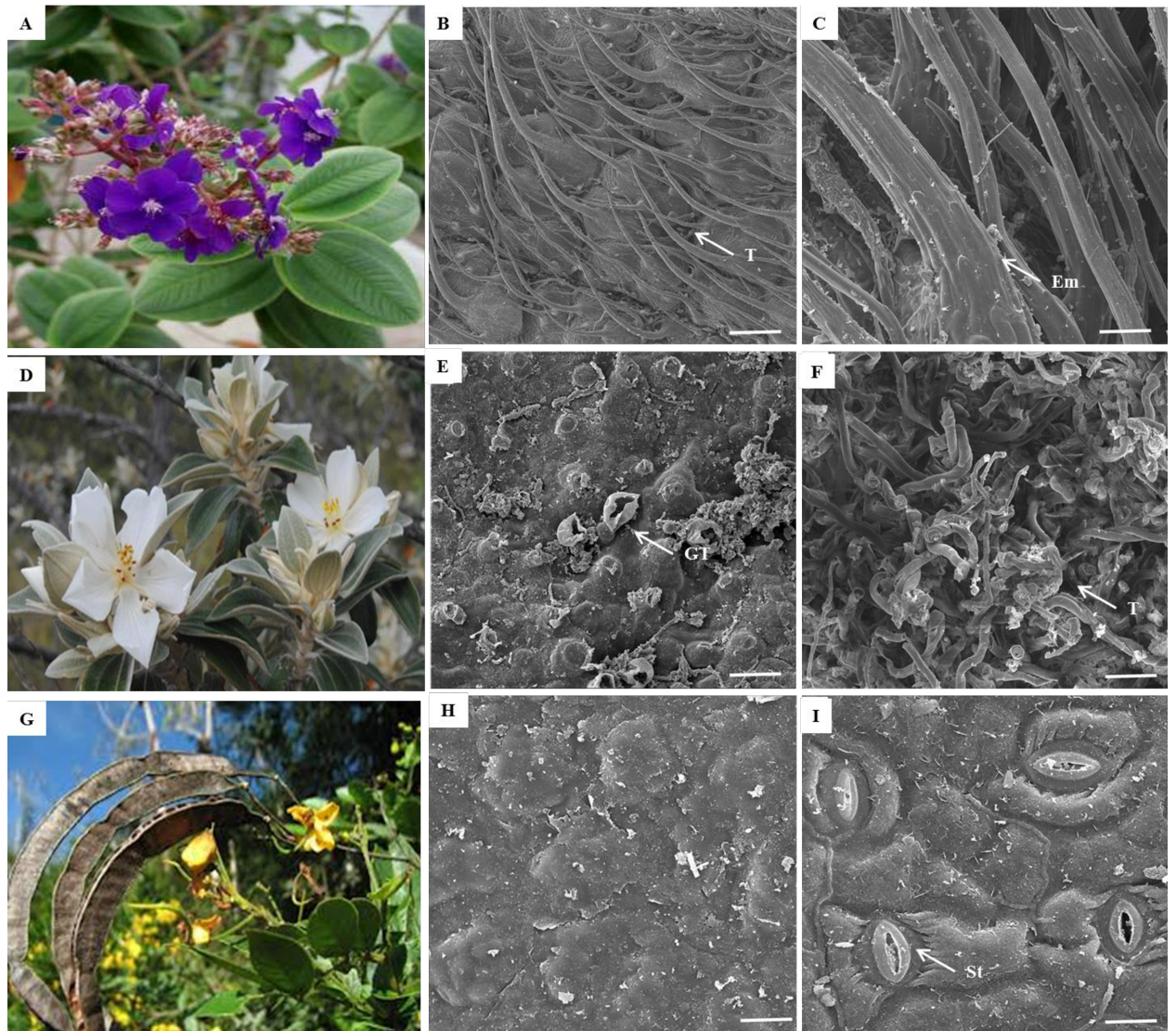
241  
 242 *Scanning electron microscopy analyses*

243  
 244 The type of the epicuticular waxes (EW) of *B. variabilis* and *O. pulchella* is of crust  
 245 with granules on their adaxial leaf surfaces (Fig. 5). *Ocotea pulchella* had the similar  
 246 type but also presented sparse trichomes. On the abaxial face, *B. variabilis* presented the  
 247 EW in crust with ribbons and *O. pulchella* presented the EW in film. In *L. australis*,

248 both on the adaxial and abaxial leaf surfaces, the EW in crust with plateles was found.  
 249 *Senna reniformis*, both on the adaxial and abaxial leaf surfaces, presented crust-like  
 250 EW. In *T. laniflora* and *P. heteromallum*, the EW in crust with granules and threads was  
 251 found on the adaxial leaf surface, and just EW in crust on the abaxial surface. The  
 252 difference between *T. laniflora* was the presence of glandular trichomes and *P.*  
 253 *heteromallum* was the presence of emergences on the adaxial surface (Fig. 5).



**Fig. 5.** Scanning electron micrographs of the adaxial and abaxial surfaces of species with speed water absorption. (A-B) *Leandra australis*, adaxial and abaxial surfaces respectively; (C-D) *Byrsonima variabilis*, adaxial and abaxial surfaces respectively; (E-F) *Ocotea pulchella*, adaxial and abaxial surfaces respectively. St: stomatal; T: Trichomes. Bars = 10  $\mu$ m.



**Fig. 6.** Scanning electron micrographs of the adaxial and abaxial surfaces of species with slow water absorption. (A-B) *Pleroma heteromallum*, adaxial and abaxial surfaces respectively; (C-D) *Trembleya laniflora*, adaxial and abaxial surfaces respectively and (E-F) *Senna reniformis*, adaxial and abaxial surfaces respectively. St: stomatal; T: Trichomes; GT: glandular trichomes; St: stomatal. Bars = 10  $\mu$ m

254 **DISCUSSION**

255

256 The ordinary role of the cuticle is to prevent excessive water loss from transpiration

257 (Riederer and Schreiber, 2001; Burghardt and Riederer, 2006). Accordingly, several

258 ecological and physiological roles have been attributed to the epicuticular wax, except  
259 leaf hydration from FWU. Contrary to the common sense, our hypothesis that different  
260 FWU capacities, especially regarding the absorption speed, are related to the cuticular  
261 wax composition of the six species of the rupestrian field was corroborated. The  
262 absorption speed ( $k$ ) was much higher than the total amount of water absorbed ( $C_{\max}$ ),  
263 which is related to leaf surface features capable of controlling the speed of water entry.  
264 The high  $C_{\max}$  is better related to leaf anatomical traits, such as tissue thickness, and cell  
265 wall components such as pectin and cellulose (Boanares et al., 2018a, b).

266 The chemical features of some classes of compounds detected in cuticle wax are  
267 already known and may interfere in FWU. The *n*-alkanes, total nonpolar compounds,  
268 has no hydroxyl radical, and tend to be associated with a low permeability of the CW  
269 (Yeast and Rose, 2013). The *n*-alkanes confer compactness to the structure of CW,  
270 negatively interfering with FWU, consequently the large amount of *n*-alkanes found in  
271 some plant species relates to their low water absorption speed ( $k$ ). Contrastingly, fast-  
272 absorbing species presented a higher proportion of classes of compounds, such as  
273 alkanols, steroids, and triterpenoids, which have hydroxyl or oxygen radicals, and some  
274 degree of polarity, H-bond interaction, and a higher solubility parameter in relation to *n*-  
275 alkanes (Fernandez et al., 2016). Alkanols tend to decrease the nonpolar part, as  
276 hydroxyls can bind to water molecules, facilitating their entry into leaf tissues.  
277 Triterpenoids and steroids disrupt *n*-alkane compaction, disaggregate the crystalline  
278 cuticle layer, promoting polar spaces, aiding water movement (Reynhardt, 1997;  
279 Riederer and Schreiber, 2001), and thereby favoring FWU. This is consistent with a  
280 model in which CW are located in crystalline or amorphous domains of the cuticle, with  
281 aliphatic compounds forming the crystalline domain that is impermeable to water. The  
282 crystalline structure forces water to diffuse through a tortuous route through the

283 amorphous domain that is formed by polar and cyclical compounds (Riederer and  
284 Schreiber, 1995), facilitating the FWU.

285 Triterpenoids such as *b*-amyirin also increase the volume of amorphous cuticle  
286 molecules, and decrease the size of the crystalline domain, blocking aggregation or  
287 increasing the distance between the nonpolar parts (Buschhaus and Jetter, 2012). Thus,  
288 the more polar compounds present in CW, the more routes through which water can  
289 diffuse may be opened or shortened, thereby decreasing the overall cuticle resistance. A  
290 common perception is that a thick cuticle is associated with lower water permeability  
291 and therefore low FWU capacity. However, the quantity/thickness of CW was unrelated  
292 to permeability or to FWU (Riederer and Schreiber 2001; Schuster, 2016; Bueno 2018).  
293 In fact, the proportion of *n*-alkanes is more significant to CW permeability than the total  
294 amount of waxes (Yeast and Rose, 2013), corroborating our data with plants and  
295 environmental condition of *Campo Rupestre*. The chemical complexity of CW was also  
296 unrelated to FWU possibly because compound classes may be equivalent in structure  
297 and function, and may not individually influence permeability. Commonly, the chain  
298 size of aliphatic compounds has been related to the water barrier (Vogg et al., 2004;  
299 Leide et al., 2007, 2011; Buschhaus and Jetter, 2012; Jetter and Riederer, 2016; Jetter  
300 and Riederer, 2016). Very long-chain aliphatic compounds build impermeable  
301 crystalline domains (Riederer and Schreiber, 1995) and constitute a barrier to water  
302 outflow and possibly to water entrance into plants, as well. Therefore, the fact that fast  
303 FWU species presented low percentages of very long-chain aliphatic compounds  
304 compared to water slow absorption species corroborates this premise.

305 Except for the *n*-alkanes, the other classes of compounds detected in the CW such as  
306 acids and alcohols, have some degree of polarity and H-bond interactivity. Thus, the  
307 polarity or potential H-binding properties of each compound found on plant surfaces are

308 more important than their hydrophobic or lipophilic traits (Fernandez et al., 2016),  
309 because one molecule alone may be irrelevant but their association with other classes of  
310 compounds can produce more polar or nonpolar sites, which may, in fact, interfere with  
311 water entrance. After all, the predominant understanding of plant cuticle as independent  
312 and lipidic is largely based on Brongniart (1830) and von Mohl's (1847) hypotheses,  
313 which were already controversial at the time. Possibly the non-responsiveness of the  
314 elements separately regards to the importance of the association of cuticle compounds to  
315 the FWU. Contrastingly, it seems that CW permeability is mainly determined by the  
316 particular mixture of epicuticular and intracuticular waxes and their morphology and  
317 organization within the cuticle architecture, and not just by an isolated class of  
318 compounds (Yeast and Rose, 2013). The overall profile of the CW mixture plays  
319 important roles in cuticle structure models because the functional groups of the wax  
320 constituents engage in spontaneous reactions within the cuticle, and confer gradual  
321 modifications to the CW composition (Riederer and Schreiber, 2001). Thus, structure  
322 and compositional variability are of particular importance for cuticular permeability to  
323 water (Schreiber, 2005). Since also in our data we find that in addition to the proportion  
324 of each compound, a mixture of each class of compound interferes with CW  
325 permeability.

326 While intracuticular waxes function primarily as a barrier to excessive water loss  
327 through transpiration (Riederer and Schreiber, 1995), epicuticular waxes (EW) strongly  
328 influence surface wettability (Bargel et al., 2006). Scanning electron microscopy  
329 revealed several ornamentations of the EW. The most complex or more ornate EW was  
330 observed on the species with the lowest FWU speed (*P. heteromallum* and *T.laniflora*,  
331 except *S. reniformis*). These data are in accordance with the fact that three-dimensional  
332 EW crystals create microstructured surfaces with low wettability (Baker, 1982; Wagner

333 et al., 2003). Therefore, a higher density of three-dimensional wax crystals provides a  
334 higher degree of leaf surface hydrophobicity (Koch, 2006). Although the presence of  
335 trichomes has a positive influence on FWU, it is important to emphasize that this  
336 influence is more related to the amount of water absorbed than to its speed (Boanares et  
337 al., 2018a). Contrastingly, the species with the highest water leaf absorption speed were  
338 the species with a simpler EW, and less ornamentation, which could have facilitated  
339 water entrance.

340 For some waxes, a distinct coherence between chemical composition and morphology  
341 in EW has been found (Koch and Ensikat, 2008). Platelets are a sub-type of plate wax  
342 observed on the adaxial face of *L. australis*, which has an intimate relationship with the  
343 EW chemical composition. It is well known that some platelets are formed by primary  
344 alcohols (Koch et al., 2006), which were the main components of the CW composition  
345 of *L. australis*. However, for some types of wax, the correlation between crystals,  
346 morphology, and chemistry is still unclear (Koch and Ensikat, 2008), which  
347 corresponds to the non-correlation between chemistry and cuticle ornamentation of the  
348 studied species.

349 For the six species of the Brazilian rupestrian fields, the different CW compositions  
350 and EW ornamentations are related to the different FWU capacities. The combination of  
351 all classes of compounds and not simply the isolated amount of one or another class of  
352 compound provides the three-dimensional structure that favors the different degrees  
353 water entrance into leaf tissues, interfering in the FWU.

354

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**Table 1.** Supplementary data. Total chemical composition of leaf cuticular waxes of six species with different FWU capacities. Each value represents the mean value  $\pm$  SD (n = 3).

Compound class	Carbon-chain-length	Coverage ( $\mu\text{g cm}^{-2}$ )					
		<i>L.australis</i>	<i>O.pulchella</i>	<i>B.variabilis</i>	<i>P.heteromallum</i>	<i>T.laniflora</i>	<i>S.reniformis</i>
<i>n</i> -alkanes	25	0.023 $\pm$ 0.008	0.016 $\pm$ 0.003	0.071 $\pm$ 0.049	0.051 $\pm$ 0.024	0.212 $\pm$ 0.04	0.018 $\pm$ 0.007
	26	0.024 $\pm$ 0.009	0.03 $\pm$ 0.013	0.077 $\pm$ 0.053	0.06 $\pm$ 0.028	0.229 $\pm$ 0.053	0.019 $\pm$ 0.009
	27	0.03 $\pm$ 0.01	0.112 $\pm$ 0.017	0.134 $\pm$ 0.028	0.058 $\pm$ 0.027	0.266 $\pm$ 0.032	0.027 $\pm$ 0.01
	28	0.035 $\pm$ 0.014	0.082 $\pm$ 0.057	0.227 $\pm$ 0.05	0.061 $\pm$ 0.017	0.181 $\pm$ 0.048	0.064 $\pm$ 0.015
	29	0.047 $\pm$ 0.014	0.379 $\pm$ 0.104	6.154 $\pm$ 2.81	0.112 $\pm$ 0.014	0.190 $\pm$ 0.019	0.606 $\pm$ 0.151
	30	0.02 $\pm$ 0.005	0.023 $\pm$ 0.007	0.157 $\pm$ 0.03	0.063 $\pm$ 0.013	0.116 $\pm$ 0.028	0.068 $\pm$ 0.006
	31	0.038 $\pm$ 0.011	0.083 $\pm$ 0.019	1.762 $\pm$ 0.88	0.428 $\pm$ 0.148	0.256 $\pm$ 0.103	0.201 $\pm$ 0.027
	32	0.012 $\pm$ 0.003			0.162 $\pm$ 0.052	0.083 $\pm$ 0.006	
	33	0.013 $\pm$ 0.001			1.054 $\pm$ 0.387	0.163 $\pm$ 0.086	
	34				0.165 $\pm$ 0.051	0.026 $\pm$ 0.002	
	35				0.397 $\pm$ 0.163	0.035 $\pm$ 0.011	
	36				0.03 $\pm$ 0.01	0.013 $\pm$ 0.002	
	37				0.056 $\pm$ 0.018		
<i>Total n-alkanes</i>		0.246 $\pm$ 0.078	0.728 $\pm$ 0.223	8.584 $\pm$ 3.90	2.703 $\pm$ 0.957	1.777 $\pm$ 0.436	1.005 $\pm$ 0.228
branched-alkanes	25					0.012 $\pm$ 0.005	
	26					0.029 $\pm$ 0.014	
	27					0.045 $\pm$ 0.020	
	28					0.017 $\pm$ 0.002	
	29					0.035 $\pm$ 0.004	
	30					0.036 $\pm$ 0.004	
	31					0.026 $\pm$ 0.006	
32					0.031 $\pm$ 0.010		

	33					0.027 ± 0.016	
	34					0.026 ± 0.011	
	35					0.019 ± 0.007	
<hr/>							
<i>Total branched alkanes</i>							
	21	0.001 ± 0.00	0.008 ± 0.001			0.014 ± 0.014	0.004 ± 0.002
	22	0.0009 ± 0.00	0.009 ± 0.00			0.043 ± 0.022	0.007 ± 0.003
	23	0.002 ± 0.00	0.01 ± 0.001			0.009 ± 0.004	0.008 ± 0.005
	24	0.004 ± 0.001	0.077 ± 0.017	0.018 ± 0.001		0.319 ± 0.208	0.072 ± 0.011
	25	0.004 ± 0.00	0.005 ± 0.00			0.025 ± 0.011	0.03 ± 0.013
	26	0.043 ± 0.034	0.118 ± 0.019	0.0248 ± 0.004		0.184 ± 0.115	0.049 ± 0.002
	27	0.008 ± 0.004	0.016 ± 0.002	0.029 ± 0.004	0.013 ± 0.003	0.012 ± 0.005	0.025 ± 0.002
alcohol	28	0.081 ± 0.045	0.104 ± 0.026	0.092 ± 0.028	0.01 ± 0.004	0.132 ± 0.081	0.167 ± 0.038
	29	0.012 ± 0.002	0.357 ± 0.044			0.017 ± 0.007	0.274 ± 0.098
	30	0.09 ± 0.015	0.093 ± 0.034	0.797 ± 0.268	0.032 ± 0.007	0.094 ± 0.058	0.123 ± 0.024
	31	0.0175 ± 0.002		0.171 ± 0.063	0.029 ± 0.01	0.016 ± 0.01	
	32	0.087 ± 0.015	0.018 ± 0.004	1.238 ± 0.379	0.069 ± 0.022	0.068 ± 0.045	0.016 ± 0.003
	33	0.016 ± 0.003	0.006 ± 0.00	0.076 ± 0.021	0.034 ± 0.01	0.009 ± 0.005	0.019 ± 0.004
	34	0.034 ± 0.003		0.043 ± 0.014	0.085 ± 0.026	0.022 ± 0.014	
	35	0.004 ± 0.00			0.021 ± 0.005		
	36	0.004 ± 0.00			0.025 ± 0.006		
<hr/>							
<i>Total primary alkanols</i>							
	29	0.01 ± 0.002					
branched-alcohol	30	0.007 ± 0.001					
	31	0.024 ± 0.13					
<hr/>							
	24					0.013 ± 0.01	
alkanals	26	0.002 ± 0.001	0.016 ± 0.002				
	28		0.043 ± 0.002			0.01 ± 0.004	
<hr/>							

	30						0.081 ± 0.011			0.13 ± 0.03
	32						0.028 ± 0.007			
<i>Total alkanals</i>	32						0.078 ± 0.012		0.023 ± 0.014	0.13 ± 0.03
	24						0.013 ± 0.008			
Coumaric acid ester	26						0.006 ± 0.003			
	28						0.007 ± 0.002			
	30						0.019 ± 0.006			
	20	0.002 ± 0.00	0.016 ± 0.00						0.022 ± 0.014	0.006 ± 0.003
	21	0.001 ± 0.00	0.005 ± 0.00						0.024 ± 0.015	0.006 ± 0.003
	22	0.002 ± 0.00	0.008 ± 0.00						0.022 ± 0.014	0.011 ± 0.002
	23	0.001 ± 0.00	0.01 ± 0.00						0.005 ± 0.003	0.01 ± 0.002
	24	0.004 ± 0.00	0.055 ± 0.002						0.054 ± 0.035	0.024 ± 0.007
	25	0.001 ± 0.00	0.01 ± 0.001	0.02 ± 0.006					0.007 ± 0.003	0.021 ± 0.006
	26	0.005 ± 0.00	0.04 ± 0.001	0.041 ± 0.022	0.01 ± 0.003				0.053 ± 0.031	0.037 ± 0.002
	27		0.023 ± 0.002	0.035 ± 0.015	0.008 ± 0.002				0.008 ± 0.003	
Alkanoic acid	28	0.007 ± 0.00	0.146 ± 0.017	0.537 ± 0.529	0.021 ± 0.008				0.077 ± 0.039	
	29		0.033 ± 0.005	0.052 ± 0.024	0.015 ± 0.004				0.013 ± 0.005	
	30		0.312 ± 0.039	0.191 ± 0.095	0.018 ± 0.004				0.042 ± 0.022	0.061 ± 0.017
	31		0.02 ± 0.005	0.087 ± 0.046	0.019 ± 0.006				0.009 ± 0.004	0.031 ± 0.006
	32		0.032 ± 0.007	0.126 ± 0.054	0.021 ± 0.006				0.026 ± 0.014	0.029 ± 0.008
	33				0.014 ± 0.004					
	34				0.036 ± 0.011					
	35									
	36				0.016 ± 0.004					
<i>Total alkanolic acids</i>		0.027 ± 0.00	0.717 ± 0.087	1.093 ± 0.793	0.183 ± 0.056	0.368 ± 0.208				0.24 ± 0.61
Alkanol acetate	26									
	28									

<i>Total alkanol acetates</i>						
	40					0.006 ± 0.0
	41					
	42		0.015636365		0.015 ± 0.11	0.014 ± 0.003
	43				0.006 ± 0.004	0.006 ± 0.00
	44	0.001 ± 0.00	0.025856349	0.052 ± 0.025	0.037 ± 0.025	0.066 ± 0.009
alkyl ester	45				0.01 ± 0.006	0.021 ± 0.002
	46	0.005 ± 0.001	0.015 ± 0.002	0.108 ± 0.023	0.059 ± 0.40	0.137 ± 0.00
	47	0.002 ± 0.007	0.008 ± 0.001		0.014 ± 0.008	0.025 ± 0.001
	48	0.01 ± 0.003	0.012 ± 0.001	0.078 ± 0.027	0.053 ± 0.036	0.0977 ± 0.002
	49	0.005 ± 0.006	0.007 ± 0.007		0.015 ± 0.008	0.024 ± 0.001
	50	0.011 ± 0.003	0.014 ± 0.001		0.03 ± 0.018	0.084 ± 0.007
	51	0.003 ± 0.00	0.011 ± 0.00			
<i>Total alkyl ester</i>						
		0.04 ± 0.011	0.11 ± 0.03	0.239 ± 0.076	0.243 ± 0.159	0.512 ± 0.031
	alpha-amyrin	0.008 ± 0.002		1.136 ± 0.317	0.209 ± 0.090	0.269 ± 0.181
	amyrin derivative			0.363 ± 0.032	0.032 ± 0.014	0.17 ± 0.01
	beta-amyrin			1.562 ± 0.345	0.274 ± 0.139	0.100 ± 0.068
	beta-amyrinon				0.014 ± 0.004	0.248 ± 0.014
	beta-sitosterol	0.004 ± 0.00				0.248 ± 0.045
	beta-tocopherol		0.012 ± 0.006	0.064 ± 0.045		
	betulin			0.124 ± 0.036		0.092 ± 0.005
	betulinic acid					0.051 ± 0.007
	cholesterol	0.007 ± 0.00	0.017 ± 0.011	0.041 ± 0.014	0.01 ± 0.002	0.027 ± 0.003
	delta-amyrin				0.023 ± 0.004	0.023 ± 0.001
	delta-tocopherol		0.004 ± 0.001			
	erythrodiol	0.005 ± 0.00			0.011 ± 0.004	0.102 ± 0.025

fridelin						0.268 ± 0.092
gama-tocopherol						0.017 ± 0.004
hederagenin	0.006 ± 0.00					
hederagenin derivative	0.005 ± 0.001					
lupenon			0.12 ± 0.055			0.705 ± 0.284
lupeol			4.927 ± 0.806	0.954 ± 0.699	0.051 ± 0.038	0.268 ± 0.059
lupeol derivative 1			0.395 ± 0.083	0.096 ± 0.085		0.439 ± 0.206
non-identified triterpenoid	0.043 ± 0.009		0.229 ± 0.09	0.086 ± 0.014		
oleanoic acid	0.074 ± 0.015		0.123 ± 0.014	0.019 ± 0.006		
ursolic acid	0.193 ± 0.041			0.017 ± 0.009	0.015 ± 0.008	
ursolic acid derivative	0.012 ± 0.002					
uvaol	0.018 ± 0.001			0.046 ± 0.016		0.076 ± 0.014
not identified	0.255 ± 0.006	0.506 ± 0.023	3.816 ± 0.913	0.314 ± 0.091	0.461 ± 0.266	0.714 ± 0.161
<i>Cuticular wax total</i>	1.41 ± 0.21	3.14 ± 0.30	25.32 ± 7.50	5.32 ± 1.64	4.69 ± 1.75	6.51 ± 0.96

*Pectin and cellulose cell wall composition enables different strategies to leaf water uptake in plants from tropical fog mountain*

1 Pectin and cellulose cell wall composition enables different strategies  
2 to leaf water uptake in plants from tropical fog mountain

3  
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13  
14 A B S T R A C T

15 Leaf water uptake (LWU) has been observed in plants of different ecosystems and this process is distinct among  
16 different species. Four plant species from the Brazilian fog mountain fields were evaluated in order to detect if  
17 leaf water uptake capacity is related to the cell wall composition of leaf epidermis. LWU measurements and their  
18 relation to anatomical and biochemical traits were analyzed. Cell wall composition was verified through  
19 immunocytochemistry using monoclonal antibodies recognizing pectin compounds, and histochemistry with  
20 calcofluor white to track cellulose. Differences in LWU among the four species were clearly revealed. Two  
21 species presented higher maximum leaf water content and the lowest values of water absorption speed. The other  
22 two species presented opposite behavior, namely, low leaf water uptake and the highest values of water  
23 absorption speed. The anatomical traits associated with the cell wall composition corroborated the data on the  
24 different LWU strategies. The species with abundant detection of cellulose in their epidermal cell walls absorbed  
25 more water, but more slowly, while those with abundant detection of pectins absorbed water at a higher speed.  
26 These results indicate that cell wall composition regarding pectin and cellulose are significant for water uptake  
27 by the leaf epidermis. Pectin provides greater porosity and absorption speed, while cellulose provides greater

28 hydrophilicity and greater water uptake capacity. Current data indicate that the composition of epidermal cell  
29 walls is a relevant trait for leaf water uptake.

30

31 *Keywords:*

32 Immunocytochemistry

33 Leaf traits

34 Rupestrian field

35 Water relations

36

37 *Abbreviations:* CW, cell walls; LWU, leaf water uptake; GalA, galacturonic acid; HGA, homogalacturonans;  
38 RG-I, rhamnogalacturonans I; RG-II, rhamnogalacturonans II;  $C_{max}$ , maximum water content;  $k$ , leaf uptake  
39 speed; LSM, leaf specific mass; LSA, leaf specific area; SU, succulence; RWC, relative water content; WSD,  
40 water saturation deficit; GLM, general linear model

41

## 42 **1. Introduction**

43

44 Leaf water uptake (LWU) is more easily observed when leaves are in the presence of a great amount of fog,  
45 and water steam can come from the atmosphere toward leaves through a difference in water potential (Burgess  
46 and Dawson, 2004; Oliveira et al., 2005; Goldsmith et al., 2013; Eller et al., 2015; Gotsch et al., 2015). Although  
47 LWU is responsible for a small portion of plant water content in relation to the amount that enters by roots, such  
48 absorption increases leaf hydration and water potential without a visible increase in soil moisture (Ewing et al.,  
49 2009; Simonin et al., 2009; Limm et al., 2009). Thus, LWU provides an ecological advantage for plants  
50 occurring at sites with water deficits, at least seasonally, playing an important role in plant survival and growth  
51 (Limm et al., 2009; Eller et al., 2013, 2016). Therefore, LWU capacity confers advantages to the vegetation by  
52 reducing the negative impacts of severe drought (Burgess and Dawson, 2004; Eller et al., 2013). Physiological  
53 studies have shown that LWU helps plants to better face drought events (Eller et al., 2016); and that it can be  
54 facilitated by leaf structures such as trichomes, stomata, emergences, hydathodes, but can also occur through the  
55 ordinary epidermal cells (Martin and von Willert, 2000; Riederer and Schreiber, 2001; Burkhardt et al., 2012;  
56 Fernandez et al., 2014).

57 Leaf water uptake may occur in many plant species of different ecosystems (Martin and von Willert, 2000;  
1 58 Gouvra and Grammatikopoulos, 2003; Oliveira et al., 2005; Breshears et al., 2008; Limm et al., 2009; Eller et  
2 3 al., 2013; Goldsmith et al., 2013; Cassana et al., 2015; Gotsch et al., 2016; Eller et al., 2016) by several possible  
4 59 mechanisms. Nevertheless, no study has investigated whether the LWU capacity is related to the presence of  
5 60 some specific structural traits, such as special cell types or trichomes, or even the composition of epidermal cell  
6 61 walls.  
7 62

11 63 The cell walls (CW) are one of the main barriers to the entrance of water into leaf cells. In plants, the CW are  
12 64 mainly composed of cellulose (Carpita and McCann, 2000), hemicelluloses and pectins (Wolf and Greiner,  
13 65 2012). Cellulose is a linear homopolymer consisting of long chains of D-glucose units, linked through  $\beta$ -type  
14 66 glycosidic bonds (1  $\rightarrow$  4). Each cellulose molecule is linked to the others by means of hydrogen bonds, forming  
15 67 sets identified as microfibrils (Zykwinska et al., 2005; Agoda-Tandjawa et al., 2012). The cellulose microfibrils  
16 68 are dispersed into water by shearing forces, when they may form gels, but when the microfibrils interact with  
17 69 each other, they may structure a strong network (Tatsumi et al., 2002; Pääkkö et al., 2007).

26 70 Pectins are a group of diversified polysaccharides rich in galacturonic acid (GalA) (Caffall and Mohnen,  
27 71 2009). They are divided into distinct groups depending on the types of monosaccharides associated with GalA  
28 72 and the types of linkage between its monomers. Among the main groups of pectins, there are homogalacturonans  
29 73 (HGA), rhamnogalacturonans I (RG-I), and rhamnogalacturonans II (RG-II) (Ridley et al., 2001; Albersheim et  
30 74 al., 2011). The HGAs are homopolymers of GalA bound through  $\alpha$ -(1  $\rightarrow$  4) bonds (Willats et al., 2001;  
31 75 Derbyshire et al., 2007; Verhertbruggen et al., 2009; Oliveira et al., 2014), they are very abundant in the CW,  
32 76 generally comprising 60-65% of the pectin set (Bush et al., 2001). The RG-I, contrastingly, are branched  
33 77 polymers of GalA, generally representing 20-35% of the pectin matrix (Oliveira et al., 2014). They have many  
34 78 structurally different side chains attached through the C-4 of the central skeletal rhamnosil with arabinosyl and  
35 79 galactosyl residues as the predominant chain components. The RG-I can alter the degrees of the CW porosity,  
36 80 flexibility and cell-cell adhesion (Mohnen et al., 2008; Verhertbruggen et al., 2009; Oliveira et al., 2014).

48 81 Pectins can regulate cell-cell signalling, cell growth, morphogenesis and defence (Agoda-Tandjawa et al.,  
49 82 2012). Moreover, pectins may allow increased water storage and transit in the CW, and regulate the stiffness  
50 83 levels, according to the degrees of methyl esterification and chain branching (Willats et al., 2001; Alba and  
51 84 Kontogiorgos, 2016). The degree of methylation (DM) of the HGA molecules is the proportion of methyl  
52 85 esterified galacturonic acids in their chain. The HGAs are synthesized in a highly methyl esterified form, and the  
53 86 structure of the polymers may undergo varying de-methylesterification degrees throughout cell development  
54 55 56 57 58 59 60 61 62 63 64 65

87 (Wolf et al., 2009). HGAs with low degree of methyl esterification, when hydrated, form gels in association with  
1 88 calcium ions at different levels of wall acidity (Axelos and Thibault, 1991).

2 89 It is speculated that the composition of the epidermal CW could influence the LWU of certain plant species,  
3  
4 90 especially those naturally involved in a great amount of fog and water steam. As models of study, four native  
5  
6 91 plant species from Brazilian tropical fog mountain fields were evaluated, using histochemical,  
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8 92 immunocytochemical, and physiological approaches, to detect whether the leaf water uptake capacity was related  
9  
10 93 to the leaf surface traits including the chemical composition of the epidermal CW.  
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14 94

## 15 95 **2. Materials and methods**

16 96

### 17 97 *2.1. Plant material and leaf water uptake*

18 98

19  
20 99 Leaves of *Heteropterys campestris* (Malpighiaceae), *Tibouchina heteromalla* (Melastomataceae), *Trembleya*  
21  
22 100 *laniflora* (Melastomataceae) and *Ocotea pulchella* (Lauraceae) were collected in ferruginous outcrops above  
23  
24 101 1,450 m of altitude at Serra da Brígida, Parque Cachoeira das Andorinhas, Minas Gerais state, Brazil, where fog  
25  
26 102 is a very frequent meteorological event during the four seasons of the year.  
27  
28

29  
30 103 To evaluate the water intake ability, we performed the following procedure: first the leaves (n = 4) were  
31  
32 104 collected in the field and immediately weighed with a portable scale. Afterwards the leaves were taken to the  
33  
34 105 laboratory and weighed once more. Subsequently, leaves were immersed in distilled water and weighed every 15  
35  
36 106 min for 2h, then every 30 min for 2 h. Finally, the leaves were immersed in water again for 1 h and weighed to  
37  
38 107 obtain the maximum water absorption curve. Before each weighing, the leaves were first dried with ultra-  
39  
40 108 absorbent microfiber synthetic towels to remove the surface water (Liang et al., 2009). Leaf water uptake was  
41  
42 109 calculated by a differential equation, which provides data about the maximum water content in leaf lamina ( $C_{max}$ )  
43  
44 110 and leaf uptake speed ( $k$ ) (Liang et al., 2009).  
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### 49 112 *2.2 Leaf functional traits*

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52 114 Leaves (n = 5) from 5 individuals of each species were collected in the field to obtain the leaf functional  
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54 115 attributes. Ten leaf discs of 1 cm diameter were removed from each leaf to obtain the leaf specific mass (LSM),  
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116 leaf specific area (LSA) (Rozendaal *et al.*, 2006), succulence (SU) (Mantovani, 1999), relative water content  
117 (RWC), and water saturation deficit (WSD) (Turner 1981).

118

### 119 2.3 *Light microscopy*

120

121 For anatomical analysis, expanded and undamaged mature leaves (n = 6) were collected from individuals (n  
122 = 3) of each species (Colleen, 2013). Leaf fragments (0.5 cm<sup>2</sup>) were fixed in Karnovsky's solution (Karnovsky,  
123 1965) for 48 h, dehydrated in n-butyl series (Johansen, 1940), embedded in Paraplast® (Kraus and Arduin,  
124 1997), sectioned in a rotary microtome (12 µm) (Jung Biocut), and stained with 9:1 (v/v) solution of astra blue  
125 and safranin (Bukatsch, 1972, modified to 0.5 %).

126 Leaf traits that may affect the absorption capacity and water storage on leaves, such as the presence of  
127 trichomes, calcium oxalate crystals, secretory cavities, mucilage presence, and the degree of compactness of  
128 spongy parenchyma cells were evaluated.

129

### 130 2.4 *Histochemical analyses*

131

132 Cutin was detected by staining with Sudan red B (Brundett *et al.*, 1991). This staining was also useful for the  
133 evaluation of cuticle thickness of the adaxial and abaxial leaf surfaces. Pectin was detected with ruthenium red  
134 (Johansen, 1940). Cellulose microfibrils were labelled with calcofluor white (Hughes and McCully, 1975), a  
135 specific dye that stains the β-glucan. All sections were mounted in water and examined under a light microscope  
136 (Leica DM500), or a fluorescence microscope (Leica DM 2500-LED). Photomicrographs were obtained with a  
137 Leica DFC 7000T camera coupled to the microscope.

138

### 139 2.5 *Immunocytochemistry of cell wall epitopes*

140

141 Part of the leaf samples fixed in Karnovsky's solution (Karnovsky, 1965), and prepared for anatomical  
142 analyses were incubated with the monoclonal antibodies JIM5, JIM7, LM5 and LM6 (Centre for Plant Sciences,  
143 University of Leeds, UK). These antibodies have specificity to binding epitopes of low methylesterified  
144 homogalacturonans (HGAs) (JIM5) and high methylesterified HGAs (JIM7), galactans (LM5) (Jones *et al.*,  
145 1997), and arabinans (LM6) (Willats *et al.*, 1998). The sections were immersed in blocking solution with 3% (w

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147

148

149

146 / v) milk powder in phosphate-buffered saline (PBS) for 30 min, and incubated with primary antibodies in  
147 powder milk/PBS solution (1:10) for 2 h at room temperature. For control tests, the primary antibodies were not  
148 added. The sections were washed in PBS, incubated with secondary antibody for IgG FITC (Sigma) in powder  
149 milk/PBS solution (1:100) for 2 h in the dark. After washing in PBS, the sections were mounted in 50% glycerol  
150 and examined under the fluorescence microscope Leica DM 2500-LED with a Leica DFC 7000T coupled  
151 camera.

152

## 153 2.6 Data analyses

154

155 General linear models (GLMs) with Quasi Poisson errors were built to evaluate if the leaf specific mass  
156 (LEM), leaf specific area (LSA), succulence (SU), percentage of relative water content (RWC), and water  
157 saturation deficit (WSD) (independent variables) influenced the maximum water content in leaf ( $C_{\max}$ ), and the  
158 water uptake speed ( $k$ ) (dependent variables) of the four species.

159

## 160 3. Results

161

### 162 3.1 Water uptake and functional leaf traits

163

164 Water uptake was fast in the first few minutes, but decreased in approximately 5 h, reaching zero value, due  
165 to leaf hydration, regarding the four plant species evaluated. According to the water absorption curves,  
166 *Trembleya laniflora* had the highest amount of water uptake, followed by *Tibouchina heteromalla* and  
167 *Heteropterys campestris*, with similar values. *Ocotea pulchella* had the lowest amount of water uptake (Fig. 1).

168 *Trembleya laniflora* had the highest rate of  $C_{\max}$ , which was significantly different from the similar  
169 intermediary values of  $C_{\max}$  in *T. heteromalla* and *H. campestris*. *Ocotea pulchella* differed significantly from  
170 the other species, with the lowest  $C_{\max}$  values (Fig. 2A). The water uptake speed ( $k$ ) was similar between *O.*  
171 *pulchella* and *H. campestris*, and it was higher than the values of the other two plant species. The lowest values  
172 of  $k$ , in *T. laniflora* and *T. heteromalla*, were statistically similar (Fig. 2B).

173 Among the analysed functional leaf traits, only the specific leaf area (SLA) had a positive and significant  
174 relationship with the  $C_{\max}$  and  $k$ . The relative water content (RWC) affected the  $C_{\max}$ , while the water saturation  
175 deficit (WSD) was more affected by the water uptake speed ( $k$ ) (Fig. 5), and these results also presented a

176 positive and significant relationship (Table 1). The other physiological leaf traits had no influence on the  
177 evaluated parameters.

178

### 179 3.2 Leaf anatomy and histochemistry

180

181 The leaves of three species are hypostomatic, and *T. heteromalla* is amphistomatic. The cuticle is thick on the  
182 adaxial leaf surface of two species, *T. laniflora* and *O. pulchella*, while the other two species, *T. heteromalla* and  
183 *H. campestris*, have thin cuticle. The cuticle is thin on the abaxial surface of all the four species (Fig. 3A-D). The  
184 four species have uniseriate epidermis with larger cells on the adaxial surface in relation to the abaxial leaf  
185 surface. Non-glandular trichomes occur on the abaxial leaf surface of *T. laniflora* and on the adaxial surface of  
186 *O. pulchella*. *Trembleya laniflora* also has short-pedunculated glandular trichomes, exclusively on the abaxial  
187 leaf surface (Fig. 3A). *Tibouchina heteromalla* has non-vascularized emergencies on both leaf surfaces,  
188 including on the venal region. These emergencies are lined by uniseriate epidermis, have some idioblasts  
189 containing druses on the cells of the ground system, and filiform sclereids interspersed (Fig. 3B). The mesophyll  
190 is dorsiventral in the four species, with 1-2 layered palisade parenchyma in three species and 3-layered in *T.*  
191 *laniflora*. The spongy parenchyma has 3-4-layers in *T. heteromalla*, *T. laniflora* and *H. campestris*, with variable  
192 area of intercellular spaces (Fig. 3C). *Ocotea pulchella* has the most distinct leaf lamina due to the occurrence of  
193 some round-shaped hypertrophied secretory idioblasts, containing oils, phenols and mucilage, interspersed to  
194 palisade parenchyma cells (Fig. 3D). The spongy parenchyma in this species has large intercellular spaces and  
195 has 5-7 cell layers. Two species, *T. laniflora* and *T. heteromalla*, have vascular bundles in bicollateral  
196 arrangement and two, *H. campestris* and *O. pulchella*, have vascular bundles in collateral arrangement (Fig. 3A-  
197 D). The bundle sheaths store phenolics and have interspersed sclereids in *T. laniflora*, and druses of calcium  
198 oxalate in *T. heteromalla* and *H. campestris* (Fig. 3 B-C). Sheath extensions with lignified fibres communicate  
199 the vascular bundles to both sides of the epidermis in *O. pulchella*. The abaxial epidermis has papillose cells and  
200 some glandular trichomes in *T. heteromalla*.

201 The leaves of *T. laniflora* and *T. heteromalla* have positive reaction for pectins detected by the ruthenium red  
202 in the CW of all tissues, including the vascular bundles, and the adaxial epidermis. In leaves of *H. campestris*,  
203 the positive reaction for ruthenium red occur in the CW of the adaxial epidermis, vascular bundle sheaths and  
204 collenchyma. There is also positive reaction for pectins in some cells of the abaxial epidermis, and in glandular  
205 trichomes. *Ocotea pulchella* has positive reaction on the CW on both epidermal surfaces and parenchyma cells

206 (data not shown). All species have positive reaction for calcofluor white in epidermal cells, which has low level  
1 of recognition in *O. pulchella* (Fig. 3 E-H).  
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### 5 209 3.3 Immunocytochemistry of cell wall epitopes 6 7

8 210  
9

10 211 In *T. laniflora*, the epitopes of low methyl-esterified HGAs were labelled by JIM5 overall the CW of the  
11 spongy parenchyma, idioblasts, and xylem and phloem (Fig. 3I). The epitopes of high methyl-esterified HGAs  
12 212 bound by JIM7 were restricted to some CW of the spongy parenchyma (Fig. 4A). No epitopes of galactans were  
13 213 labelled by LM5 (Fig. 4E). Epitopes of arabinans were abundantly labelled by LM6 in the CW of the palisade  
14 214 parenchyma (Fig. 4I).  
15  
16 215

17  
18 216 In *T. heteromalla*, the low methyl-esterified HGAs had a low level of recognition by JIM5 in the CW of both  
19 217 epidermal surfaces, emergencies, spongy parenchyma, xylem and phloem (Fig. 3I). The high methyl-esterified  
20 218 HGAs had an overall but low level of recognition by JIM7 in the CW of all leaf tissues (Fig. 4B). The epitopes  
21 219 of galactans were labelled by LM5 throughout the CW of the palisade parenchyma and abaxial epidermis (Fig.  
22 220 4F). The epitopes of arabinans labelled by LM6 were restricted to the stomata (Fig. 4J).  
23  
24 221

25 222 In *H. campestris*, the epitopes of low methyl-esterified HGAs were labelled by JIM5 in the CW of the  
26 223 adaxial epidermis (Fig. 3K). The high methyl-esterified HGAs had abundant detection by JIM7 in the CW of  
27 224 adaxial epidermis and palisade parenchyma (Fig. 4C). The epitopes of galactans labelled by LM5 were restricted  
28 225 to the CW of the idioblasts and the epitopes of arabinans were labelled by LM6 only in some cells of the  
29 226 vascular bundles (Fig. 4G, K).  
30  
31 227

32 228 In *O. pulchella*, the epitopes of low methyl-esterified HGAs were labelled by JIM5 in the CW of adaxial  
33 229 epidermis, in parenchyma cells, and in the vascular bundles (Fig. 3L). The high methyl-esterified HGAs labelled  
34 230 by JIM7 were overall abundant in the CW of all leaf tissues (Fig. 4D). The epitopes of galactans labelled by  
35 231 LM5 were restricted to a few cells of spongy parenchyma (Fig. 4H), while the epitopes of arabinans recognized  
36 232 by LM6 were restricted to some cells of the vascular bundles (Fig. 4L).  
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## 43 233 4. Discussion 44 45

46 234

### 47 235 4.1 LWU, morphology and leaf anatomy 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65

236 Different strategies of LWU were clearly revealed in all the four species, with peculiarities among them. The  
1  
2 237 amount and speed of water absorption are not only related to the presence of trichomes and thin cuticle, but also  
3  
4 238 to more compactly arranged tissues and mucilaginous cells in the mesophyll. Water storage sites indicate greater  
5  
6 239 capacity for water absorption, while the CW with high detection of pectin are related to an increase in water  
7  
8 240 absorption speed in leaves.

9  
10 241 As the RWC describes the plant water status at a given time (Kramer, 1969), it is interesting to note that there  
11  
12 242 is a close relationship between the RWC and the LWU strategies. The fastest absorbing species are those with a  
13  
14 243 higher initial RWC, when compared to the slower absorbing species. In other words, a higher RWC is a  
15  
16 244 consequence of a higher absorption rate, since a smaller value of WSD positively interferes in the value of  $k$ .  
17  
18 245 Thus, it may be highlighted that the species with faster absorption have higher RWC values, which may be a  
19  
20 246 strategy to avoid drought effects. Moreover, different strategies were observed; *T. laniflora*, followed by *T.*  
21  
22 247 *heteromalla* had higher rates of  $C_{max}$  and the smallest values of  $k$ . *O. pulchella* and *H. campestris* had the highest  
23  
24 248 values of  $k$  and smallest values of  $C_{max}$  (Fig. 2).

25  
26 249 Morphological and anatomical features may significantly interfere in the LWU. The cuticle is the main route  
27  
28 250 through which water must pass during leaf absorption (Eller et al., 2013; 2016). However, the degree of  
29  
30 251 permeability may be related to the cuticle chemical composition and to the arrangement of the epicuticular wax,  
31  
32 252 and not necessarily to cuticle thickness (Schuster et al., 2016), as herein observed for the four species. The leaves  
33  
34 253 of *T. laniflora* and *T. heteromalla*, despite the differences in cuticle thickness, absorbed similarly more water in  
35  
36 254 the 5-hour period of the experiment, but more slowly. The trichomes positively influence the LWU (Lusa et al.,  
37  
38 255 2014; Eller et al., 2016; Vitarelli et al., 2017), as observed in *H. campestris* and *O. pulchella*, the only species  
39  
40 256 with non-glandular leaf trichomes. Accordingly, only *T. heteromalla* has emergencies on its leaves (Fig. 3B),  
41  
42 257 which may also facilitate the LWU, while the mucilage in the leaf tissues of *H. campestris* and *O. pulchella* may  
43  
44 258 enable water retention due to its hygroscopic trait, as indicated by Fahn and Cutler (1992).

45  
46 259 The compactness of the spongy parenchyma in *T. laniflora* and *T. heteromalla* has a direct relationship with  
47  
48 260 the strategy of these species to absorb a larger amount of water (Fig. 3A-B). Since water is mainly stored in the  
49  
50 261 vacuoles, the higher the number of vacuolated large cells, the higher is the availability of cell compartments to  
51  
52 262 store water. Thus, these anatomical attributes help water retention within leaf tissues, and with more  
53  
54 263 compartments for water storage and therefore higher  $C_{max}$  (Tomlinson, 2003). Contrastingly, *O. pulchella*, which  
55  
56 264 had the second smaller  $C_{max}$ , has a loose spongy parenchyma, with large intercellular spaces. Also, the presence  
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265 of oil-containing idioblasts also decreases the number of vacuoles for water storage, corroborating the initial  
1 premises.  
2

3  
4 267 Current results indicate a trade-off between the LWU capacity and the water storage. Species with greater  
5  
6 268 LWU capacity tend to have limited water storage capacity (Gotsch et al., 2015), compensating it with rapid leaf  
7  
8 269 water uptake. However, they can avoid the loss of cell turgor, keeping high structural allocation and water  
9  
10 270 storage, as seen in epiphytes of Costa Rica, which have a thicker hydrenchymal layer (Gotsch et al., 2015).  
11  
12 271 Accordingly, similar strategies should occur with two of the four species currently studied, *T. laniflora* and *T.*  
13  
14 272 *heteromalla*, which have a slower water uptake, but a higher degree of water storage.  
15

16 273

#### 17 18 274 4.2 Cell wall composition

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21  
22 276 There is direct relationship between the absorption rate, the amount of water absorbed, and the CW  
23  
24 277 composition in the four species herein studied. Species that have abundant detection of cellulose in their  
25  
26 278 epidermal CW, i. e., *T. laniflora* and *T. heteromalla*, can absorb more water, but more slowly, while those with  
27  
28 279 abundant detection of pectins, especially HGAs, can absorb water at a higher rate.

29  
30 280 Once water molecules have passed the cuticle, the next barrier is the external periclinal CW. As one of the  
31  
32 281 main components of the CW, cellulose was histochemically detected in distinct levels of recognition in the CW  
33  
34 282 of the adaxial epidermal cells in all the studied species by the calcofluor. For both *H. campestris* and *O.*  
35  
36 283 *pulchella*, low fluorescence was considered indicative of less cellulose in their epidermal cells in relation to the  
37  
38 284 other species. Cellulose microfibrils, when interacting with water, form gels that may increase the amount of  
39  
40 285 water absorbed, but even though these gels have hygroscopic property and keep wall hydration, they slow the  
41  
42 286 absorption rate due to their physical properties (Pääkkö et al., 2007; Agoda-Tandjawa et al., 2012). Current data  
43  
44 287 indicate that the interlacing of cellulose microfibrils may reduce the CW porosity, and therefore slow the speed  
45  
46 288 of water uptake.

47  
48 289 Cell walls can compensate the low proportion of cellulose by the deposition of HGAs (His et al., 2001). The  
49  
50 290 proportion of cellulose:pectin can differ between species, and once it is very dynamic, it can be modified in  
51  
52 291 response to several external and internal factors (Cosgrove, 2005). Currently, pectins were detected by ruthenium  
53  
54 292 red in the CW of the adaxial epidermal cells, in the four species studied. However, the MABs detected slight  
55  
56 293 differences in the pectin classes. The absence of labelling for all the tested MABs in the CW of *T. laniflora*  
57  
58 294 epidermis indicates low pectin proportion, as proposed as an overall assumption (Albersheim et al., 2011),  
59

295 making it difficult to be detected. The greater amount of water absorbed by *T. laniflora* can be explained by the  
1 overall and higher detection of cellulose microfibrils in the CW of the adaxial epidermis, which is reinforced by  
2  
3  
4 297 the CW traits of *T. heteromalla*, the second species with the higher LWU.

5  
6 298 The HGAs of high and low methyl esterification are the most abundant pectin group in the CW  
7  
8 299 (Verhertbruggen et al., 2009), as is true for two of the four species of the tropical fog mountain fields herein  
9  
10 300 studied. The overall labelling of HGAs in *H. campestris* and *O. pulchella* corroborates the assumption that their  
11  
12 301 cell walls are pectin-rich, mainly rich in HGAs, and indicates the role of HGAs as determinant in the increase of  
13  
14 302 the water absorption speed in these two species. Thus, abundant pectins in the CW of the adaxial epidermis of  
15  
16 303 this two species may explain the high speed of LWU.

17  
18 304 The pectins form hydrated gels that displace the cellulose microfibrils, and as they fill in the microfibrillar  
19  
20 305 spaces, they avoid the aggregation and collapse of the cellulose network (Wolf et al., 2009). A study carried out  
21  
22 306 with *Phaseolus vulgaris* evidenced that reducing the amount of pectins at root apices under osmotic stress altered  
23  
24 307 the structure of the CW, due to a rearrangement of wall polymers, affecting cell porosity (Cosgrove, 1997;  
25  
26 308 Cosgrove, 2005). Therefore, pectins ensure the CW porosity, and consequently a higher water absorption rate, as  
27  
28 309 observed for *H. campestris* and *O. pulchella*, which presented the fastest LWU.

29  
30 310 The degree of methyl esterification of the HGAs has a direct influence on the CW structure and consequently  
31  
32 311 on cell physiology (Wolf et al., 2009). The HGAs with low degree of methyl esterification form bonds with  $Ca^{2+}$ ,  
33  
34 312 forming an "egg-box" structure (Liners et al., 1989), and consequently, a greater interaction with the water.  
35  
36 313 Therefore, the decrease in the degree of methyl esterification increases the interaction with water, and makes the  
37  
38 314 CW more hydrophilic (Faravash and Ashtiani, 2007). Accordingly, the HGAs with a low degree of methyl  
39  
40 315 esterification should be more related to LWU than those with a high degree of methyl esterification. The  
41  
42 316 increasing in the detection of HGAs with low methyl esterification may indicate a compensation to the  
43  
44 317 decreasing in the restricted detection of cellulose in the CW. In *H. campestris* and *O. pulchella*, cellulose  
45  
46 318 microfibrils had a restricted detection, in the epidermal cells, by the calcofluor white, but the HGAs with low  
47  
48 319 degree of methyl esterification were abundantly detected by JIM5. Consequently, it is interesting to note that an  
49  
50 320 increase in the labelling of low methyl esterified HGAs can balance the apparently low synthesis and deposition  
51  
52 321 of cellulose in the CW (His et al., 2001; Manfield et al., 2004).

53  
54 322 The substitution of part of the HGAs by RG-I or RG-II is related to an increase in the CW porosity, and thus  
55  
56 323 to the movement of proteins and hydrolytic enzymes within the wall matrix (Iraki et al., 1989). Peculiarly, *H.*  
57  
58 324 *campestris* was the only species where galactans were restricted labelled in some abaxial epidermal cells, and as  
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325 galactans are related to stability and rigidity in CW of the leaf epidermis (Xu et al., 2011), a greater amount of  
1 326 this lateral chains of RG-I may be associated with a higher rate of absorption. The CW composition of *H.*  
2  
3 327 *campestris* seems to favour the porosity of the wall and the speed of water absorption in its leaves.  
4  
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## 7 329 **5. Conclusion**

9 330  
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11 331 The plants that presented the fastest leaf water absorption have an overall labelling of pectins, mainly HGAs,  
12  
13 332 in their epidermal CW. However, plants that absorb more water, even more slowly, have abundant detection of  
14  
15 333 cellulose microfibrils in the CW of epidermal cells. These results indicate that cellulose and pectins are  
16  
17 334 determinant in water absorption by leaf epidermis, conferring greater porosity and absorption speed, regarding  
18  
19 335 pectin composition; or greater hydrophilicity and greater absorption capacity, regarding cellulose. In conclusion,  
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21 336 it seems that the determining trait in leaf water absorption in the studied species is the composition of the  
22  
23 337 epidermal CW, although other traits such as trichomes are known to influence the process. In addition, the  
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25 338 presence of more compact tissues and mucilaginous cells in leaves may also influence leaf water absorption and  
26  
27 339 water storage capacity.  
28  
29 340

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42 347

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503 **Figure captions**

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4 505 **Fig. 1.** Dynamic water content in the leaves by submerging time (min). Equations: *Tibouchina heteromalla*  
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6 506 ( $y=0.55751-e^{-0.0372t}$ )  $r^2=0.9341$ ; *Ocotea pulchella*  $y=0.0789(1-e^{-0.0624t})$   $r^2= 0.971$ ; *Trembleya laniflora*  
7  
8 507  $y=0.7985(1-e^{-0.026t})$   $r^2= 0.936$ ; *Heteropterys campestris*  $y=0.4508(1-e^{-0.0621t})$   $r^2= 0.869$ .

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12 509 **Fig. 2.** Maximum water content (Cmax) in (A) and water uptake speed (k) in (B) of the evaluated species. The  
13  
14 510 mean values ( $\pm$ SE) of the leaf uptake maximum amount standardized by projected leaf area. The standardized  
15  
16 511 mean values ( $\pm$ SE). ( $P \leq 0.05$  between treatments).

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20 513 **Fig. 3.** Immunocytochemistry of plant cell wall pectins. (A, E, I) *Trembleya laniflora*. (B, F, J) *Tibouchina*  
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22 514 *heteromalla*, (C, G, K) *Heteropterys campestris* (D, H, L) *Ocotea pulchella*. (A–D) Safranin and astra blue; (E–  
23  
24 515 H) Calcofluor white; (I–L) JIM5. AE: adaxial epidermis; PP: palisade parenchyma; SP: spongy parenchyma; Em:  
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26 516 emergencies; Ae: aerenchyma; SE: bundle sheath extension.

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30 518 **Fig. 4.** Immunocytochemistry of plant cell wall pectins. (A, E, I) *Trembleya laniflora*. (B, F, J) *Tibouchina*  
31  
32 519 *heteromalla*, (C, G, K) *Heteropterys campestris* (D, H, L) *Ocotea pulchella*. (A–D) JIM 7, (E–H) LM5, (I–L)  
33  
34 520 LM6. AE: adaxial epidermis; PP: palisade parenchyma; SP: spongy parenchyma; ID: idioblast; ST: stomata; Xy:  
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36 521 xylem; Ph: phloem.

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40 523 **Fig. 5.** Relative water content (RWC) and water saturation deficit (WSD) (%) of *Heteropterys campestris*,  
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42 524 *Ocotea pulchella*, *Tibouchina hetemalla* and *Tremblyea laniflora*.

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## Contributions

The study was conceived by MGCF and RMSI. DB, BGF, ARZ and HCS performed the experiments.

DB computed and analyzed data and DB, MGCF and RMSI wrote the manuscript.

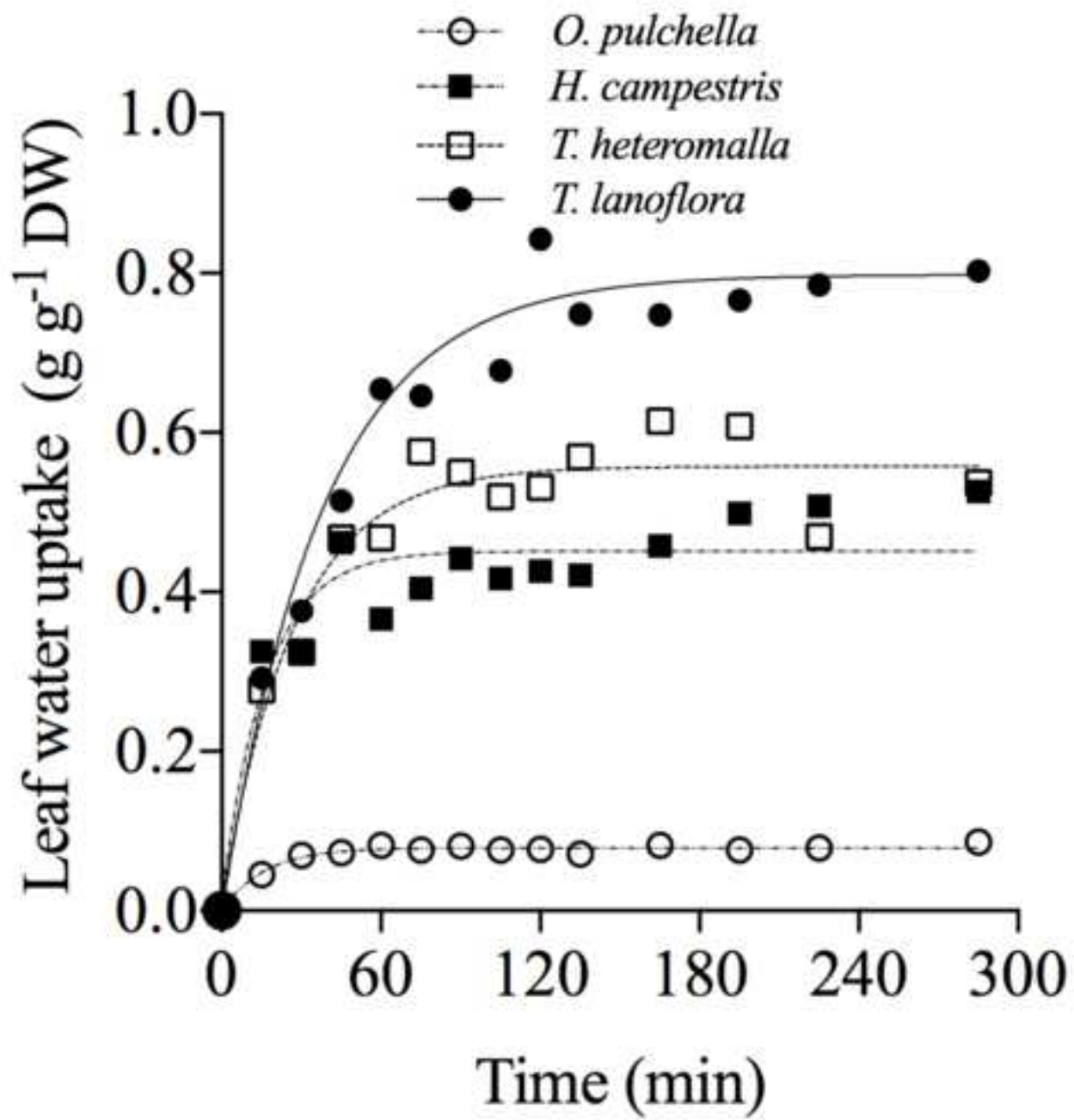
**Table 1**

Water leaf content ( $C_{\max}$ ) and water uptake speed ( $k$ ) related to leaf traits of the evaluated plants. Specific leaf weight (SLW), specific leaf area (SLA), succulence (SU) and relative water content (RWC) and water saturation deficit (WSD). In these analyses, dependent variables were maximum leaf water content ( $C_{\max}$ ) and water uptake speed ( $k$ ).

Dependent variable	Independent variable	Deviance	F	DV	<i>P</i>
$C_{\max}$	SLW	3.5368	0.1731	18	0.6837
	SLA	2.0768	15.1418	17	0.0016 **
	SU	2.0742	0.0268	16	0.8722
	RWC	1.6541	4.3563	15	0.05064 *
	WSD	1.5381	1.2034	14	0.2911
$k$	SLW	0.4944	0.0025	18	0.9607
	SLA	0.2512	23.2187	17	0.0002 ***
	SU	0.2421	0.8686	16	0.3671
	RWC	0.2289	1.2574	15	0.281
	WSD	0.1433	8.1740	14	0.0126**

The general linear models (GLMs), with Quasipoisson errors, for the biotic independent variables evaluated. In these analyses, dependent variables were maximum water leaf content ( $C_{\max}$ ) and water uptake speed ( $k$ ). \*\*\* Indicates  $P \leq 0.001$ ; \*\*Indicates  $P \leq 0.01$  and \* Indicates  $P \leq 0.05$ .

Figure 1  
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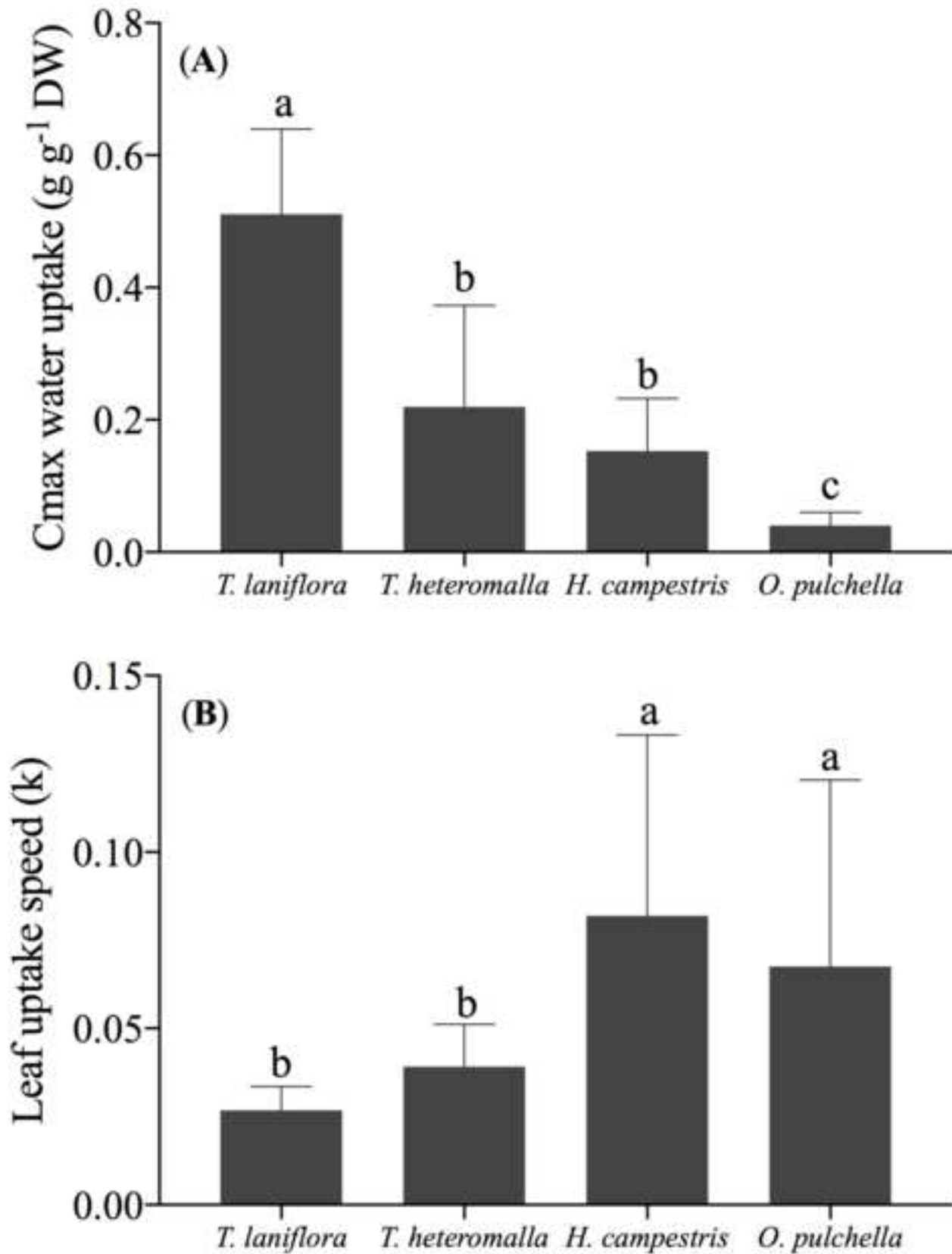


Figure 3  
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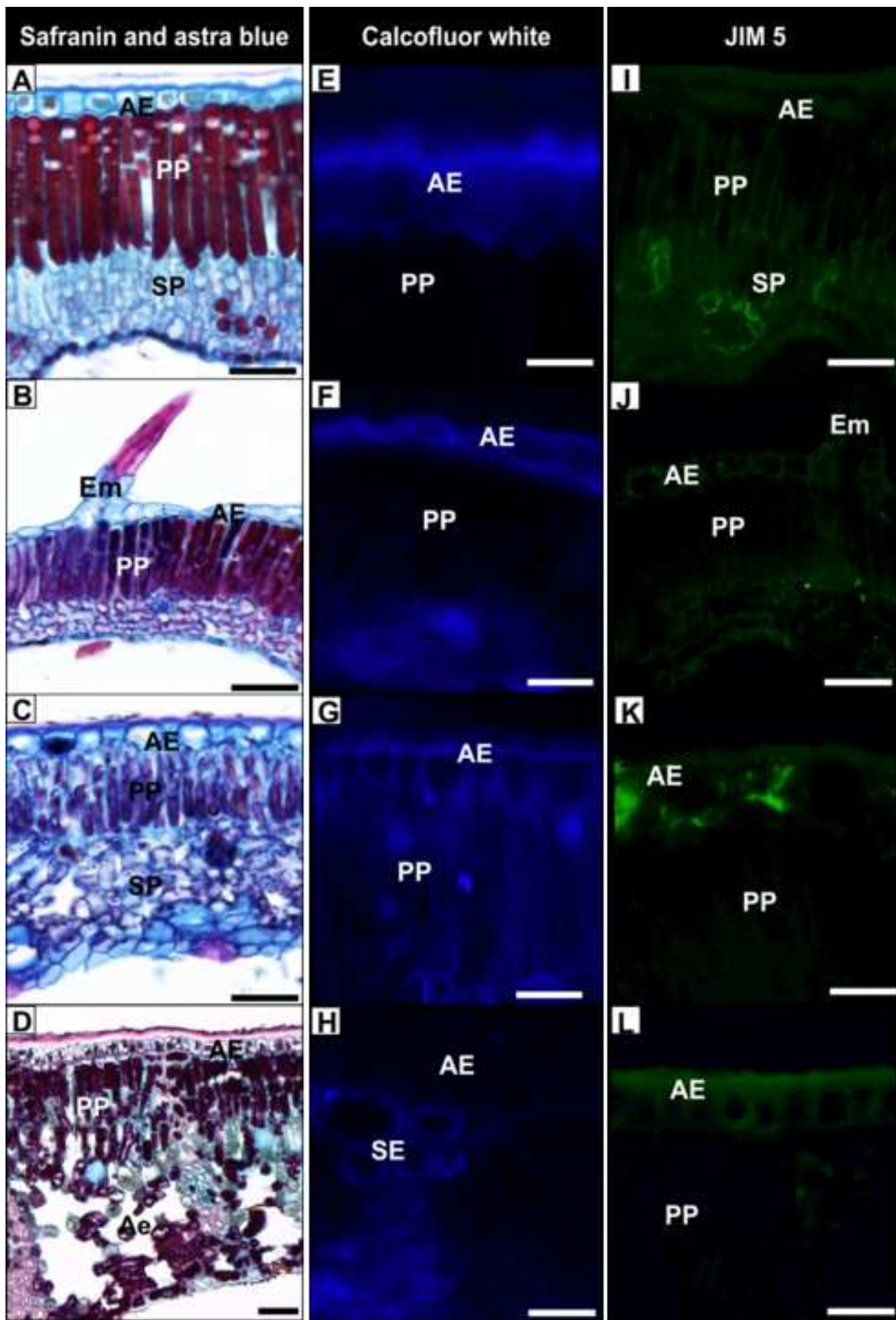


Figure 4  
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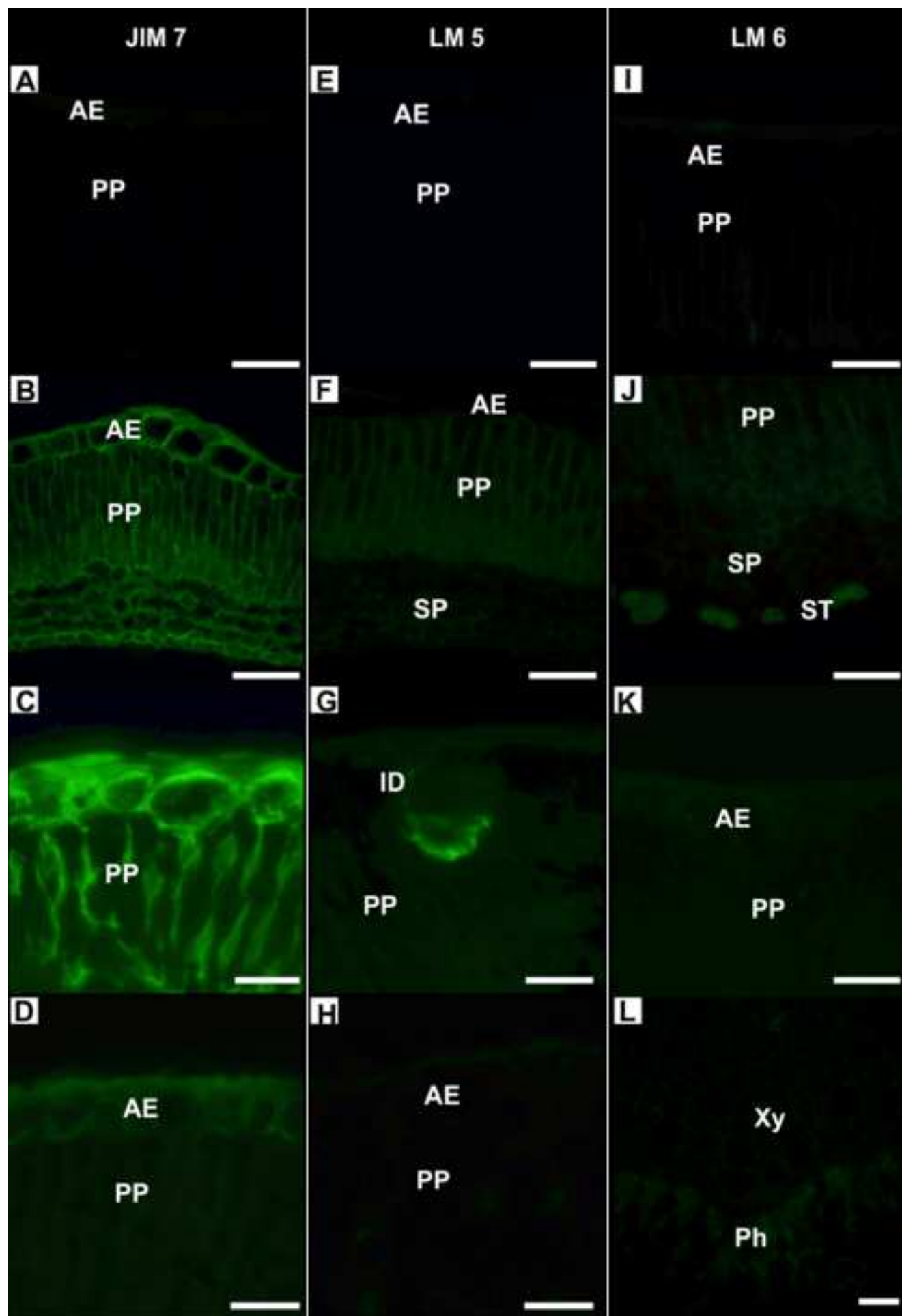
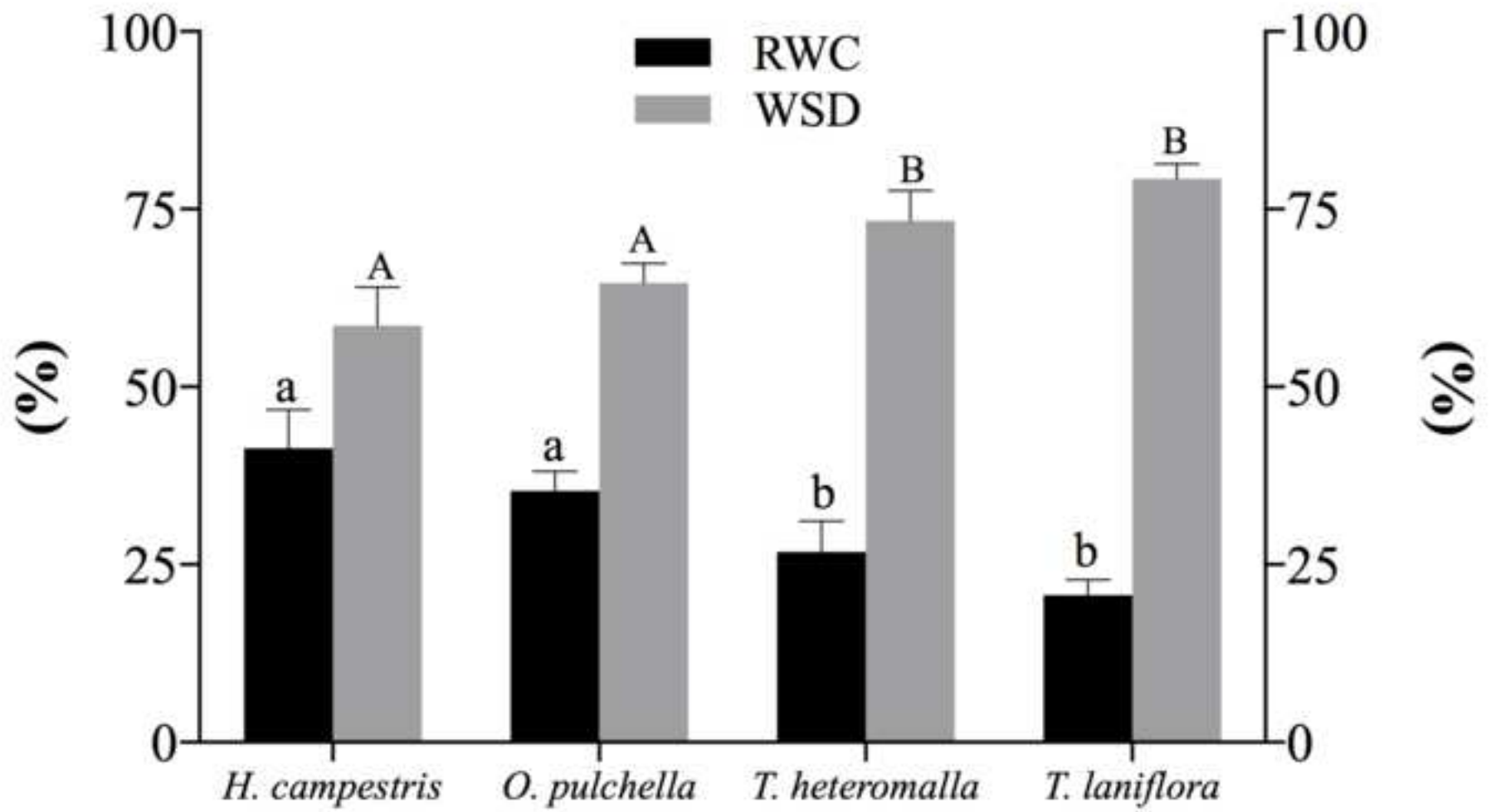
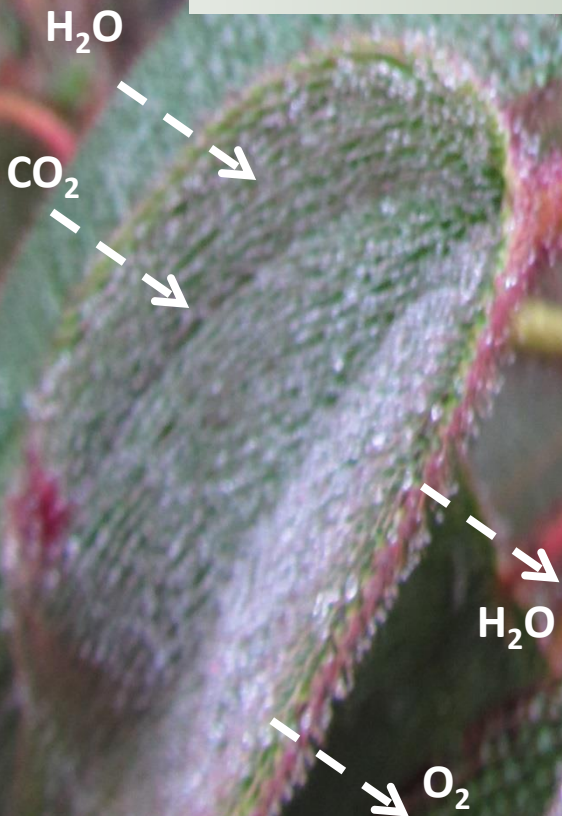


Figure 5  
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*Foliar water-uptake strategies are related to leaf water status and gas exchange in plants from a ferruginous rupestrian field*



1 **Foliar water uptake strategies are related to both leaf water status and gas**  
2 **exchanges in plants from a ferruginous rupestrian field**

3

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14

15 **PREMISE OF THE STUDY:** Fog plays an important role in the physiology of several plant  
16 species. It is known that foliar water uptake (FWU) of fog happens in different ways: a faster  
17 and a slower water fog uptake. However, water fog may play a negative role in CO<sub>2</sub>  
18 assimilation. Thus, the interference of different strategies of FWU in the water and carbon  
19 balance were evaluated to verify if fog may mitigate possible leaf water deficit.

20

21 **METHODS:** Four plant species with different FWU strategies were studied in a ferruginous  
22 rupestrian field, where fog is a frequent event. Measurements of gas exchanges and water  
23 potential in predawn and midday were performed during dry and rainy seasons, separating  
24 foggy from non-foggy days during the dry season.

25

26 **KEY RESULTS:** The FWU speed negatively influences CO<sub>2</sub> assimilation in the dry season,  
27 possibly because of its negative relationship with stomatal conductance, since reduced  
28 stomatal aperture impairs carbon entrance. Fog presence increased leaf water potential both in  
29 early morning and mid-day during the dry season. However, during the rainy season, the  
30 values of leaf water potential were higher at noon than at midday, which favors leaf gas  
31 exchanges.

32

33 **CONCLUSIONS:** FWU interferes negatively, but briefly with CO<sub>2</sub> assimilation.  
34 Nevertheless, FWU prevents water loss through transpiration and increases the water status of  
35 plants in the dry season. That is, FWU generates a compensation between CO<sub>2</sub> assimilation  
36 and foliar hydration, which, in fact, is beneficial to the plants of this ecosystem.

37

38 **KEY WORDS;** gas exchanges; photosynthesis; Rupestrian field, water relations.

39

40 The ferruginous rupestrian fields are considered an ancient climatically protected infertile  
41 landscape (OCBIL) such as Kwongkan in South-West Australia, the fynbos in South-West  
42 South Africa, and the tepuis in the Shield of Guyana in Northern South America (Hopper,  
43 2009). This ecosystem has great diversity with high endemism, and rare species (Silveira et  
44 al., 2016). Ferruginous rupestrian fields are usually characterized by a mosaic of grassy-shrub  
45 and arboreal vegetation, prone to fire occurring on outcrops hematite rich sites, such as  
46 itabirites or *cangas*, that is, on shallow and discontinuous soils. A peculiar feature of this  
47 ecosystem is the presence of fog due to its elevated altitude. In addition to contributing to the  
48 hydrology of ecosystems, fog may play a relevant ecological role as an alternative source of  
49 water for vegetation (Burgess and Dawson, 2004), once many species depend on an  
50 alternative source of water in drought periods (Dawson, 1998; Williams et al., 2008). During

51 periods without fog, tropical altitudinal environments can be considered arid due to the high  
52 evaporative demand (Leuschner, 2000).

53 When fog is present, foliar water uptake (FWU) is a strategy for leaf hydration. Foliar  
54 absorption occurs when water surpasses a leaf surface in response to the water potential  
55 gradient diffusing through the cuticle (Gouvra and Grammatikopoulos 2003), absorbent  
56 trichomes (Benzing et al., 1978) and through the hydrotodes (Martin and von Willert, 2000).  
57 Water absorption and particles can occur soon after different forms of precipitation, such as  
58 fog and rain.

59 Although the interception and absorption of water by the leaves have been reported in  
60 publications since the 1960 (Stone 1963), studies on the mechanisms and pathways of foliar  
61 water uptake (FWU) have become more present in recent decades (Burgess and Dawson,  
62 2004; Anderson, 2005; Lai et al., 2007; Liang et al., 2009; Qiu et al., 2010; Goldsmith, 2013;  
63 Eller et al., 2016; Berry et al., 2018). Several studies have already revealed the advantages of  
64 FWU in the physiology of plant species, such as increased water status and benefits for gas  
65 exchange (Grammatikopoulos and Mannetas, 1994; Martin and von Willert, 2008; Simonin et  
66 al., 2009). The importance of FWU for gas exchanges and leaf water status has not yet been  
67 described for any shrub and woody species of Brazilian ferruginous rupestrian fields. The  
68 water acquired by FWU is considered of ecological relevance for dry or seasonally dry  
69 environments, where fog events are frequent (Oliveira et al., 2014). This strategy may  
70 facilitate the physiological activity, growth and establishment of plant species, since the  
71 frequency of fog seems to be functionally linked to the patterns of distribution of certain  
72 species and types of ecosystems (Qiu et al., 2010).

73 Due to selective limitations imposed by water, species that occur in places with water  
74 deficit have different mechanisms of water capture or maintenance (Reich et al., 2003), and  
75 consequently different adaptive strategies. In some species, water can be absorbed quickly by

76 leaves, but in smaller amounts. However, there are species whose leaves absorb more water  
77 but more slowly. Indeed, 11 species of this ecosystem have different water absorption  
78 capacities (Boanares et al., 2018a and b), and such knowledge may help clarify the  
79 coexistence of species adapted to the temporal and spatial water availability variations that the  
80 tropical environment is subject to. Such adaptations are key factors for the understanding of  
81 species diversity (Tyree et al., 2003), mainly in relation to climate changes. After all,  
82 pragmatically, metal-rich environments, such as ferruginous rupestrian fields, have a flora  
83 with restricted geographic distribution and subjected to several iron mining activities (Silveira  
84 et al., 2016). In this way, several species are also endangered due to this anthropic activity  
85 (Martinelli and Moraes, 2013).

86 The hypothesis of this study is that FWU strategies interferes positively in the water and  
87 carbon balance of the species studied. Thus, increased carbon assimilation and leaf hydration  
88 would attenuate possible water stress.

89

## 90 **MATERIALS AND METHODS**

91

### 92 **Study area and plant species**

93

94 This study was carried out in a region of ferruginous rupestrian field in the "Cachoeira das  
95 Andorinhas" Environmental Protection Area, located in the municipality of Ouro Preto, Brazil  
96 (20 ° 21'30 "S / 43 ° 30'11 "W). The local ecosystem is characterized by species of herbs,  
97 shrubs and treelets growing on shallow soils of rock outcrops of itabirites or *cangas* (Valim et  
98 al., 2013). The area has an average altitude of 1,492 m, where fog events are frequent (Fig.1).  
99 The study was conducted in February and July, 2017, the most evident months of rainy and  
100 dry seasons, respectively. The average annual temperature and precipitation were 16.9°C and

101 100.4 mm, respectively, with the total annual rainfall of 1,204.7 mm, 95.5% concentrated  
102 from October to March (Fig. 2). Fog accounts for approximately 13% of rainfall entries  
103 during the year. However, in the dry season, it contributed 10 times more in relation to  
104 rainfall (Baeta, 2012).

105 The studied Phanerophytes species were: *Leandra australis* (Cham.) Cogn. (shrub) and  
106 *Miconia corallina* Spring. (arboreal) (Melastomataceae), *Eremanthus erythropappus* (DC.)  
107 MacLeish, (Asteraceae) (arboreal), *Byrsonima variabilis* A. Juss., (Malpighiaceae) (shrub), all  
108 endemic of Brazil except *Leandra australis*.

109 These species were selected according to their different FWU capacities (Boanares et al.,  
110 2018b). *Leandra australis* and *Byrsonima variabilis* have a higher speed of leaf water  
111 absorption, but uptake a small amount of water. However, *Eremanthus erythropappus* and  
112 *Miconia corallina* have slow absorption, but uptake a large amount of water through the  
113 leaves (Table 1).

114

#### 115 **Measurements of foliar water uptake**

116

117 To evaluate leaf water absorption capacity, leaves ( $n = 4$ ) were collected in the field and  
118 immediately weighed with a portable digital scale. Leaves were then taken to the laboratory  
119 and weighed once more, and the petiole was sealed with hot paraffin. Subsequently, the  
120 leaves were immersed in distilled water and weighed every 15 min for 2 h, then weighed  
121 every 30 min for 2 h. Finally, the leaves were immersed in water again for 1 h and weighed to  
122 obtain the maximum water absorption curve. Before each weighing, the leaves were dried  
123 with synthetic towels of ultra-absorbent microfibers to remove surface water (Liang et al.,  
124 2009). Leaf water uptake was calculated by a differential equation, which provides data on  
125 maximum leaf water ( $C_{\max}$ ) and leaf uptake velocity ( $k$ ) ( $\text{g g}^{-1}$  dry weight) (Liang et al., 2009).

126

127 **Measurements of water potential and gas exchanges**

128

129 The water potential ( $\Psi_w$ ) was measured on three leaves of four individuals of each species at  
130 predawn ( $\Psi_{pd}$ ) and at midday ( $\Psi_{md}$ ) with a Scholander type pressure chamber in the dry  
131 (July) and rainy seasons (February). Predawn measurements were taken between 5:00 and  
132 6:00 am, and midday measurements were taken between 12:00 and 1 pm. In the dry season,  
133  $\Psi_w$  was measured at midday with and without fog.

134 Gas exchanges were measured in leaves of four individuals of each species in the dry and  
135 rainy season. A portable infrared gas analyzer (IRGA) (LI6400, LI-COR, Lincoln, Nebraska)  
136 equipped with artificial light and an integrated fluorometer (Li-6400-40, Li-Cor, Inc) was  
137 used for light curves and one-off measurements. The light curve measurements were  
138 performed from 9:00 to 11:00 am, with the addition of photosynthetic active radiation (PAR)  
139 from 0  $\mu\text{mol m}^{-2} \text{s}^{-1}$  to 2600  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . One-off measurements of gas exchanges were taken  
140 at 9:00 am with PAR saturation obtained by the light curve (1500  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ).  
141 Measurements of net  $\text{CO}_2$  assimilation ( $A$ ), transpiration ( $E$ ), stomatal conductance ( $g_s$ ) and  
142 internal carbon and internal: ambient carbon ratio ( $C_i/C_a$ ) were obtained. Water use efficiency  
143 ( $\text{WUE} = A/E$ ) and intrinsic water use efficiency ( $i\text{WUE} = A/g_s$ ) were also calculated.

144

145 **Data analysis**

146

147 Data normality was tested using the Kolmogorov-Smirnoff test with 5% significance. A  
148 variance analysis (ANOVA) followed by Tukey test at 5% was performed to verify if there  
149 was a difference in the FWU among plant species from a comparison between the mean  
150 values of  $C_{\text{max}}$  and  $k$  in the dry and rainy seasons. Linear Mixed Models (LMM) (Bolker et al.,

151 2009) were used to evaluate the influence of water absorption capacity on the physiological  
152 parameters related to gas exchange. The two parameters related to water absorption capacity,  
153 maximum water content ( $C_{\max}$ ) and leaf uptake speed ( $k$ ) were used as fixed explanatory  
154 variables and the response variables were:  $A$ ,  $E$ ,  $g_s$ ,  $C_i$ ,  $C_i/C_a$ , WUE and  $iWUE$ . The identity  
155 of each measured plant was used as a random effect variable to control for the potential  
156 pseudoreplication biases in our models (*sensu* Hurlbert 1984). Highly correlated variables  
157 were removed from the models ( $C_i$  and  $iWUE$ ).

158 To analyze the relationship between FWU and  $\Psi_w$ , a model selection was performed. Thus,  
159 an interaction between  $C_{\max}$  and  $k$  was made as two categorical factors and, as explanatory  
160 variables, the value of the water potential at predawn ( $\Psi_{pd}$ ) and at midday ( $\Psi_{md}$ ) in the dry  
161 and rainy seasons were selected. Individuals and species in both models were used as random  
162 factors (Appendix S1).

163 The full model was subjected to model selection using the Akaike Information Criteria of  
164 second order (AICc), and the best set of models was selected through multimodal inference  
165 analysis (Burnham et al., 2011). Any given model was considered superior to the other,  
166 whenever the deltaAICc between the top model and the others was higher than two (Burnham  
167 et al., 2011). For all adjusted models used in this study, residual analysis was performed to  
168 check for model suitability and adequacy of the distribution used (Crawley, 2012). All  
169 analyzes were performed on the R platform (R-Core-Team, 2015) and the computational  
170 packages used were: Ordinations and PERMANOVA - *vegan*, *gdata*, *GLMM-lme4*.

171

## 172 **RESULTS**

173

### 174 **Strategies of FWU and gas exchanges**

175

176 A significant relationship was observed among foliar water absorption rate ( $k$ ) and  $A$ ,  $g_s$  and  
177  $E$ . There was no relationship between  $C_{max}$  and gas exchange parameters (Table 2), and  $A$ ,  $g_s$   
178 and  $E$  were inversely proportional to  $k$  (Fig. 3).  $A$  and  $g_s$  are significantly influenced by  $k$ , by  
179 species and by different seasons (Table 2). The differences between  $A$  and  $g_s$  in relation to  $k$   
180 are more evident in the dry season (Fig. 3). The dry season influenced negatively these  
181 parameters, except for *E. erythropappus* that did not have differentiated values of  $A$  and  $g_s$   
182 between the seasons. Only *M. corallina* and *E. erythropappus* differed significantly from each  
183 other in relation to  $A$ . For *M. corallina*,  $g_s$  is significantly different from *E. erythropappus* and  
184 *B. variabilis*. Unlike  $A$  and  $g_s$ ,  $E$  was influenced only by  $k$  and by species and not by seasons.  
185 Only *M. corallina* and *E. erythropappus* differed significantly from each other.

186 The strategy of rapidly absorbing water i.e., high  $k$  favored  $A$ , but decreased the  $E$  with  
187 decreasing of  $g_s$ . However, with the decrease of  $g_s$ , there is also a negative influence on  $A$ . No  
188 distinct behavior was observed between species according to their strategy of foliar absorption  
189 and gas exchange.

190

### 191 **Strategies of FWU and water potential**

192

193 Only one best model remained after model selection, with a  $\Delta AICc = 4$  to the second-  
194 best model. In this model, leaf water potential varied significantly between predawn and  
195 midday and between dry and rainy seasons with and without fog (Fig. 4). The presence of fog  
196 increased the mean values of leaf water potential in all species during the dry season. The  $\Psi_w$   
197 was positively influenced by  $k$  ( $\beta = 5.68$ ) and dry season with fog ( $\beta = 3.01$ ). However, the  
198  $\Psi_w$  was negatively influenced by  $C_{max}$  ( $\beta = -0.39$ ) and dry season without fog ( $\beta = - 8.76$ ).  
199 These results corroborate with the antagonistic traits between  $C_{max}$  and  $k$  parameters. The  
200 presence of fog increased leaf  $\Psi_w$  in both predawn and midday, during the dry season. The

201 fog water absorption generated higher values of leaf  $\Psi_w$  at midday during the dry season than  
202 at the rainy season.

203

## 204 **DISCUSSION**

205

### 206 **Strategies of FWU and gas exchanges**

207

208 The uptake of water by the leaves under different strategies has been proven in the species  
209 studied herein (Boanares et al., 2018b). These different strategies of FWU are a compensation  
210 because once the plant is slow to absorb water through its leaves, it compensates by absorbing  
211 a larger amount of water. In contrast, a plant with limitations in storing water in its tissues can  
212 adjust such disadvantage by absorbing faster, and hydrating its photosynthetic tissues  
213 (Boanares et al., 2018a and b). Currently, it is demonstrated that these strategies interfere in  
214 the water and carbon balance in *in situ* conditions, and the benefits water fog uptake in the dry  
215 season were verified. FWU increases the water potential of plant species, minimizing the  
216 water stress due to low rainfall of this season.

217 The  $k$  parameter, which confers speed to the FWU, negatively influences  $A$  in the dry  
218 season, because it also has negative relation with  $g_s$ . Thus, reduced stomatal conductance  
219 impairs gas exchange, and consequently carbon input due to the limited storage of water in  
220 plant tissues. Therefore, when leaves are less hydrated, stomatal closure reduces the  $g_s$   
221 inducing a decrease in the transpiration rate, which is beneficial for the plant. For the plant  
222 species studied herein, the strategy of fast leaf water absorption through the leaves grants  
223 disadvantages in terms of carbon assimilation, but offers advantages against loss of water in  
224 the dry season. The stomatal closure restricts the damage caused by dehydration, since the  
225 potential for leaf transpiration is much greater than the water supply and / or storage capacity

226 (Medrano et al., 2015). Then, species with higher  $C_{max}$ , such as *E. erythropappus*, may be less  
227 able to control stomatal closure and opening, showing a greater dependence on the continuous  
228 supply of water for its tissues (Sperry et al., 2002).

229 Much wetting next to the leaves through fog or rainfall, provides a decrease in photon flux  
230 density, which can negatively interfere with light absorption due to its dispersion (Egri et al.,  
231 2010). Indeed, this wetting can also negatively interfere in  $CO_2$  assimilation (Burgess and  
232 Dawson, 2004; Eugster et al., 2006). This process occurs because diffusing  $CO_2$  through  
233 water is 10,000 times slower than through air, consequently photosynthetic carbon uptake is  
234 reduced (Brewer and Smith, 1995; Letts and Mulligan, 2005). There is also the possibility of  
235 water entering the stomata and consequently causing a decrease in leaf gas exchanges, since it  
236 has been seen that there is evidence supporting the role of stomata of FWU (Burgess and  
237 Dawson, 2004; Burkhardt, 2010). Stomatal clog up may also occur by water droplets without  
238 the actual absorption of this water, preventing  $CO_2$  entrance (Gerlein-Safdi et al., 2018).  
239 However, the opposite behavior, i.e., leaf absorption varying positively in relation to gas  
240 exchange in different species of different ecosystems may occur (Grammatikopoulos and  
241 Mannetas, 1994; Martin and von Willert, 2008; Simonin et al., 2009; Burkhardt et al., 2012;  
242 Eller et al., 2013; Berry et al., 2014; Rosado et al., 2018). Leaf wetting changes leaf energy  
243 balance, decreasing temperature and consequently stomatal closure (Dawson and Goldsmith  
244 2018; Gerlein-Safdi et al., 2018). Thus, in addition to leaf hydration, there is a decrease in  
245 transpiration. Due to the different availabilities of water and  $CO_2$  resources, improvement of  
246 leaf wetting serves as a support to the limited water soil content (Boucher et al., 1995).

247 The wet soils during the rainy season does not represent a favorable condition for FWU  
248 (Simonin et al., 2009), which explain the non-significant relationship of the rainy season and  
249 FWU. Meteorological events of fog saturating the atmosphere, which reduce the incidence of  
250 photons, and consequently carbon assimilation, is a temporary situation. As soon as plant

251 tissues hydrate and precipitation ceases, it will assimilate carbon again, and water stress will  
252 no longer impair photosynthesis, which means a compensation between CO<sub>2</sub> assimilation and  
253 leaf hydration.

254 The increase of gas exchange may be directly related to the increase of water potential in  
255 these species, which does not imply a decrease in CO<sub>2</sub> assimilation (Eller et al., 2013). During  
256 the rainy season, there is an increase in precipitation frequency, which can provide more  
257 favorable sunlight for photosynthesis, if compared to clear sky conditions. Thus, a reduction  
258 in the risk of photoinhibition from the penetration of diffuse and non-direct light may occur  
259 (Lüttge et al., 2007). The strategies of plant species to repel superficial water, as seen in many  
260 species of plants growing in areas with frequent leaf wetting, may favor the relation of FWU  
261 with gas exchanges (Brewer and Smith, 1995).

262 A decrease in gas exchange in the dry season may be related to the amount of water in leaf  
263 tissues, which is not enough for gas exchanges to occur in total capacity. Therefore, in the dry  
264 season, two of the four plant species studied here, *Byrsonima variabilis* and *Leandra australis*  
265 cannot store enough water to maintain leaf gas exchange. However, *E.*  
266 *erythropappus* presented similar *A* values between dry and rainy seasons, and possibly, this is  
267 the consequence of a large water storage in its tissues (the highest *C*<sub>max</sub>), and therefore this  
268 species will not suffer constraints related to water loss, by keeping its stomata open. Also, the  
269 possible deep root and xylem hydraulic differences in *E. erythropappus* can be related to  
270 maintenance of this physiological process in dry season (Zimmermann, 1983).

271

## 272 **Strategies of FWU and water potential**

273

274 Several studies have demonstrated positive effects of fog on the water balance of some  
275 plant species (Johnson and Smith, 2008; Ritter et al., 2009; Eller et al., 2016). Currently,

276 different strategies of foliar absorption ( $k$  and  $C_{\max}$ ) interfere in the  $\Psi_w$  of the species. Just as  
277 in FWU, there is an antagonistic relationship between  $k$  and  $C_{\max}$ , as is also true for the  $\Psi_w$ .  
278 The  $\Psi_w$  is positively influenced by the  $k$  and negatively by the  $C_{\max}$ . This relationship occurs  
279 because the foliar water rapid absorption ( $k$ ) is directly related to the increase in plant water  
280 status. In contrast, a plant with slow absorption will not change its water status at the same  
281 speed as plants with fast absorption. However, considering the period of the day, the  $C_{\max}$   
282 positively influences the  $\Psi_w$ , and  $k$  negatively influences the the  $\Psi_w$  in predawn. Such  
283 influences may be explained by the strategy of plants with higher  $C_{\max}$  to slowly absorbing  
284 larger amounts of water, losing less water than the plants that absorb fast, keeping their  
285 tissues more hydrated for a longer period of time. Thus, in the early hours of the day, their  $\Psi_w$   
286 is larger than that of the fast-absorbing plants.

287 During the dry season when fog occurs, all the species increased their  $\Psi_w$  in relation to the  
288 dry season when fog was absent. *M. corallina* and *E. erythropappus* ( $C_{\max}$ ) increased their  
289 water status the most, when the midday of the dry season with no fog is compared to the  
290 midday of the dry season with fog. The positive relationship between FWU and gas exchange  
291 in the dry season is a consequence of the positive role of fog in plant physiology (Dawson,  
292 1998; Limm et al., 2009; Eller et al., 2013; Cassana et al., 2015).

293 Fog can alter plant microclimate to a large extent by reducing the vapor pressure deficit,  
294 and consequently decreasing plant transpiration close to zero (Burgess and Dawson, 2004;  
295 Johnson and Smith, 2008; Vesala et al., 2017). Many studies have shown that despite the  
296 potentially negative effects, cloud forest species may benefit from cloud immersion as the  
297 transpiration can increase considerably in the dry season (Burgues and Dawson, 2004;  
298 Johnson and Smith, 2008). Also, the existence of some species in a given environment may  
299 be correlated to their FWU (Limm et al., 2009; Berry et al., 2014). The occurrence of fog in  
300 the ferruginous rupestrian fields confers a favorable water balance for all species studied,

301 especially in the dry season. Thus, it can be observed that the studied species of the Brazilian  
302 ferruginous rupestrian fields do not experience water stress as much as expected formerly,  
303 since fog mitigate the damages caused by water deficit.

304 Current data corroborate the concept of decoupling of shoot physiology from soil water  
305 potential due to the foliar absorption capacity (Simonin et al., 2009). Transpiration and  
306 photosynthesis are independent of the amount of water in the soil, contrary to which was  
307 previously predicted by the unidirectional model of the continuous soil-plant-atmosphere  
308 (SPAC). Additionally, our results represent a complement to the concept of the SPAC model,  
309 with implications to the climatic models, once the strategy of leaf water absorption of these  
310 plants can be dissociated from the soil water potential (Goldsmith, 2013).

311 Although FWU interferes negatively in CO<sub>2</sub> assimilation, this interference is brief and  
312 prevents water loss through transpiration by increasing plant water status in the dry season.  
313 Accordingly, there is a compensation between CO<sub>2</sub> assimilation and foliar hydration, which  
314 is, in fact, beneficial to the plants of this ecosystem.

315 Studies on fog FWU in ferruginous rupestrian fields may provide information on the  
316 functional traits of species more or less adapted to carbon and water flux differences,  
317 considering changes in thermal and hydrological patterns, since global climate changes  
318 models predict a decrease in the fog formation (IPCC, 2013). Because of such changes, it is  
319 expected that the distribution of species and phytophysionomies will be strongly changed in  
320 the landscape and that the same pattern will be found in other OCBILs. Such knowledge can  
321 provide subsidies which will help to understand the dynamics of the community, its operation  
322 and flows.

323

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325

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330

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**Table 1.** Mean values of  $C_{\max}$  and  $k$  ( $\text{g g}^{-1}$  dry weight) in leaves of four species studied in dry and rainy seasons.

Species	Season	$C_{\max}$		Species	Season	$k$	
<i>E. erythropappus</i>	Dry	0.9729	<b>a</b>	<i>L. australis</i>	dry	0.1822	<b>a</b>
<i>M. corallina</i>	Wet	0.8754	<b>ab</b>	<i>B. variabilis</i>	wet	0.1607	<b>a</b>
<i>E. erythropappus</i>	Wet	0.8363	<b>ab</b>	<i>B. variabilis</i>	dry	0.1387	<b>a</b>
<i>M. corallina</i>	Dry	0.6907	<b>b</b>	<i>L. australis</i>	wet	0.1196	<b>a</b>
<i>L. australis</i>	Wet	0.3521	<b>c</b>	<i>E. erythropappus</i>	wet	0.03356	<b>bc</b>
<i>B. variabilis</i>	Wet	0.2173	<b>c</b>	<i>M. corallina</i>	wet	0.03032	<b>bc</b>
<i>B. variabilis</i>	Dry	0.183	<b>c</b>	<i>E. erythropappus</i>	dry	0.02846	<b>bc</b>
<i>L. australis</i>	Dry	0.1805	<b>c</b>	<i>M. corallina</i>	dry	0.0194	<b>c</b>

Different letters indicate a significant difference ( $P \leq 0.05$ ) between the means of the same column.

**Table 2.** Water leaf content ( $C_{\max}$ ) and water uptake speed ( $k$ ) related to the gas exchange parameters and ratios of all four species analyzed that were related to the parameters of foliar water uptake. Net CO<sub>2</sub> assimilation ( $A$ ), transpiration ( $E$ ), stomatal conductance ( $g_s$ ).

Independent variable	Dependent variable	F	DF	Deviance	p-value	
$A$	$C_{\max}$	0.301	1	72.97	0.5845	
	$k$	6.717	1	110.95	0.0108	*
	Species	6.558	3	109.06	0.0004	***
	Season	64.923	1	109.92	0.0001	***
$g_s$	$C_{\max}$	0.091	1	72.97	0.9244	
	$k$	13.126	1	110.95	0.0001	***
	Species	4.628	3	109.062	0.0043	**
	Season	11.135	1	109.92	0.0011	**
$E$	$C_{\max}$	0.006	1	72.97	0.9341	
	$k$	6.276	1	110.97	0.0136	*
	Species	4.029	3	109.06	0.0092	**
	Season	2.94	1	109.92	0.0892	

The above data correspond to the general model. The data of the appropriate minimum model are in the supplementary data. \*\*\* Indicate  $P \leq 0.001$ ; \*\*Indicate  $P \leq 0.01$  and \* Indicates  $P \leq 0.05$ .

**Appendix S1.** Model selection table of the Global model call: lmer (formula = Potential ~ C<sub>max</sub> + k + hour of day + season + C<sub>max</sub>: season + k: season + (1 | Specie/Ind.))

Model	(Int)	C <sub>max</sub>	Season	k	Hour of day	C <sub>max</sub> : season	C <sub>max</sub> : hour of day	Season:k	k: Hour of day	R <sup>2</sup>	df	logLik	AICc	Delta	Weight	
256	-8.75	-0.39	+	5.67	+	+	+	+	+	0.726	15	-	230.531	497.1	0.00	0.742
240	-7.52	-1.84	+	1.23	+		+	+	+	0.724	13	-	235.261	501.0	3.90	0.106
128	-8.05	-1.01	+	1.78	+	+	+	+		0.725	14	-	234.269	501.7	4.66	0.072
224	-11.40	2.75	+	15.23	+	+		+	+	0.714	14	-	234.409	502.0	4.94	0.063
112	-6.81	-2.46	+	-2.66	+		+	+	+	0.723	12	-	238.992	505.7	8.68	0.010
208	-10.17	1.29	+	10.78	+			+	+	0.712	12	-	239.162	506.1	9.02	0.008

496 **Figure legends**

497

498 **FIGURE 1.** Ferruginous rupestrian field (A) without fog and (B) with fog.

499

500 **FIGURE 2.** The average annual rainfall and average annual temperature from “Serra da  
501 Brígida”. Bars represent total precipitation volume (mm) and the line represents monthly  
502 average.

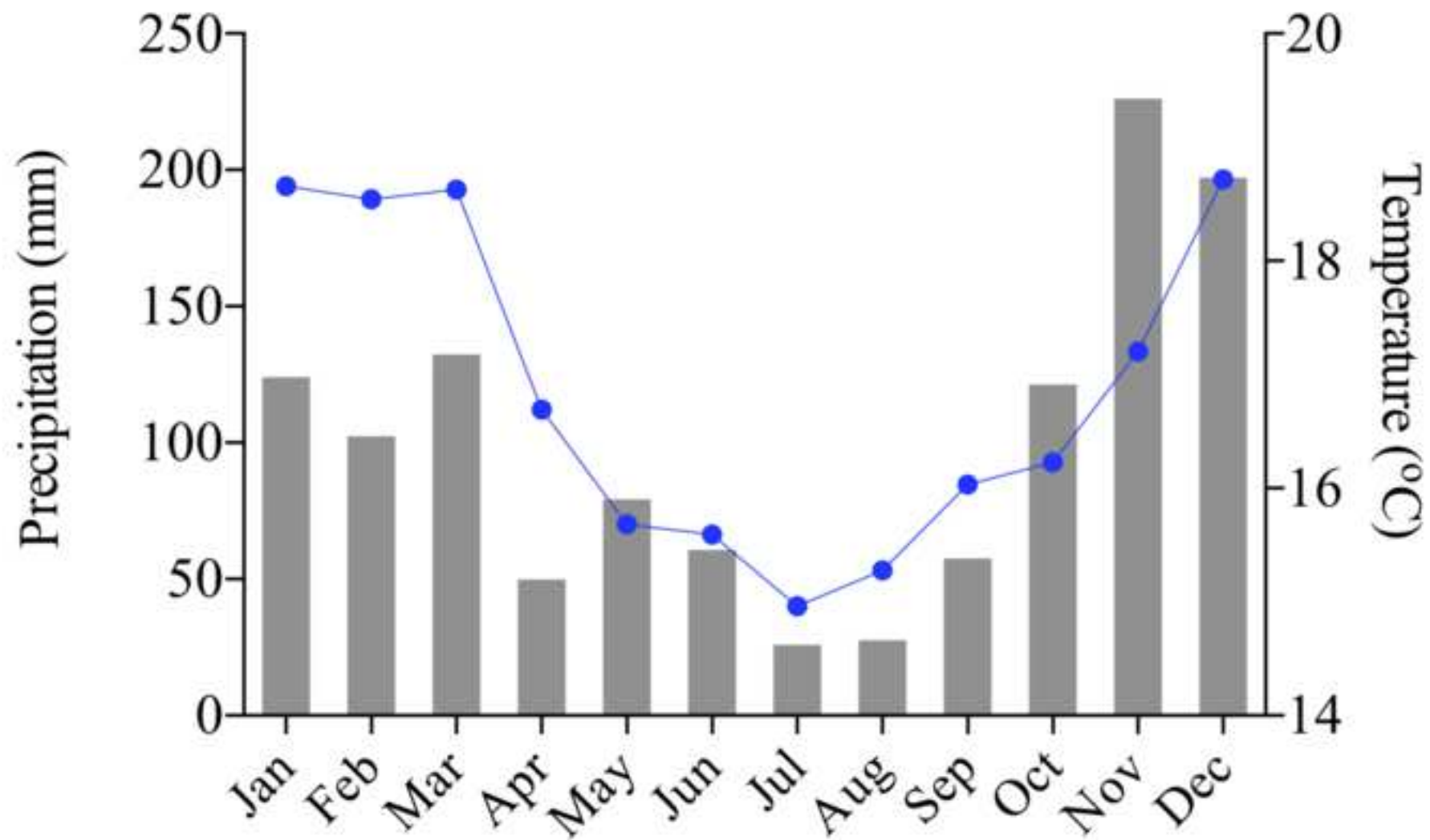
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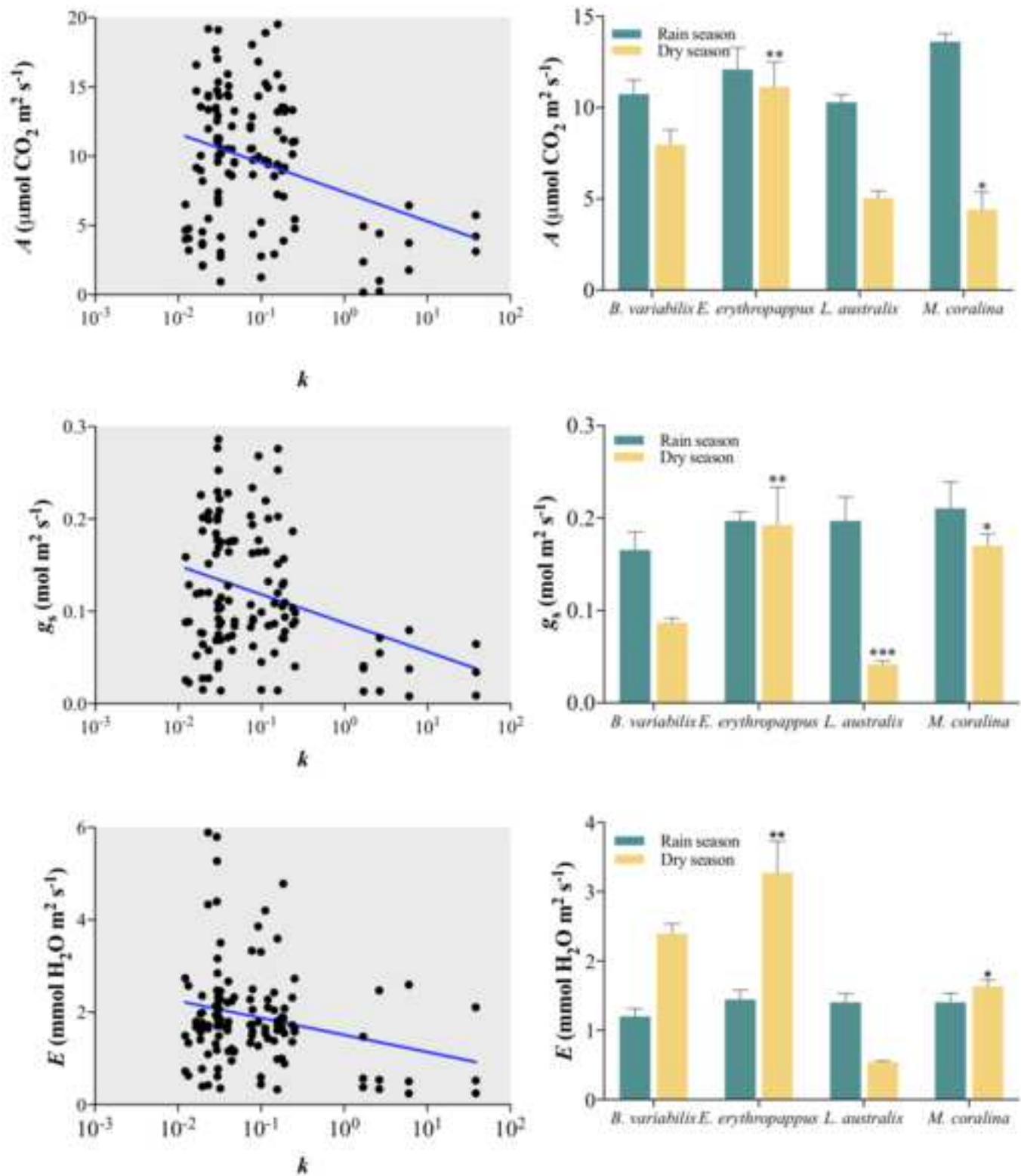
504 **FIGURE 3.** Gas exchanges in leaves of *Eremanthus erythropappus*, *Miconia corallina*,  
505 *Leandra australis* and *Byrsonima variabilis* in the rainy and dry seasons. Parameters that  
506 were significantly inflated by  $C_{\max}$  and  $k$ . Net CO<sub>2</sub> assimilation (A), stomatal conductance (B)  
507 and transpiration (C).

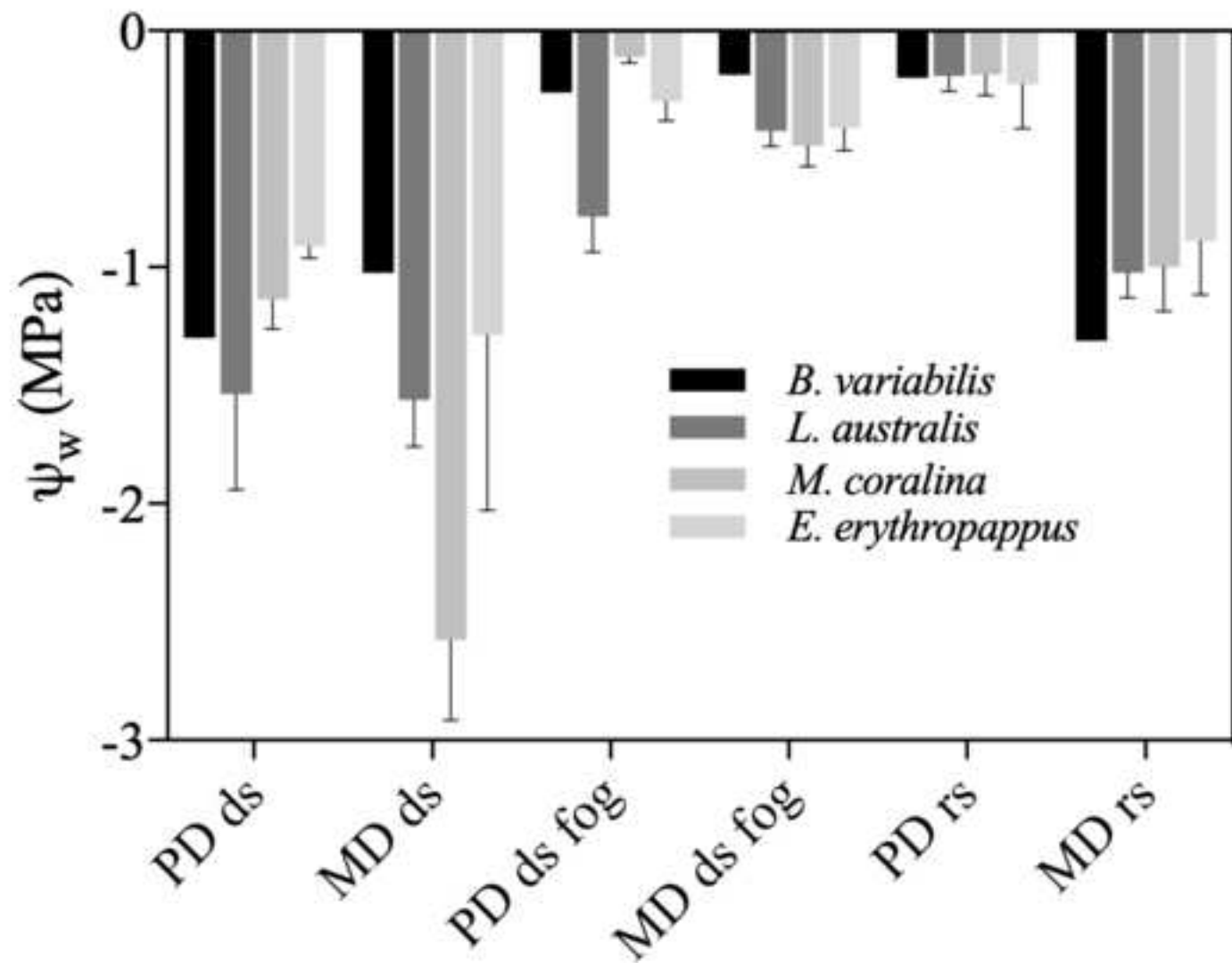
508

509 **FIGURE 4.** Leaf water potential ( $\Psi_w$  in MPa) of the branches of the four species studied.  
510 Period: Predawn dry season without fog (PD d); Predawn dry season with fog (PD d + f),  
511 Predawn rainy season (PD r); Midday dry season without fog (MD d); Midday dry season  
512 with fog (MD d + f); Midday rainy season (MD r).









## Chapter 4

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*Foliar water uptake is an important trait related to thermotolerance responses in plants from ferruginous rupestrian fields*

1 **Photosynthetic heat tolerance in plants with different foliar water uptake**  
2 **strategies**

3  
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15

16 **AUTHORS' CONTRIBUTIONS**

17

18 D.B., J.P.L.F., R.M.S.I. and M.G.C.F. designed the research, D.B. collected the data for the  
19 research, and D.B. analyzed the data and D.B., J.P.L.F., R.M.S.I. and M.G.C.F wrote the  
20 manuscript. All the authors contributed critically to the drafts and gave final approval for  
21 publication.

# 1 **Photosynthetic heat tolerance in plants with different foliar water** 2 **uptake strategies**

3

## 4 **Abstract**

5

- 6 1. The Campo rupestre is an old climatically buffered infertile landscape (OCBIL) and  
7 we have previously characterized in this protected ecosystem different strategies for  
8 foliar water uptake (FWU) as follows: quickly and lower water absorption (QL) or  
9 slower and higher water absorption (SH).
- 10 2. For the first time the relationship between FWU, photoinhibition and  
11 thermotolerance was investigated in seven species from this ecosystem.
- 12 3. The SH species are those that presented less photoinhibition. Contrastingly, the QL  
13 species presented greater thermotolerance when their leaves were totally hydrated.  
14 However, when there is greater leaf dehydration, the most thermotolerant species  
15 were those with SH strategy.
- 16 4. Results indicated that FWU is an important plant trait from this ecosystem to face  
17 excessive light intensity and higher temperatures. These plants may be differentially  
18 affected by future global warming and it was observed that the best strategy to deal  
19 with this expected climate change is the SH strategy.

20

## 21 **KEYWORDS**

22 Brazilian OCBIL, climate change, fog, foliar water uptake, heat tolerance,  
23 photosynthesis

24

## 25 **1 | INTRODUCTION**

26

27 The occurrence of fog is observed in the mountains of Minas Gerais, Brazil due to the  
28 high altitude characteristic of Tropical montane cloud Forests (Jarvis & Mulligan,  
29 2011). This meteorological phenomenon plays an important role in the water balance of  
30 plants from Campo rupestre. In these tropical highland environments, the water balance  
31 is favored by fog episodes. The fog is intercepted by the canopy toward leaf water  
32 absorption, which is also favored by reduced evapotranspiration conditions (Aparecido  
33 et al., 2018). This is the case of the vegetation of the rupestrian fields, which also have

34 shallow soils and rocky outcrops (Silveira et al., 2016), and are frequently subjected to  
35 high light incidence on sunny days. In addition, the high air vapor pressure deficit often  
36 results in restriction of transpiration due to the decrease of stomatal conductance. These  
37 overall conditions result in thermal stress by the increasing leaf temperature as  
38 described for species from the montane Atlantic forest of southeastern Brazil (Fauset et  
39 al., 2017).

40 Temperature is a strong driver of plant species distribution and even survival  
41 (Marias et al., 2016). Likewise, the increase of mean global temperature (IPCC, 2014)  
42 can cause thermal stress and induce a significant constraint for vegetation survival  
43 (Wahid et al., 2007). High temperatures can threaten the survival of sensitive species,  
44 resulting in local extinctions, large scale changes and altered vegetation composition  
45 (Taylor et al., 2019). Unfortunately, global warming may, by the end of this century,  
46 increase the temperature in the tropics resulting in a higher frequency and intensity of  
47 thermal stress in plants, and similarly compromise the hydric regime through the  
48 decrease of precipitations events, such as rain and fog (Corlett, 2011; IPCC, 2014).  
49 Therefore, this heating can change the structure, function, services and earth-  
50 atmosphere interactions in the ecosystems (Sastry & Barua, 2017). However, these  
51 high-temperature impacts on the physiological performance of plants are not fully  
52 understood (Marias et al., 2016).

53 Mountain species grown in the Campo rupestre (Jacobi et al., 2007) are the most  
54 susceptible to global climate change (Colwell et al., 2008). This tropical environment is  
55 considered an OCBIL for it is an old, climatically buffered, infertile landscape (Hopper,  
56 2009). The progressive increase in mean atmospheric temperature will force the species  
57 to move to areas beyond their current distribution (Corlett & Westcott, 2013). However,  
58 such movement is improbable for several species have limited dispersion capacity in  
59 this environment (Colwell et al., 2008; Silveira et al., 2016). In this sense, global  
60 warming is a major threat for the survival of species that already suffer great anthropic  
61 pressure due to iron mining activity (Martinelli & Moraes, 2013).

62 Above a certain thermal limit, temperature increases result in damage to the  
63 photosynthetic apparatus, and a consequent decrease in photosynthetic rates (Berry &  
64 Bjorkman, 1980). The photosystem II (PSII) and the oxygen-evolving complexes are  
65 sensitive to stress factors, such as light excess and extreme temperatures (O'Sullivan et  
66 al., 2016). The temperature limits and the consequence of temperature increasing for  
67 tropical species are poorly known. Tropical species are subjected to interactions with

68 several environmental constraints, and may present variations in the tolerance of the  
69 photosynthetic apparatus to progressive temperature increasing and light excess (Sastry  
70 & Barua, 2017). PSII tolerance to high temperatures can be enhanced by drought  
71 (Havaux, 1992; Godoy et al., 2011). Anyway, an increase in heat tolerance, even in a  
72 few degrees, can be extremely important for plant survival regarding the intensification  
73 of heat waves and drought episodes resulting from predicted climate change.

74 In addition, rupestrian field plants are exposed to high light intensity and under  
75 excess light conditions; non-photochemical thermal dissipation provides an energy  
76 imbalance between light absorption and utilization, minimizing the possibility of  
77 photooxidative damage (Demmig-Adams & Adams, 2006). The damage to the PSII  
78 under excessive light has been reported for some species in the rupestrian fields (Luttge  
79 et al., 2007; Morales et al., 2014; Castro et al., 2016).

80 Foliar water uptake (FWU) has been studied in the last decades (Berry et al., 2019)  
81 and this physiological process is characterized by the water entrance, whether in its  
82 liquid form or vapor through the leaf surface (Burgess & Dawson, 2004). The  
83 ecological role of this small but significant water intake has been studied (Berry et al.,  
84 2019). The FWU can improve the hydric status of the plant through increase of leaf  
85 water potential, decrease stomatal conductance and improvement of the photosynthesis  
86 capacity and water use efficiency (Burgess & Dawson, 2004; Eller et al., 2016; Vesala  
87 et al., 2017; Berry et al., 2019). It was observed that 11 species of the ferruginous  
88 rupestrian field have FWU capacity, but with different strategies of water influx. Some  
89 species absorb water rapidly in smaller amounts and other species absorb water slowly  
90 in greater amounts (Boanares et al., 2018 a, b).

91 Hypothetically, PSII heat tolerance can differ among the species with different FWU  
92 strategies, due to the increase in the water content in leaves, which allows a greater loss  
93 of water through transpiration providing the cooling of these leaves. According to the  
94 theory OCBIL, past climate stability in the Campo rupestre may have favored the  
95 existence of several areas of endemism and refuge (Collevatti et al., 2009; Bonatelli et  
96 al., 2014; Barbosa et al., 2015) and the temperature increase will represent a great  
97 challenge for the species of this environment. Therefore, understanding the relationship  
98 between FWU and thermotolerance and high light intensity is essential to assess the  
99 vulnerability of these species in the global climate change scenario (Wang et al., 2012;  
100 Sastry & Barua, 2017) and this is not yet investigated.

101 We investigate if species with differential FWU strategies also have differential PSII  
102 tolerance to heat, and if they respond differently to excessive light. Thus, we aim to  
103 verify if there is a better FWU strategy related to the increase of the mean temperatures  
104 expected due to climate change for studied species. We believe that species, which  
105 absorb a greater amount of water but slowly, present greater photosynthetic efficiency  
106 and thermotolerance than those species with rapid absorption, but with lesser amount of  
107 water.

108

## 109 2 | MATERIALS AND METHODS

110

### 111 2.1 | Plant material

112

113 This study was carried out in a Campo rupestre, a Brazilian altitudinal ecosystem (20°  
114 21'30"S/43°30'11"W). The area has an average altitude of 1,492 m a.s.l., and fog is  
115 frequent throughout the year both during day night. Local vegetation is present on  
116 shallow soils on rock outcrops of quartzite, sandstone or ironstone with iron formation  
117 such as itabirites and *cangas* and is characterized by herbs, shrubs and treelets (Valim et  
118 al., 2013). Data were obtained in March 2019, duration the transition from rainy to dry  
119 season, and the average annual temperature was 16.9°C with a total annual rainfall of  
120 1,204.7 mm.

121 The studied species were: *Leandra australis* (Cham.) Cogn. (Shrub), *Trembleya*  
122 *laniflora* (D. Don) Cogn. (Shrub), *Pleroma heteromallum* (D. Don) D. Don. (Shrub),  
123 *Miconia corallina* Spring. (tree) (Melastomataceae), *Eremanthus erythropappus* (DC.)  
124 MacLeish, (Asteraceae) (tree), *Byrsonima variabilis* A. Juss., (Malpighiaceae) (shrub)  
125 and *Ocotea pulchella* (Nees and Mart.) Mez, (Lauraceae) (tree). All species are native  
126 and endemic to Brazil, except *Leandra australis* and *Ocotea pulchella* that were not  
127 endemic to Brazil. *T. laniflora* is the species with the most restricted distribution,  
128 followed by *B. variabilis*. Both occur only in the Campo rupestre. Contrastingly,  
129 *Pleroma heteromallum*, *Eremanthus erythropappus* and *Ocotea pulchella* have the  
130 widest distribution among the species studied. These species were selected according to  
131 their different water absorption capacities (Boanares et al., 2018b). *Leandra australis*,  
132 *Ocotea pulchella* and *Byrsonima variabilis* have a higher speed of leaf water absorption,  
133 but uptake a small amount of water (Figure 1).

134

## 135 2.2 | Foliar water uptake measurement

136

137 To evaluate the different capacities of FWU, the following procedure was performed:  
138 first, mature and intact leaves ( $n = 4$ ) of 5 individuals were collected in the field, and in  
139 laboratory conditions the leaves were immersed in distilled water and weighed during 4  
140 h, every 15 min for 2 h. Then every 30 min for 2 h and 1 h to finish the experiment to  
141 obtain the maximum water absorption curve. Before each weighing, leaves were dried  
142 with ultra-absorbent microfiber synthetic towels to remove the surface water (Liang et  
143 al., 2009). Leaf water uptake was calculated by a differential equation, which provides  
144 data on the maximum water content in leaf lamina ( $C_{\max}$ ) and leaf uptake speed ( $k$ )  
145 (Liang et al., 2009). From a statistical analysis two opposite strategies were observed:  
146 species with higher  $C_{\max}$  presented lower  $k$  and species with low  $C_{\max}$  presented higher  $k$   
147 (Boanares et al., 2018a, b).

148

## 149 2.3 | Leaf dehydration measurement

150

151 To assess the leaf dehydration capacity of these species, mature leaves ( $n = 4$ ) of 5  
152 individuals were totally submerged in distilled water, room temperature, for 4 h to full  
153 hydration after the petiole were sealed. These leaves were dried with ultra-absorbent  
154 microfiber synthetic towels to remove the surface water (Liang et al., 2009), weighed  
155 with an electronic scale, every 30 min in the first 6 h, and then they were weighed two  
156 more times after 12 h and 24 h. The leaves were kept in the light throughout the  
157 experiment. The total amount of water leaf lost ( $C_{\max}^L$ ) and water leaf lost speed ( $k^L$ )  
158 were defined from a differential equation according to Liang et al. (2009).

159

## 160 2.4 | Photosynthetic efficiency evaluation

161

162 In order to observe the occurrence of photoinhibition, measurements of the  
163 photosynthetic efficiency were performed three times a day, at 9 am, 3 pm and 6 pm,  
164 using a modulated fluorescence meter (MINI-PAM Waltz, Germany) in leaves of 5  
165 individuals in the field. The average temperature ranged from 18° C to 32.5°C, the  
166 humidity was 36% to 83%, and the maximum radiation was 3101 kJ/m<sup>2</sup>. After  
167 adaptation of leaf lamina to the dark for 30 min the potential quantum yield ( $Fv/Fm$ ) of

168 photosystem II (PSII) was evaluated (Rascher et al., 2008). Subsequently, to calculate  
169 the  $F_v/F_m$  ( $F_v = F_m - F_0$ ), the  $F_v$  (variable fluorescence) was measured in leaves adapted  
170 to the dark, using leaf clips provided by the manufacturer, and  $F_m$  (maximum  
171 fluorescence) was measured in leaves adapted to the dark after a pulse of saturating  
172 light, and  $F_0$  is the basal fluorescence produced by leaves adapted to the dark.

173

## 174 **2.5 | Photosystem II thermotolerance assessment**

175

176 Leaf disks (2 cm<sup>2</sup>) from 4 leaves from each species were collected at 9 am and also at 1  
177 pm, totalling 10 discs per species for each sampling time, were used to evaluate heat  
178 tolerance immediately after the sampling. The thermotolerance assessment was  
179 conducted using a thermostatic bath. The leaf discs were placed on a thermally  
180 controlled plate, and covered with plastic film to avoid contact of the discs with water  
181 of thermostatic bath. To evaluate thermotolerance at different levels of leaf hydration,  
182 measurements were made on new obtained disks with 100%, 80% and 60% of the full  
183 foliar hydration. Leaf discs were kept on a workbench in order to obtain the hydration  
184 levels of 60% and 80%, weighted on analytical scale and considering the fully hydrated  
185 disc weights. Leaf discs were subjected to a progressive temperature increase from 25°C  
186 to 65°C, with gradual increases of 2°C every 3 min in the dark. The values of  $F_v/F_m$   
187 were obtained in intervals of 2°C. The  $F_v/F_m$  decay temperatures at 50% ( $T_{50}$ ) values  
188 were calculated from a sigmoidal equation adjustment between the  $F_v/F_m$  data at each  
189 temperature increase (Chaves et al., 2015).

190

## 191 **2.6 | Data analysis**

192

193 General linear models (GLMs) with gaussian errors were built to evaluate if the  
194 maximum water content in leaves, the water uptake speed and time of the day (9 am or  
195 3 pm) (independent variables) influenced the potential quantum yield, and  $T_{50}$   
196 (dependent variables) of the seven species. GLM were built to evaluate if there was a  
197 relation between the water parameter efflux ( $C_{\max}^L$  and  $k^L$ ) with thermotolerance data  
198 ( $T_{50}$ ) and FWU parameters ( $C_{\max}$  and  $k$ ). To verify if different levels of hydration  
199 interfere in thermotolerance, a GLM was built, including levels of hydration  
200 (independent variable) and  $T_{50}$  (dependent variable). All analyses were performed using  
201 the R platform (R Core Team, 2018).

202

203 **3 | RESULTS**

204

205 **3.1 | Leaf dehydration measurements**

206

207 There was a negative relation between the amount of water lost by the leaves ( $C_{\max}^L$ )  
208 and the foliar water absorption speed ( $k$ ). The species with higher values of  $k$  are those  
209 that lose less water ( $C_{\max}^L$ ). Contrastingly, species that absorb a higher amount of water  
210 ( $C_{\max}$ ) lose water by the leaves more quickly ( $k^L$ ) (Figure 2). The foliar water loss  
211 parameters influenced thermotolerance responses of all studied species. The most  
212 thermotolerant species were those that lost water more slowly and in smaller quantity,  
213 i.e., lower values of  $C_{\max}^L$  and  $k^L$ . These species, *L. australis*, *O. pulchella* and *B.*  
214 *variabilis*, were represented by the lower values of  $k^L$  (Table 1). However, these species  
215 that absorb water faster (high  $k$ ) and lose less water through their leaves (low  $C_{\max}^L$ ) are  
216 not the most resistant to dehydration. That is, with the decrease of their foliar hydration,  
217 these species reduced the thermotolerance responses.

218

219 **3.2 | Photoinhibition evaluation**

220

221 There was a significant relation between  $C_{\max}$  and time of day with  $Fv/Fm$  values and  
222 species ( $F_{1,82}=22.7$ ,  $F_{2,79}=14.6$  and  $F_{6,73}=8,06$ , respectively;  $P\leq 0.001$ ). The species with  
223 higher values of  $Fv/Fm$  were those that have the strategy of higher leaf water absorption  
224 and lower water absorption speed (high  $C_{\max}$  and low  $k$ ): *E. erythropappus*, *T. laniflora*  
225 and *P. heteromallum*. Contrastingly, the species with lower  $Fv/Fm$  values, *L. australis*  
226 and *B. variabilis*, presented opposite strategies, that is low  $C_{\max}$  and high  $k$ . However,  
227 this second group of species includes *M. corallina*, which presents the FWU strategy  
228 similar to that of the first group of species (Figure 3). Importantly, *O. pulchella* that has  
229 the lowest value of  $C_{\max}$ , also showed the lowest value of  $Fv/Fm$ . The species in which  
230 photoinhibition was detected ( $Fv/Fm$  values lower than 0.8) are those related to the  
231 strategy of higher speed ( $k$ ) and a smaller amount of water absorbed ( $C_{\max}$ ) (Figure 3).

232 Regarding the different times of the day when  $Fv/Fm$  was measured, at 3 pm the  
233 lowest values of  $Fv/Fm$  were observed and, consequently, photoinhibition occurred, at  
234 least in RL species. On the other hand, the highest  $Fv/Fm$  values occurred at 6 pm,

235 when there was solar radiation and lower temperature - environmental conditions which  
236 enable the occurrence of PSII recovery - in fact,  $F_v/F_m$  values at 9 am showed  
237 intermediate values.

238

### 239 **3.3 | PSII thermotolerance assessment**

240

241 The two parameters used to estimate FWU ( $C_{max}$  and  $k$ ) and plant species were  
242 significantly related to  $T_{50}$  ( $F_{1,33}=21.04$ ;  $P\leq 0.001$ ,  $F_{1,32}= 4.13$ ;  $P\leq 0.05$  and  $F_{6,26}=5.03$ ;  
243  $P\leq 0.01$ , respectively). Species with higher values of  $T_{50}$ , that is, the most thermotolerant  
244 ones, were the species with higher foliar absorption speed (high  $k$ ) and lower values of  
245  $C_{max}$ , i.e, *B. variabilis*, *L. australis* and *O. pulchella* obtained at 46°C. The parameters  
246 with opposite traits related to FWU distinguished the species according to the  
247 thermotolerance capacity. The less tolerant species to temperature increase were those  
248 that had the strategy to absorb water slowly by the leaves but in greater quantity: *E.*  
249 *erythropappus*, *T. laniflora*, *M. corallina* and *P. heteromallum* obtained at 44°C (high  
250  $C_{max}$ ) (Figure 4).

251 Considering leaves hydration levels, there was a significant relation between the  
252 hydration levels ( $F_{2,207}=32.48$ ;  $P\leq 0.001$ ) and the different species ( $F_{6,201}=3.81$ ;  $P\leq 0.01$ )  
253 in relation to  $T_{50}$  values (Figure 4). A contrary standard for thermotolerance results was  
254 observed, i.e., the most thermotolerant species to the decrease of dehydration were the  
255 species that have the strategy of absorbing a larger quantity of water but more slowly  
256 (high  $C_{max}$  and low  $k$ ): *M. corallina*, *T. laniflora* and *P. heteromallum* obtained at 45°C.  
257 The less thermotolerant species with the increase of dehydration were those of the  
258 opposite strategy with high  $k$  and low  $C_{max}$ : *L. australis*, *O. pulchella* and *B. variabilis*  
259 obtained at 43°C, except *E. erythropappus* that despite presenting high  $C_{max}$  remained in  
260 this group (Figure 5).

261

## 262 **4 | DISCUSSION**

263

### 264 **4.1 | Water influx and efflux in leaves**

265

266 The different FWU traits influenced the photosynthetic efficiency and thermotolerance  
267 of the species studied. Although the relation between leaf hydration levels with

268 photosynthesis and thermotolerance was expected, the relation between leaf hydration  
269 from FWU with these parameters was not known. An observed pattern was that the  
270 species that absorb the most amount of water through the leaves (high  $C_{\max}$ ), but in a  
271 slower speed (low  $k$ ) lose water more quickly (high  $k^L$ ). Contrastingly, species that  
272 absorb less water (low  $C_{\max}$ ) more quickly (high  $k$ ) lose less water (low  $C_{\max}^L$ ). These  
273 different water losses strategies are the cause of a lower value of relative water content  
274 (RWC) for the species with high  $C_{\max}$  and high  $k^L$ , and higher values of RWC for  
275 species with high  $k$  and low  $C_{\max}^L$  (Boanares et al., 2018b). However, the leaf  
276 ultrastructural and anatomical traits that grant these distinct FWU behaviors are not the  
277 same for water loss (Riederer & Schreiber, 2001). The cell wall composition with  
278 different proportions of cellulose and pectins that allow these different FWU strategies  
279 (Boanares et al., 2018b) does not influence the different water exit trait. Moreover, the  
280 total amount of water absorbed by the leaf ( $C_{\max}$ ) should also depend on a different set  
281 of traits, such as the leaf hydraulic capacitance (McCulloh et al., 2014; Berry et al.,  
282 2019).

283       Probably there is a compensation mechanism between FWU traits and water efflux.  
284 Species that absorb more water lose water faster, consequently, these leaf species need  
285 to absorb a greater amount of water. However, leaves that absorb water quickly cannot  
286 absorb a large amount of water due to their anatomical traits. Their anatomical structure  
287 is one of the causes of this restriction because their leaves have less space for water  
288 storage (Boanares et al., 2018a). On the other hand, they lose less water, compensating  
289 the limitation to absorb a greater amount of water (low  $C_{\max}$ ). However, even though an  
290 individual leaf can eventually stop absorbing water because its internal water storage  
291 capacity is saturated, in a whole-plant context this water would have been redistributed  
292 to the other tissues, and the leaf would be able to keep absorbing water (Burgess &  
293 Dawson, 2004; Eller et al., 2013).

294

#### 295 **4.2 | Photosynthetic efficiency**

296

297 In turn, the species with the high  $C_{\max}$  trait are more efficient photosynthetically than  
298 the species that absorb water faster (high  $k$ ), due to higher values of water potential  
299 induced by fog (Boanares et al., 2019). Moreover, the late afternoon recovery of  $F_v/F_m$   
300 values and the absence of photoinhibition are indicative of the effectiveness of  
301 mechanisms that dissipate the excess light of these plants (Castro et al., 2016). These

302 species have anatomical traits, which offer more possibilities of water storage in their  
303 leaves, and this may protect the photosynthetic apparatus, since a higher amount of  
304 water plays a key role in this process. The species with rapid absorption (higher  $k$ ) and  
305 low amount water absorption (low  $C_{\max}$ ) present anatomical structure that is one of the  
306 causes of this restriction, because their leaves have less space for water storage  
307 (Boanares et al., 2018a). On the other hand, they lose less water, compensating the  
308 limitation to absorb a greater amount of water (low  $C_{\max}$ ). Moreover, this opposite  
309 pattern interfered in the physiological parameters analyzed. The species that absorb  
310 rapidly and lose little water are not efficient enough photosynthetically, since  
311 photoinhibition was observed during a more intense irradiation period of the day. The  
312 lower values of  $\text{CO}_2$  assimilation of the species with lower  $k$  corroborate these results  
313 (Boanares et al., 2019). This behavior occurs because the  $\text{CO}_2$  suppression fixation may  
314 increase the extent of PSII photoinhibition, inhibiting the synthesis of repair proteins  
315 (Murata et al., 2007). The different strategies of water absorption are closely linked to  
316 the strategies of drought tolerance of these species. The slow absorption species can be  
317 closely related to the desiccation tolerance strategy, protecting the proteins linked to the  
318 photosynthesis (Challabathula et al., 2018). Contrastingly, species with higher speed  
319 absorption may be related to the strategy of drought avoidance, reducing the possibility  
320 of dehydration of their tissues, because they cannot deal with low leaf hydration.

321

### 322 **4.3 | Heat tolerance**

323

324 Water contributes to higher heat dissipation by its higher specific heat capacity, granting  
325 a greater leaf thermotolerance (Fauset et al., 2018), therefore attenuating the daily  
326 thermal gradient and the highest irradiance flux. A greater amount of water in leaves  
327 may favor greater transpiration and therefore heat dissipation. In addition, there is a  
328 variation in the thermotolerance degree because of the different gene expressions of the  
329 proteins related to thermotolerance (Liao et al., 2016). The difference in this protein  
330 expression may be directly related to the water amount in leaves (Bechtold et al., 2013).  
331 The species with high  $C_{\max}$  are not as thermotolerant as the other species with high  $k$ ,  
332 when their leaves are kept fully hydrated. This relation is plausible since species with  
333 faster FWU benefit from total hydration in a more efficient way to face high  
334 temperatures. In fact, the species with higher  $k$  lose less water to the environment and  
335 induce higher leaf hydration levels although they have a slower water uptake. There are

336 several repair mechanisms to PSII, such as cyclic electron flux around PSI, xanthophyll  
337 cycle, photorespiratory pathway, ROS elimination systems and thermal dissipation of  
338 absorbed light energy (Szymanska et al., 2017). The species with high  $C_{max}$  are not as  
339 thermotolerant as the species with high  $k$  when their leaves are fully hydrated. So, it is  
340 probable that these species prevent or reduce the chances of photoinhibition through a  
341 more efficient system for thermal dissipation. Thus, it is possible that even large  
342 increases both in the internal repair temperature and in the external temperature can  
343 occur due to high light intensity, which is beyond the levels these species usually  
344 encounter.

345 Although the species with high  $k$  are more thermotolerant, this advantage is limited,  
346 once it decreases with the increasing leaf dehydration. Plants under water deficit have  
347 the stomatal opening reduced to maintain more internal water and, consequently, there  
348 may occur an increase in leaf temperature (Sharkey, 2005). It is possible that these  
349 species will not be able to metabolic rearrange to the extent of maintaining their  
350 thermotolerance with a low amount of water in leaf tissues. This rearrangement would  
351 help in tolerating the lower thermotolerance level of these species, when they are less  
352 hydrated.

353 Thermal stress causes loss of plant water, severely damaging membrane structure  
354 and function (Tiwari et al., 2018). However, species whose strategy is high  $C_{max}$  and  
355 high  $k^L$  present a higher tolerance to water deficit, since these species had the highest  
356 values of  $T_{50}$  in the less hydrated leaves. It is known that plant thermotolerance is  
357 influenced by exposure to higher temperatures, reduced water availability and high light  
358 intensities increasing heat tolerance (Havaux et al., 1992; Sastry & Barua, 2017). One of  
359 the explanations would be a rapid induction of proteins sensitive to dehydration (Sastry  
360 & Barua, 2017). The expression of these proteins as well as the chaperones are directly  
361 related to better tolerance against other abiotic stresses such as thermal stress (Liu et al.,  
362 1998; Sakuma et al., 2002) which prevents protein denaturation and protects organisms  
363 under thermal stress (Parsell & Lindquist, 1993). Furthermore, when plants suffer from  
364 heat stress there is the activation of several proteins related to dehydration generating  
365 defense mechanisms. These proteins play a crucial role in regulating gene expression in  
366 response to this constraint (Takahashi & Badger, 2011). In this way, these species  
367 manage to cope better with lower water content in their foliar tissues being better  
368 adapted because they lose water more quickly. Therefore, these species must protect

369 themselves from the inherent water deficit in their physiological traits of water loss and,  
370 consequently, are more adapted to a lesser leaf hydration.

371 However, it was verified in this work that the different strategies of FWU have no  
372 relation with the degree of distribution (more or less restricted) of the species. An  
373 example are the species *T. laniflora* and *B. variabilis*, both of which are restricted to the  
374 rock field environment and present distinct ecophysiological traits. The same occurred  
375 with other species in this work, that is, species with different FWU strategies coexisting,  
376 corroborating the consensus that FWU is not an exception, but a rule for the species,  
377 differentiating only in its magnitude (Berry et al., 2019).

378

## 379 5 | CONCLUSIONS

380

381 This study indicates that native species may be vulnerable and affected differently by  
382 future global warming. Considering this climate scenario, among the different strategies  
383 of FWU and thermotolerance, it is noticed that the best strategy to deal with this  
384 expected climate change is the one that grants higher  $C_{max}$  to plants. These species with  
385 higher  $C_{max}$  will be able to better perform photosynthetically, when they experience  
386 water deficit due to rainfall decrease. These strategies will help finding out species more  
387 adapted to the future climate scenarios, and consequently indicated for revegetation of  
388 this ecosystem, which is already impacted, and it will also help to predict the possible  
389 changes in the landscape and which species will suffer the most.

390

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## Figure captions

**FIGURE 1** Relationship between water absorption speed ( $k$ ) and maximum water content ( $C_{\max}$ )  $P \leq 0.05$  (a) and the  $C_{\max}$  values among species (b). In black are the species that present slower and higher water absorption and in gray are the species that present quickly and lower water absorption. Data represent mean values ( $\pm$ SE). Different letters indicate differences between species ( $P \leq 0.05$ ).

**FIGURE 2** Maximum water loss ( $C_{\max}^L$ ) and Water speed loss ( $k^L$ ). In black are the species that present quickly and lower water absorption and in gray are the species that present slower and higher water absorption. Different letters indicate differences between species ( $P \leq 0.05$ ).

**FIGURE 3** Potential quantum yield ( $F_v/F_m$ ) measured after 30 min of dark adaptation from leaves of the different species observed at three different times of the day: 9 am, 3 pm and 6 pm. Different letters indicate differences between species at the same time of day at  $P < 0.05$ . Bars represent  $\pm$  SE.

**FIGURE 4** Relationship between the temperature of  $F_v/F_m$  50% decline ( $T_{50}$ ) with (a) water loss leaf content ( $C_{\max}^L$ ) ( $\text{g g}^{-1}$  dry weight) and (b) loss water speed ( $k^L$ ). Linear regressions at  $P < 0.05$ .

**FIGURE 5** Thermotolerance in different species and different leaf hydration levels. Error bars represent  $\pm$  SE. Thermotolerance measured as decay temperatures at 50% ( $T_{50}$ ) of PSII.

**TABLE 1** Water loss leaf content ( $C_{\max}^L$ ) ( $\text{g g}^{-1}$  dry weight), loss water speed ( $k^L$ ) and temperature ( $^{\circ}\text{C}$ ) drop 50% ( $T_{50}$ ) related to the FWU parameters of all seven species analyzed. Water leaf content ( $C_{\max}$ ) and water uptake speed ( $k$ ).

Independent variable	Dependent variable	F	DF	Deviance	$p$ -value	
$C_{\max}^L$	$C_{\max}$	1.1983	1	0.178	0.28809	
	$k$	7.7568	1	1.153	0.01222	*
	$k^L$	0.872	1	5.8696	0.02618	*
	Species	39.493	6	109.92	0.0001	***
$k^L$	$C_{\max}^L$	1.5374	1	0.0001	0.2309	
	$C_{\max}$	26.852	1	0.002	0.0001	***
	$k$	0.5504	1	0.0004	0.4677	
	Species	13.369	6	0.0002	0.0001	***
$T_{50}$	$C_{\max}^L$	34.873	1	23.481	0.0001	***
	$k^L$	33.373	1	22.471	0.0001	***
	Species	4.3844	6	17.713	0.006	**

Data correspond to the statistical analysis of linear general model (GLM). \*\*\* Indicate  $P \leq 0.001$ ; \*\* Indicate  $P \leq 0.01$  and \* Indicates  $P \leq 0.05$ .

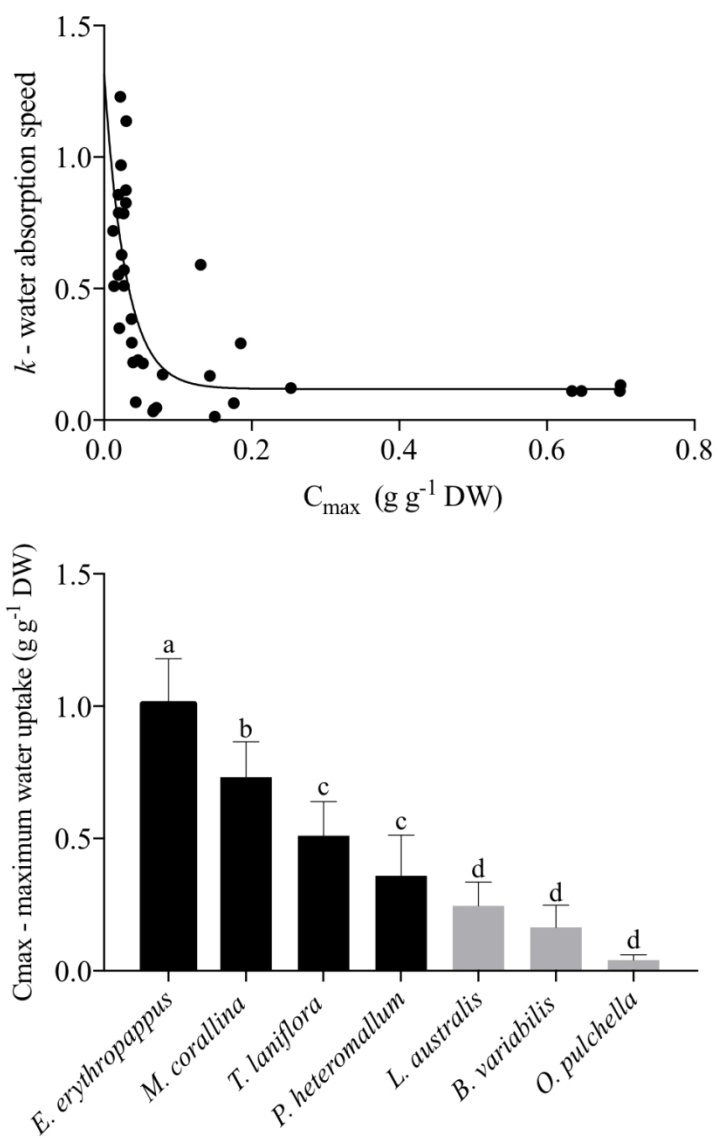


Figure 1

92x135mm (300 x 300 DPI)

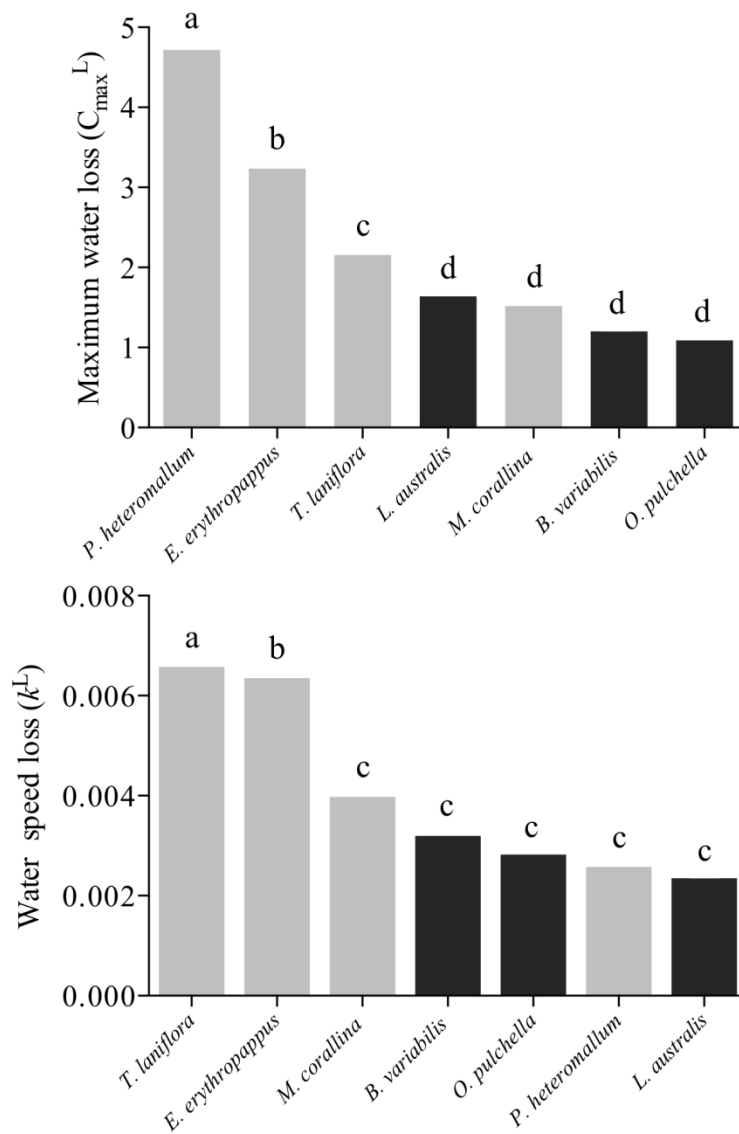


Figure 2

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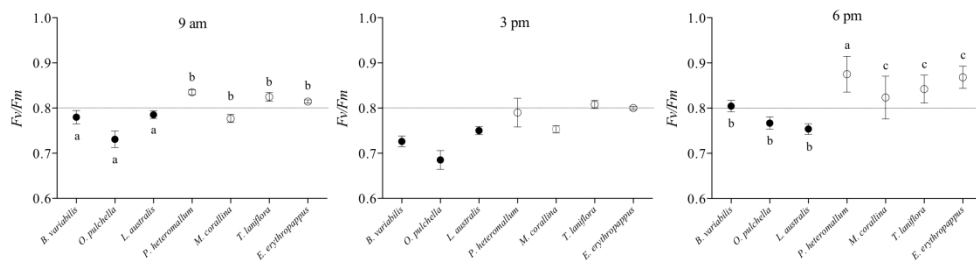


Figure 3

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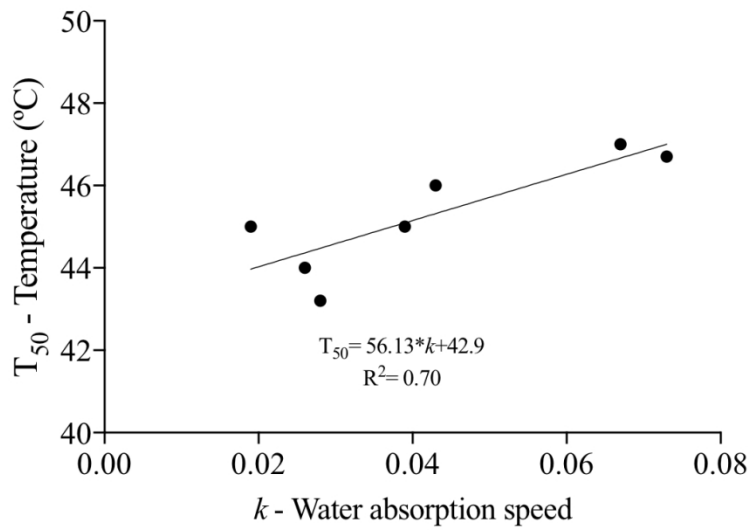
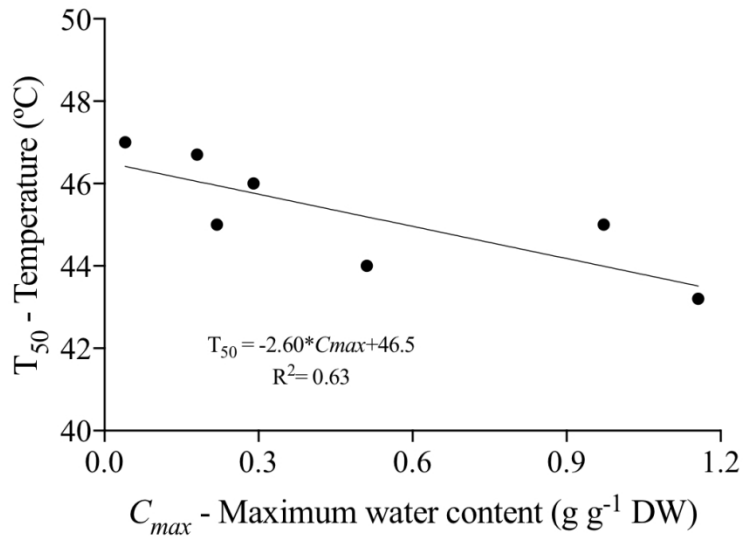


Figure 4

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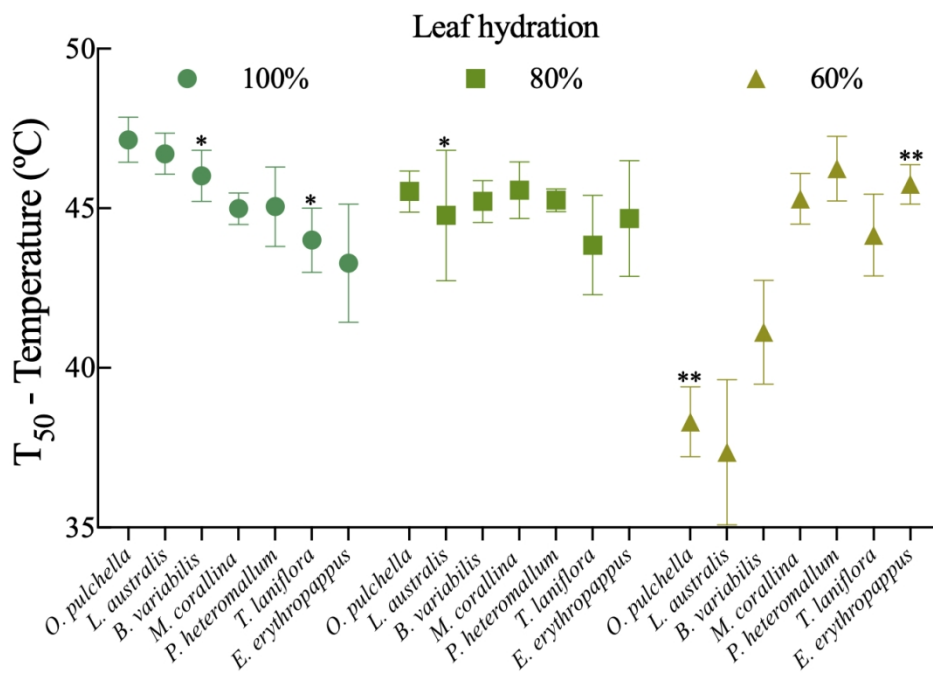


Figure 5

120x88mm (300 x 300 DPI)

*Oxidative metabolism in plants from  
Brazilian rupestrian fields and its  
relation with foliar water uptake in dry  
and rainy seasons*



1 Oxidative metabolism in plants from Brazilian rupestrian fields and its  
2 relation with foliar water uptake in dry and rainy seasons

3  
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12  
13 A B S T R A C T

14 In an altitudinal Brazilian ecosystem, fog is a frequent event in both the dry and rainy seasons. The drought  
15 stress is aggravated in the dry season due to elevated evaporative demand and this constraint can induce  
16 oxidative stress in plants. In this ecosystem, there are plants which present different foliar water uptake (FWU)  
17 capacities - species that absorb less water more quickly (LQ) and those that absorb more water more slowly  
18 (MS). In this study, the relationship between oxidative stress responses and the different FWU strategies was  
19 evaluated in dry and rainy seasons. The oxidative stress was assessed by H<sub>2</sub>O<sub>2</sub> production and lipid peroxidation  
20 as well as by the antioxidant enzymes system as superoxide dismutase (SOD), catalase (CAT) and ascorbate  
21 peroxidase (APX). During the dry season, plants had higher oxidative stress compared to rainy season plants  
22 which exhibited moderate oxidative damage. The FWU strategies were closely related to oxidative stress  
23 responses, since the LQ species presented the higher H<sub>2</sub>O<sub>2</sub> content and oxidative defense system. Contrastingly,  
24 it was found that MS species have the lowest values of H<sub>2</sub>O<sub>2</sub> and less SOD, CAT and APX activities. However,  
25 the lipid peroxidation did not present any relation with FWU strategies. Altogether results revealed that plants,  
26 which present MS strategy, are more adapted to cope with the higher H<sub>2</sub>O<sub>2</sub> concentrations generated in the dry  
27 season and drought stress events than those that present LQ strategy.

29 *Keywords:*

30 Adaptive strategies

31 Drought stress

32 Reactive oxygen species

33 Cell signaling

34

## 35 **1. Introduction**

36

37 Because they are sessile, plants have developed several strategies to cope with the challenges (Mignolet-  
38 Spruyt et al., 2016) imposed by various biotic and abiotic constraints (Anjum et al., 2011). These different  
39 stresses are minimized by adaptive strategies that depend on the type, severity, and combination of  
40 environmental traits, and require specific and flexible combinations of signaling components to trigger  
41 adaptation and acclimatization responses (Suzuki et al., 2012, 2014). The reactive oxygen species (ROS), such as  
42 hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), superoxide anion (O<sub>2</sub><sup>•-</sup>), the oxygen singlet (1O<sub>2</sub>) and hydroxyl radical (HO<sup>•</sup>), which  
43 are formed mainly in chloroplast and mitochondria during the cellular oxidative process, are involved in the  
44 beginning and signaling regulation of the oxidative events (Baxter et al., 2014).

45 The excessive production of ROS can profoundly affect plant metabolism due to damage to cell membranes,  
46 proteins, and genetic material (Foyer and Noctor, 2013). To minimize oxidative stress, plants possess complex  
47 non-enzymatic and enzymatic antioxidant systems, that include superoxide dismutase (SOD), ascorbate  
48 peroxidase (APX) and catalase (CAT) among others (Apel and Hirt, 2004). The cell signaling or oxidative  
49 damages occurrence is determined by balance between ROS and antioxidative enzyme activities (Moller et al.,  
50 2007). Previously, ROS were considered only negative byproducts for optimal cellular metabolism.  
51 Nevertheless, the role of these molecules has been better understood and has emerged as essential messengers  
52 involved in redox signaling (Mittler et al., 2011; Vaahtera et al., 2014).

53 Among the environmental constraints, drought is one of the most limiting factors to plant growth,  
54 establishment, and survival (Cruz de Carvalho, 2008). An ecosystem in which plants face very frequent water  
55 deficit mainly in the dry season is the Brazilian rupestrian fields. This ecosystem is characterized by a typically  
56 altitudinal tropical climate, with mainly two distinct seasons, a dry and a rainy one. However, because it is  
57 located in sites with an altitude above 1,000 m, fog is a frequent meteorological phenomenon (Bruijnzeel, 1990;

58 Baeta, 2012). Besides, this environment can be considered arid due to the high evaporative demand in the dry  
59 season without fog (Leuschner, 2000), emphasizing its importance to this ecosystem.

60 Moreover, many plants from this ecosystem can absorb vapor water or liquid water through the leaves  
61 (Boanares et al., 2018a). Foliar water uptake (FWU) occurs in response to the water gradient potential when  
62 water in the leaf surface diffuses into the internal tissues through the cuticle (Yates and Hutley, 1995; Gouvra  
63 and Grammatikopoulos, 2003), absorbent trichomes (Benzing et al., 1978) and through hydathodes (Martin and  
64 Willert, 2000). Thus, the FWU provides an ecological advantage for plants that occur in the environments that  
65 reveal water deficit, at least seasonally, playing an important role in their cell physiological process and plant  
66 growth (Limm et al., 2009; Eller et al., 2013).

67 The FWU capacity has already been verified in 11 species from this ecosystem and different FWU strategies  
68 have been revealed (Boanares et al., 2018a, b). There are plants which present different foliar water uptake FWU  
69 capacities - species that absorb less water more quickly (LQ) and those that absorb more water more slowly  
70 (MS) and coexist in this same ecosystem (Boanares et al., 2018a, b). Therefore, the hypotheses of this study are:  
71 1) there is a relation between FWU capacity and oxidative stress/signaling response, mainly in the dry season  
72 when plants have to cope with water deficit; 2) that the different FWU strategies are related to this metabolic  
73 oxidative response.

## 74 75 **2. Material and Methods**

### 76 77 *2.1. Site study*

78  
79 This study was carried out in a Brazilian ferruginous rupestrian field in the municipality of Ouro Preto (20°  
80 21'30"S/43°30'11"W). The area has an average altitude of 1,492 meters above sea level, and fog is frequent and  
81 accounts for approximately 13% of the total precipitation during the year. However, in the dry season, it  
82 contributed 10-fold more than rainfall (Baeta, 2012). Local vegetation is present on shallow soils on rock  
83 outcrops of itabirites or *cangas* and is characterized by herbs, shrubs and treelets (Valim et al., 2013). Data were  
84 obtained in January and July 2018, characterized as the rainiest and the driest months of both seasons,  
85 respectively. The site, therefore, provides a possibility of drought stress for plants in the dry season. The average  
86 annual temperature was 16.9°C with the total annual rainfall of 1,204.7 mm (Fig. 1).

87

88 2.2. Plant material and FWU measurements

89  
90 Leaves of *Pleroma heteromallum*, *Trembleya laniflora*, *Miconia australis* and *Miconia corallina*  
91 (Melastomataceae), *Ocotea pulchella* (Lauraceae) and *Byrsonima variabilis* (Malpighiaceae) were harvested to  
92 evaluate the FWU. Leaves (n = 4) were collected in the field and immediately weighed with a portable scale.  
93 After the leaves were taken to the laboratory, they were weighed once again. Then, leaves were immersed in  
94 distilled water and weighed every 15 min for 2 h, and every 30 min for 2 h. At last, leaves were immersed in  
95 water for 1 h and weighed to obtain the maximum water absorption curve. Before each weighing, leaves were  
96 then dried with ultra-absorbent microfiber synthetic towels to remove the surface water (Liang et al., 2009).  
97 FWU was calculated by a differential equation  $\Delta C = \Delta C_{\max}(1 - e^{-kt})$ , where  $\Delta C = (C - C_i)$  is the difference  
98 between the amount of water present in the leaves in an interval of 300 min, and  $\Delta C_{\max} = (C_s - C_i)$  the difference  
99 between the maximum amount of water after the end of the experiment, providing data on the maximum water  
100 content in leaf surface ( $C_{\max}$ ) and leaf uptake speed ( $k$ ) (Liang et al., 2009).

101

102

103 2.3.  $H_2O_2$  and lipid peroxidation evaluation

104

105 Frozen leaves of the six species studied (0.3 g) were ground into powder in mortar with liquid nitrogen and  
106 homogenized with 0.1 % (v/v) trichloroacetic acid (TCA). Then, plant samples were centrifuged at  $13,000 \times g$  at  
107  $4^\circ C$  for 20 min and the supernatant was collected for further analysis of  $H_2O_2$  and malondialdehyde (MDA). The  
108 amount of  $H_2O_2$  was determined essentially as described by Velikova et al. (2000). In the reaction medium  
109 presenting 1 mM of phosphate buffer (pH 7.0) and 1 M potassium iodide, volumes of 300  $\mu\text{m}^3$  of supernatant  
110 were added. After, samples were incubated at  $30^\circ C$  for 30 min, absorbance was determined at 390 nm. The  
111  $H_2O_2$  concentrations were estimated based on a calibration curve.

112 The concentration of MDA was used as a parameter to evaluate lipid peroxidation. For this purpose, the  
113 supernatant was incubated in a reaction medium containing 0.5% thiobarbituric acid (w/v) and 10% TCA (w/v)  
114 at  $90^\circ C$ . The reaction was stopped with an ice bath for 10 min, 30 min later samples were centrifuged for 5 min  
115 at  $13,000 \times g$ . The absorbance was measured at 535 and 600 nm, and the MDA concentration was calculated as  
116 described by Cakmak and Horst (1991).

117

#### 118 2.4. Activity of antioxidant enzymes evaluation

119

120 Frozen leaves of the six species studied (0.3 g) were ground in mortar with liquid nitrogen, and proteins were  
121 extracted with a 100 mM phosphate buffer (pH 7.8) containing 100  $\mu$ M ethylenediaminetetraacetic acid, 20 mM  
122 ascorbic acid and 50% (w/w) polyvinylpyrrolidone. After centrifuging at  $13,000 \times g$  at  $4^\circ\text{C}$  for 20 min, the  
123 supernatant was collected for further analysis of the superoxide dismutase (SOD), catalase (CAT) and ascorbate  
124 peroxidase (APX) enzymes. The method of nitroblue tetrazolium (NBT) photoreduction (Giannopolitis and Ries,  
125 1977) was used to evaluate SOD (EC 1.15.1.1) activity. One unit of SOD was defined as the amount of enzyme  
126 required to inhibit NBT reduction by 50%. To determine the CAT (EC 1.11.1.6) activity, plants extracts were  
127 incubated with 100 mM phosphate buffer (pH 7.0) and 12.5 mM  $\text{H}_2\text{O}_2$  (Azevedo et al., 1998). For the enzyme  
128 activity determination, the molar extinction coefficient ( $\epsilon$ ) of  $= 39.4 \text{ M cm}^{-1}$  from measurements of  $\text{H}_2\text{O}_2$   
129 degradation at 240 nm was used. To evaluate APX (EC 1.11.1.11) activity, 100 mM of phosphate buffer (pH  
130 7.0), 0.5 mM ascorbic acid and 0.1 mM  $\text{H}_2\text{O}_2$  was used (Nakano and Asada, 1981). The rate of ascorbic acid  
131 oxidation was monitored at 290 nm and the quantification was carried out using  $\epsilon$  equal to  $2.8 \text{ mM}^{-1} \text{ cm}^{-1}$ .

132

#### 133 2.5. Data analyses

134

135 General linear mixed models (GLMMs) with gaussian errors were built to evaluate the influence of FWU  
136 capacity on oxidative response related to oxidative stress. The  $\text{H}_2\text{O}_2$ , MDA, SOD, APX, and CAT were used as  
137 fixed explanatory variables, and the response variables were maximum water content in leaf ( $C_{\text{max}}$ ), the water  
138 uptake speed ( $k$ ) and season. The identity of each measured plant was used as a random effect variable to control  
139 for potential pseudoreplication biases in our models (Hurlbert, 1984). For all models, distributions were tested  
140 through residual analysis. All analyses were performed using the R platform (R Core Team, 2015) using the  
141 function GLMER from package lme4 (Bates et al., 2015).

142

### 143 3. Results

144

145 There were significant differences among species analyzed according to the FWU strategies. Highest  $C_{\text{max}}$   
146 and lowest  $k$  values were observed in *M. corallina*, *P. heteromallum* and *T. laniflora*. Contrastingly, *M. australis*,

147 *B. variabilis* and *O. pulchella* presented a different pattern, that is, smaller values of  $C_{\max}$  and higher values of  $k$   
148 in dry season (Fig. 2) and rainy season (see Supplementary Fig. S1).

149 As expected, the highest values for all parameters of oxidative stress responses occurred in the dry season.  
150 The comparison between dry and rainy season data can be found in the Supplementary data (Fig. S2). However,  
151  $H_2O_2$ , besides having a significant relationship among different plant species and seasons it is related to the two  
152 parameters of FWU -  $C_{\max}$  and  $k$  (Table 1). This relation of FWU parameters is antagonistic since the increase of  
153  $H_2O_2$  was directly related to the amount of water absorbed ( $C_{\max}$ ) and inversely proportional to the rate of water  
154 absorption ( $k$ ). In other words, the species with the highest values of  $C_{\max}$  presented the highest  $H_2O_2$  content and  
155 the species with higher values of  $k$  presented the lowest  $H_2O_2$  content, mainly in the dry season (Fig. 3). These  
156 data corroborate the opposite pattern of the parameters  $C_{\max}$  and  $k$  according to the statistical difference among  
157 species. Only *T. laniflora* that presents strategy with higher  $C_{\max}$  value and lower  $k$  value, and *B. variabilis* that  
158 presents strategy with higher  $k$  value and lower  $C_{\max}$  value did not fit the patterns of the different FWU  
159 strategies. However, the lipid peroxidation estimated by MDA values had no significant relation with any FWU  
160 parameters ( $C_{\max}$  and  $k$ ). This relation of lipid peroxidation was only found between species and seasons (Table  
161 1). This non-relation with MDA and FWU was observed in the significant differences among species in which  
162 there was no clear separation between the FWU strategies, that is, species *M. corallina*, *P. heteromallum* and *T.*  
163 *laniflora* that have different FWU strategy than *B. variabilis* and *O. pulchella* showed no significant differences  
164 for lipid peroxidation (Fig. 3). However, the antioxidant enzymes SOD, APX and CAT analyzed were  
165 significantly related to  $C_{\max}$ , plant species and season (Table 1). These enzymes had a positive relationship with  
166  $C_{\max}$  mainly in the dry season (Fig. 4). APX and CAT were the enzymes that best separated the species  
167 according to FWU strategies.

168

#### 169 4. Discussion

170

171 It is widely known that oxidative metabolism, which comprises the production of ROS and the antioxidant  
172 response, is closely linked to the plant water status, as well as plant species and season. Also, we know that the  
173 ability of plants to absorb water through leaves is an adaptive advantage, because despite a small amount of  
174 water entry, this water is enough to enable rapid leaf hydration, which is the primary organ for photosynthesis  
175 and gas exchanges. Although the plant response to oxidative stress depends on the type of ROS molecule and its  
176 concentration, but also the site of production and interaction with other molecules (Moller et al., 2007), it was

177 shown that the different strategies of FWU reflect in distinct H<sub>2</sub>O<sub>2</sub> production. Among the different ROS  
178 molecules, H<sub>2</sub>O<sub>2</sub> was analyzed for being relatively stable and present at higher levels of concentration concerning  
179 the other ROS molecules, being, therefore, the most representative molecule (Moller et al., 2007). In addition,  
180 H<sub>2</sub>O<sub>2</sub> has the ability to cross membranes, possibly through aquaporins (Bienert et al., 2007), and is the main  
181 secondary messenger acting on a signal transduction pathway for stress response (Mittler, 2002; Vranova et al.,  
182 2002; Miller and Mittler, 2006). Thus, their concentrations can have different consequences for the plant, if  
183 maintained at relatively low levels, it is likely that the function is to act as a signaling pathway for stress, defense  
184 responses/stress acclimatization (Moller et al., 2007). However, upon reaching a certain level of concentration,  
185 ROS become extremely harmful, activating uncontrolled oxidative cascades that result in biochemistry stress  
186 and, eventually, cell death (Mignolet-Spruyt et al., 2016).

187 Therefore, the different concentrations of H<sub>2</sub>O<sub>2</sub> in the studied species are related to characteristics of the  
188 species, but also the FWU strategies, culminating in differences in leaf hydration, especially in the dry season,  
189 when the highest concentration of this molecule occurred. In the rainy season, there was also the detection of  
190 H<sub>2</sub>O<sub>2</sub>, despite the lower concentration in relation to the dry season. Probably it is due to its role of signalization  
191 and not of its involvement in drought stress, since, in this condition, plants continue to produce ROS (Cruz de  
192 Carvalho, 2008; Zandalinas et al., 2018). Plants can use a stable cellular ROS level to monitor their intracellular  
193 stress level (Mittler, 2002), but this stationary level must be strictly regulated to avoid an oxidative burst on the  
194 ROS accumulation, which would result in extensive cellular damage (Dinakar et al., 2010; Baxter et al., 2014;  
195 Choudhury et al., 2017).

196 Plants are extremely tolerant to H<sub>2</sub>O<sub>2</sub> and antioxidant systems appear to function as rigid controllers of the  
197 cellular redox state rather than destroyers of all intracellular H<sub>2</sub>O<sub>2</sub> (Mittler et al., 2004). In this sense,  
198 antioxidants are key components in the modulation of the ROS signal, since they determine lifetime and  
199 intensity of this signal (Foyer and Noctor, 2013). Regarding the antioxidant response and the FWU strategies,  
200 only C<sub>max</sub> parameter had a significant influence on the SOD, CAT and APX enzymes production since the  
201 highest concentrations of these enzymes are proportional to H<sub>2</sub>O<sub>2</sub> production. Thus, the species with the highest  
202 concentrations of antioxidant enzymes were those with the highest C<sub>max</sub> values. SOD is the first enzyme in the  
203 line of defense against damages produced by ROS and the most general one, also having affinity with other ROS  
204 molecules (Boguszewska et al., 2010). One of its functions is the elimination of ROS during drought stress,  
205 possibly through the water-water cycle, thus protecting the photosynthetic apparatus from oxidative damage  
206 induced by drought (Cruz de Carvalho, 2008), since all species need a larger amount of water in their tissues for

207 a more effective hydration, justifying a junction of the FWU strategies among the species and the concentrations  
1  
2 208 of SOD.

3  
4 209 Due to the high affinity of APX with  $H_2O_2$ , this activity increase in the dry season has shown that APX is  
5  
6 210 responsible for the elimination of high intracellular levels of  $H_2O_2$  produced under this constraint condition  
7  
8 211 (Cruz de Carvalho, 2008). Thus, this result reveals the clear separation between different FWU strategies, that is,  
9  
10 212 the highest APX concentrations was related to the highest  $H_2O_2$  content present in species that have the highest  
11  
12 213  $C_{max}$  strategy. This combination of increased SOD and APX concentration in the dry season demonstrates a high  
13  
14 214 efficiency in increasing tolerance to water deficit-induced oxidative damage due to simultaneous scavengers  
15  
16 215 (Lee et al., 2007).

17  
18 216 However, CAT has the characteristic of being a sequestering molecule less susceptible than APX in relation  
19  
20 217 to the oxidative stress. This finding indicates that only under severe drought stress the activity of CAT is  
21  
22 218 increased. However, under moderate drought stress,  $H_2O_2$  removal is preferably done by ascorbic acid from the  
23  
24 219 ascorbate / glutathione cycle (Moller et al., 2007). Because it has a lower affinity for  $H_2O_2$  than APX, CAT only  
25  
26 220 acts when there is excess of  $H_2O_2$ , because this may inhibit APX. Therefore, CAT activity is more probable to  
27  
28 221 occur in maintaining APX activity under severe drought stress (Cruz de Carvalho, 2008).

29  
30 222 Maintaining the normal level of intracellular ROS requires a delicate balance between ROS production and  
31  
32 223 elimination. Considering the drought-induced water stress, this balance is shifted upwards, resulting in an  
33  
34 224 increase in ROS production due to stomatal closure and the consequent limitation in  $CO_2$  fixation, as seen in the  
35  
36 225 results of oxidative stress/signaling and the antioxidant response in this study. However, rather than having an  
37  
38 226 immediate detrimental effect, this increase in ROS production is likely to be beneficial for the plant to be kept  
39  
40 227 under balanced control. Higher ROS production activates defense pathways and acclimatization responses,  
41  
42 228 resulting in an adaptation of the plant to environmental changes (Shi et al., 2013; Gollmack et al., 2014).

43  
44 229 The majority of higher  $C_{max}$  strategy species had the highest values of  $H_2O_2$  compared to  $k$  strategy species in  
45  
46 230 the dry season. We believe that this higher production of  $H_2O_2$  is related to its greater need for foliar hydration  
47  
48 231 due to the lower speed of water in leaf tissue and slower absorption (Boanares et al., unpublished data).  
49  
50 232 However, in the dry season, regardless of the FWU strategies and the  $H_2O_2$  content observed, the lipid  
51  
52 233 peroxidation occurred in all studied species, which was confirmed through the non-significant relation between  
53  
54 234  $C_{max}$  and  $k$  with the lipid peroxidation. So, the sensitivity to lipid peroxidation is related only to the plant species  
55  
56 235 and the season. However, the production of  $H_2O_2$  which can culminate in lipid peroxidation is related to both  
57  
58 236  $C_{max}$  and  $k$ . Although the antioxidant response was proportional to  $H_2O_2$  content, the production of antioxidant

237 enzymes was not sufficient to prevent lipid peroxidation. Therefore, the capacity of FWU, with the temporary  
238 and rapid hydration of the foliar tissues, should attenuate ROS production, reducing oxidative stress and  
239 decreasing the chances of chronic stress and consequent more cell damage. Thus, the plants of this environment  
240 depend on the alternative source of water from the fog and collected by the leaves present in the dry season to  
241 attenuate the water deficit imposed by this season.

242 Possibly, there is a fine adjustment between the foliar absorption capacity and the oxidative signaling.  
243 Perhaps due to water deficit pressure experienced by plants, especially in the dry season, a selection of different  
244 FWU strategies was induced. A possible relationship between FWU and ROS is in the role of H<sub>2</sub>O<sub>2</sub> being  
245 involved in cell wall differentiation (Gapper and Dolan, 2006). Its role in the process of cell wall differentiation  
246 is not well known. However, it is known that H<sub>2</sub>O<sub>2</sub> can act by raising the activity of cellulose synthases that are  
247 activated during the growth of cotton fiber. In this way, it can be inferred that H<sub>2</sub>O<sub>2</sub> can increase the proportion  
248 of cellulose, which consequently increases the capacity to absorb a greater amount of water throughout the high  
249 C<sub>max</sub> plants (Boanares et al., 2018b). This relationship may explain the higher content of H<sub>2</sub>O<sub>2</sub> in these species,  
250 and may have a key role in the proportion of pectin/cellulose (Boanares et al., 2018b) in the cell wall to grant  
251 greater amount of absorbed water.

252

## 253 5. Conclusion

254

255 It was verified that species with the high *k* value strategy are more sensitive to lipid peroxidation than species  
256 with the high C<sub>max</sub> value strategy, because they needed a smaller amount of ROS to induce the lipid peroxidation.  
257 Species that have a higher speed of leaf water absorption (higher *k*), but uptake a small amount of water (lower  
258 C<sub>max</sub>) are more sensitive. Thus, these species should have faster foliar water absorption to mitigate the damage  
259 caused by ROS. The species with higher C<sub>max</sub> value, because they tolerate a higher concentration of ROS, use  
260 H<sub>2</sub>O<sub>2</sub> as a signal and thus possibly modify the proportion of the constituents of the leaf cell wall, generating a  
261 slower FWU, but in a larger quantity. In this way, it can be inferred that the oxidative stress response is  
262 associated with the FWU strategies (Fig. 5). Therefore, it is assumed that fog as an alternative source of water  
263 for vegetation has an important ecological role for the maintenance of these species, being the only source of  
264 water available to plants to deal with the oxidative stress/signaling induced by drought stress in this ecosystem.

265

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267

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386 **Figure captions**

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4 388 **Fig. 1.** Mean annual rainfall and mean annual temperature at Brazilian ferruginous rupestrian field at 2018. —  
5  
6 389 represent total precipitation volume (mm); - - - represents monthly temperature mean.

7  
8 390  
9  
10 391 **Fig. 2.** Dynamic water content in leaves by submerging time (min) in dry season. Equations: *P. heteromallum*,  $y$   
11  
12 392 =  $0.5575(1-e^{-0.0372t})$ ,  $r^2 = 0.93$ ; *T. laniflora*,  $y = 0.7985(1-e^{-0.026t})$ ,  $r^2 = 0.94$ ; *M. corallina*,  $y = 0.0789(1-e^{-0.0624t})$ ,  
13  
14 393  $r^2 = 0.97$ ; *B. variabilis*,  $y = 0.4508(1-e^{-0.0621t})$ ,  $r^2 = 0.87$ ; *L. australis*,  $y = 0.4508(1-e^{-0.0621t})$ ,  $r^2 = 0.87$ ; *O.*  
15  
16 394 *pulchella*,  $y = 0.0789(1-e^{-0.0624t})$ ,  $r^2 = 0.97$ .

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18 395  
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20 396 **Fig. 3.** (A) Relationship between  $H_2O_2$  and maximum leaf water absorption ( $C_{max}$ ) and (B) leaf uptake speed ( $k$ )  
21  
22 397 in the dry season. (C) Quantity of MDA that represent lipid peroxidation among species. Black and grey bars  
23  
24 398 indicate species with higher  $C_{max}$  and species with lower  $C_{max}$ , respectively. Regression values are means of the  
25  
26 399 four individuals for each species. Significant differences among species were tested by contrast analysis of R  
27  
28 400 program with letters comparing among species ( $P < 0.01$ ).

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30 401  
31  
32 402 **Fig. 4.** Relationship between activities of antioxidant enzymes and maximum leaf water absorption ( $C_{max}$ ) in dry  
33  
34 403 season: (A) superoxide dismutase (SOD), (B) ascorbate peroxidase (APX) and (C) catalase (CAT). Black and  
35  
36 404 grey bars indicate species with higher  $C_{max}$  and species with lower  $C_{max}$ , respectively. Regression values are  
37  
38 405 means of the four individuals for each species. Significant differences among species were tested by contrast  
39  
40 406 analysis of R program with letters comparing among species ( $P < 0.01$ ).

41  
42 407  
43  
44 408 **Fig. 5.** Schematic comparison of two contrasting foliar water uptake (FWU) strategies. (A) species that absorb  
45  
46 409 less water more quickly (LQ) and (B) species that absorb more water more slowly (MS), indicating the  
47  
48 410 differences in plant water regime and ROS ( $H_2O_2$ ) and antioxidant enzyme (catalase, superoxide dismutase and  
49  
50 411 ascorbate peroxidase) levels in the leaf.

51  
52 412  
53  
54 413 **Supplementary material**

55  
56 414

415 Fig. S1. Dynamic water content in leaves by submerging time (min) in rainy season. Equations: *P. heteromallum*  
1  
2 416  $y = 0.0474(1-e^{-0.08t})$ ,  $r^2 = 0.94$ ; *T. laniflora*  $y = 0.0275(1-e^{-0.002t})$ ,  $r^2 = 0.98$ ; *M. corallina*  $y = 0.05117(1-e^{-0.004t})$ ,  
3  
4 417  $r^2 = 0.96$ ; *B. variabilis*  $y = 0.0333(1-e^{-0.113t})$ ,  $r^2 = 0.84$ ; *L. australis*  $y = 0.082(1-e^{-0.0145t})$ ,  $r^2 = 0.83$ ; *O. pulchella*  $y$   
5  
6 418  $= 0.028(1-e^{-0.0055t})$   $r^2 = 0.98$ .

7  
8 419  
9  
10 420 Fig. S2. Relationship between H<sub>2</sub>O<sub>2</sub> content and activities of antioxidant enzymes in different plant species from  
11  
12 421 ferruginous rupestrian field (A) H<sub>2</sub>O<sub>2</sub> content, (B) superoxide dismutase (SOD), (C) catalase (CAT) and (D)  
13  
14 422 ascorbate peroxidase (APX). Black and grey bars indicate species with higher C<sub>max</sub> and species with lower C<sub>max</sub>,  
15  
16 423 respectively. Significant differences among species were tested by contrast analysis of R program with letters  
17  
18 424 comparing among species ( $P < 0.01$ ).

19  
20 425

## Contribution

Daniela Boanares and Marcel Giovanni Costa França designed research, performed all analyses and wrote the first draft of the manuscript. Cristiane Jovelina Silva and Rosy Mary dos Santos Isaias performed the experiment and helped to collect data. All the authors contributed to the experimental design, and significantly improved later drafts.

**Table 1.** Oxidative stress parameters, such as H<sub>2</sub>O<sub>2</sub> concentration, MDA concentration to evaluate lipid peroxidation and antioxidant enzymes (SOD), (APX) and (CAT) related to two FWU parameters: maximum leaf water content (C<sub>max</sub>) and water uptake speed (k).

Dependent variable	Independent variable	Deviance	F	DV	P value
H <sub>2</sub> O <sub>2</sub>	C <sub>max</sub>	390.04	6.3109	11	0.012*
	k	390.04	6.4441	11	0.0111*
	Species	390.04	41.843	11	0.0001***
	Season	390.04	87.873	11	0.0001***
MDA	C <sub>max</sub>	479.3	0.763	11	0.3824
	k	479.3	3.0775	11	0.0793
	Species	479.3	21.424	11	0.0001***
	Season	479.3	84.928	11	0.0001***
SOD	C <sub>max</sub>	94.826	8.7544	11	0.0030**
	k	94.826	2.1928	11	0.1387
	Species	94.826	36.791	11	0.0001***
	Season	94.826	56	11	0.0001***
APX	C <sub>max</sub>	-7.1862	14.731	11	0.0001***
	k	-7.1862	0.662	11	0.4159
	Species	-7.1862	14.152	11	0.0146*
	Season	-7.1862	34.877	11	0.0001***
CAT	C <sub>max</sub>	325.49	18.212	11	0.0001***
	k	325.49	1.4351	11	0.2309
	Species	325.49	29.63	11	0.0001***
	Season	325.49	35.365	11	0.0001***

The general linear mixed models (GLMMs) for the biotic independent variables evaluated. \*\*\* Indicates  $P \leq 0.001$ ; \*\*Indicates  $P \leq 0.01$  and \* Indicates  $P \leq 0.05$ .

Figure 1  
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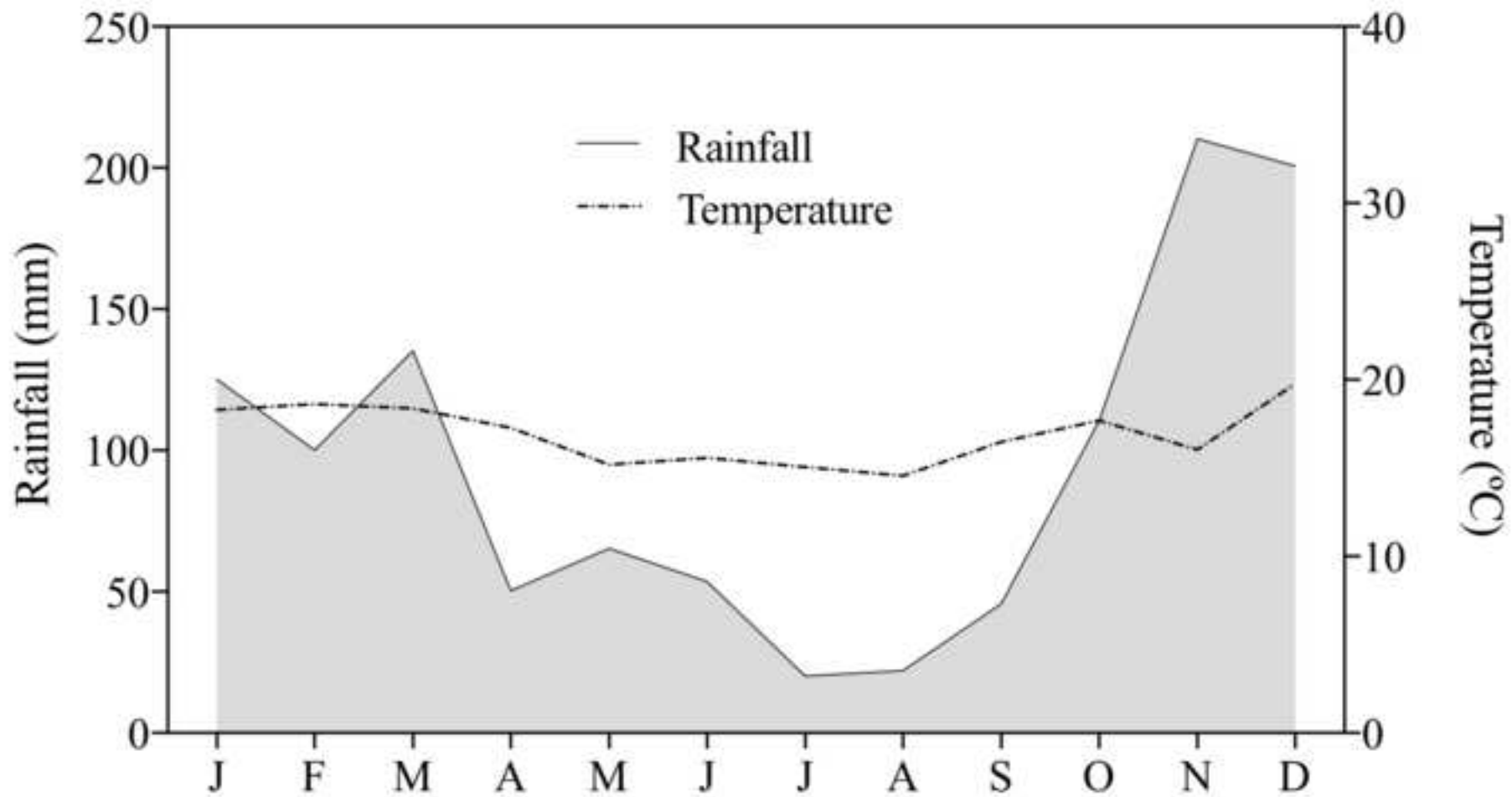


Figure 2  
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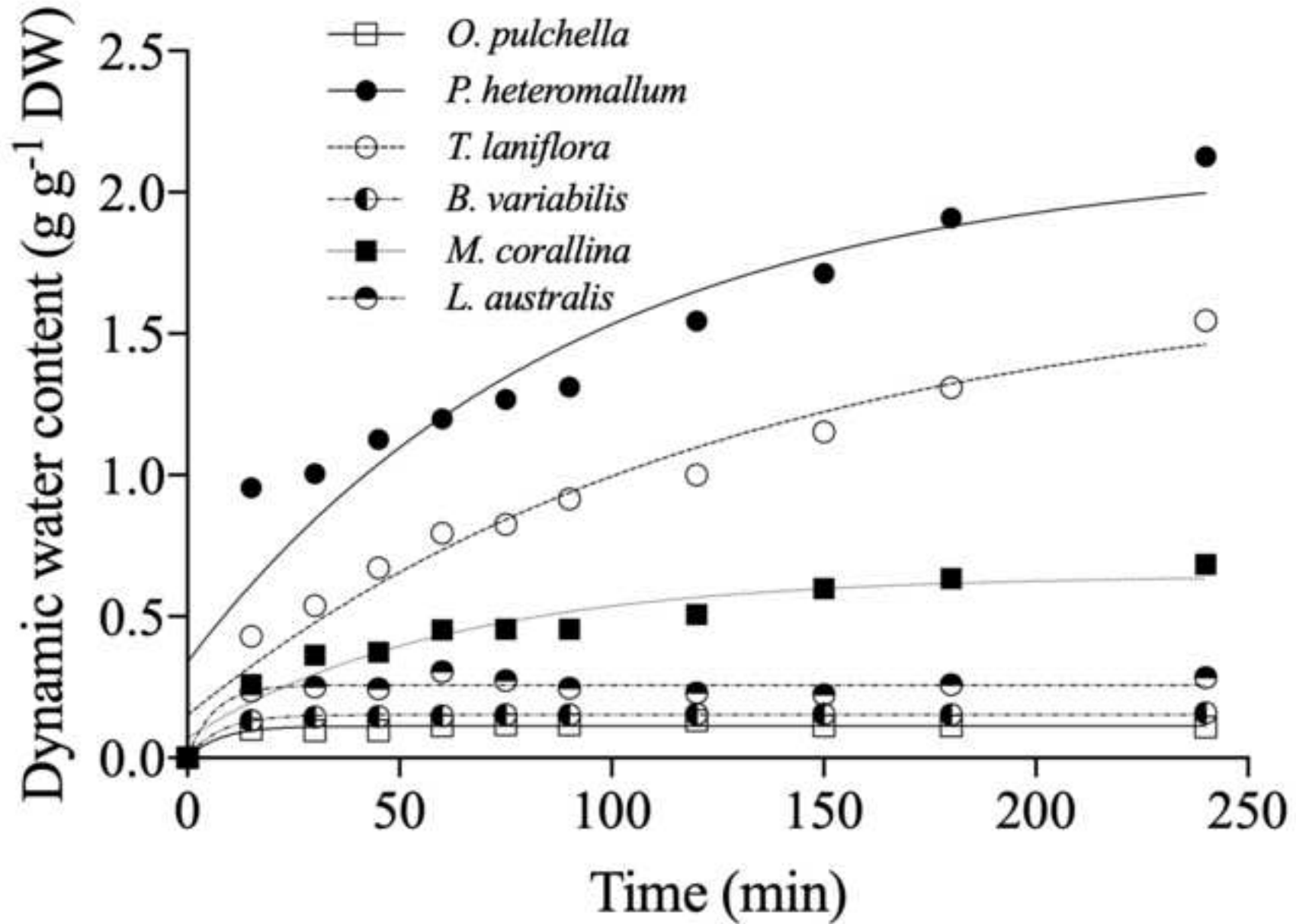


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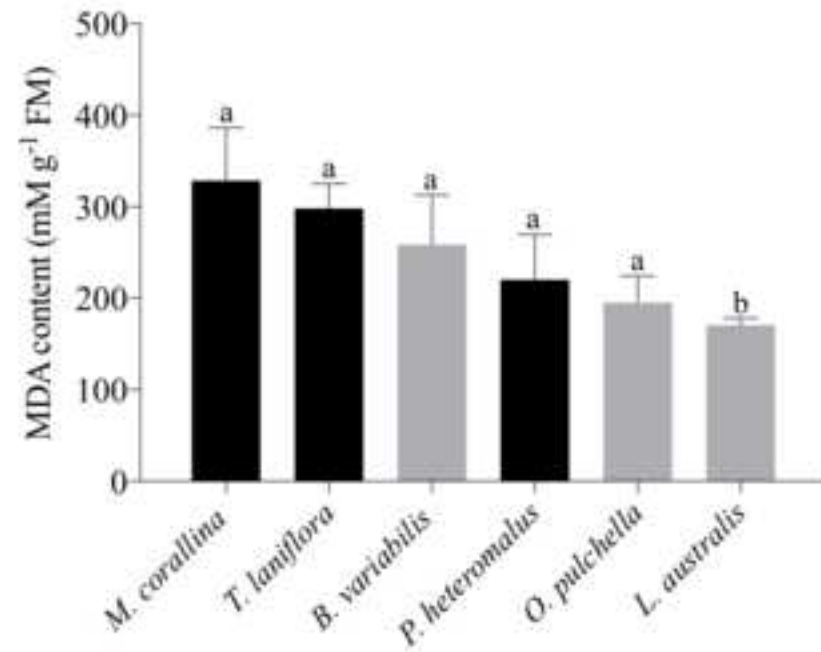
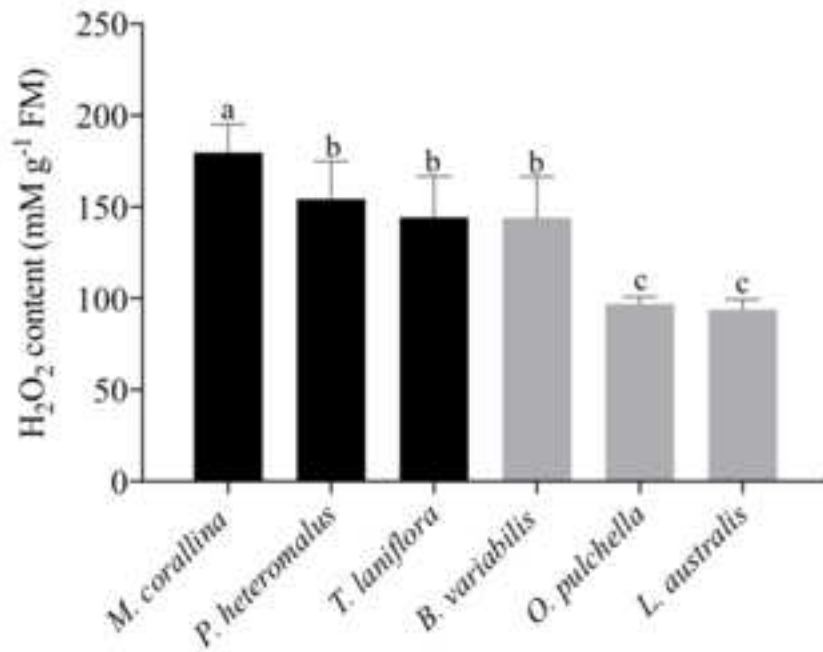
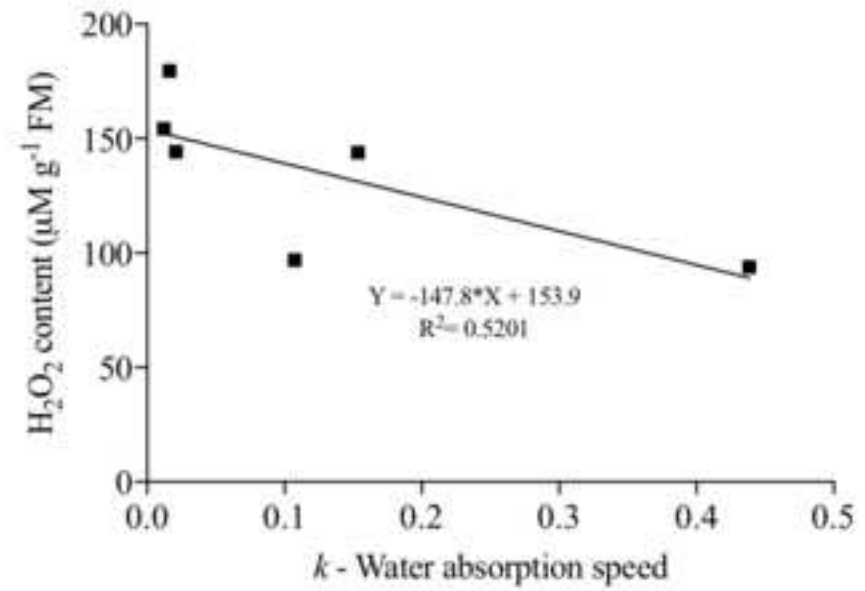
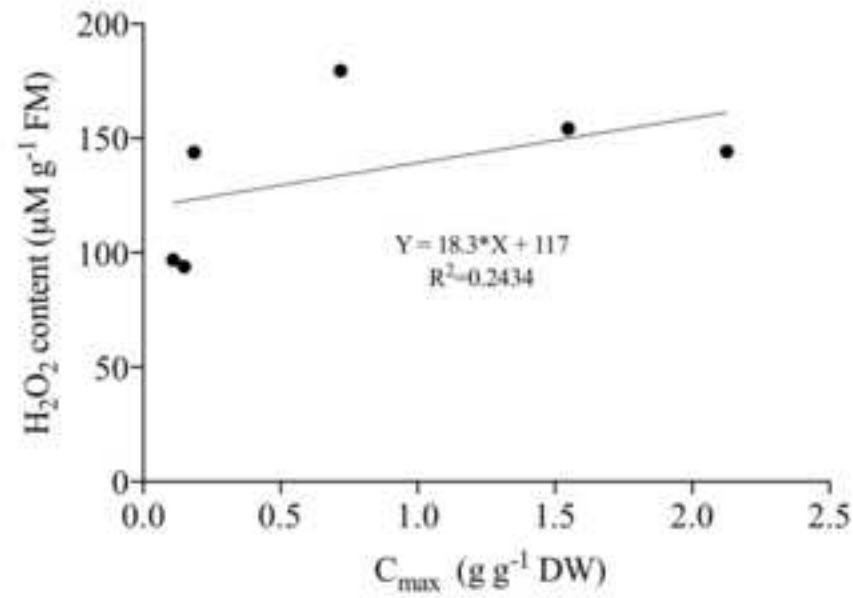
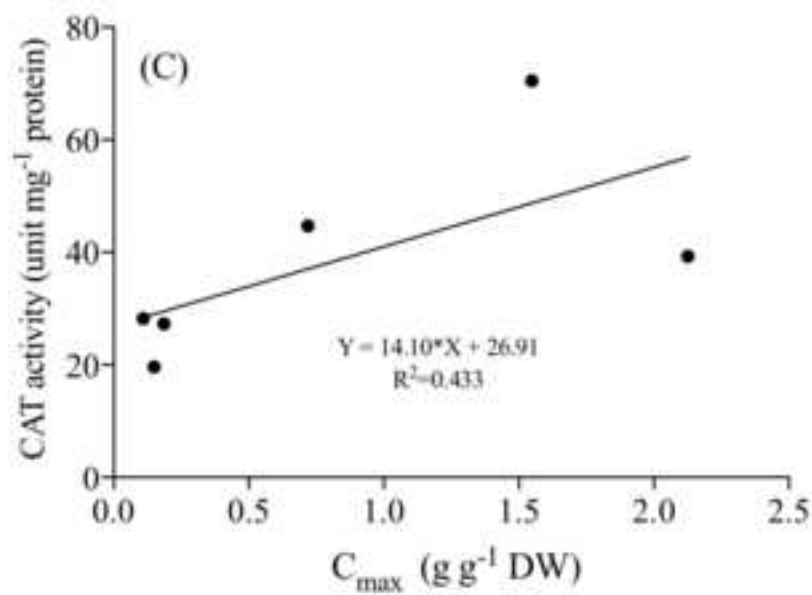
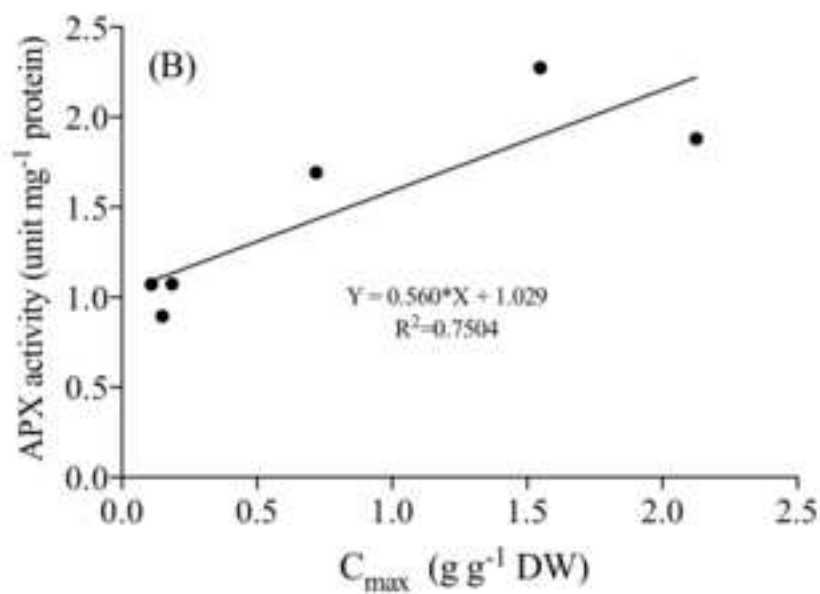
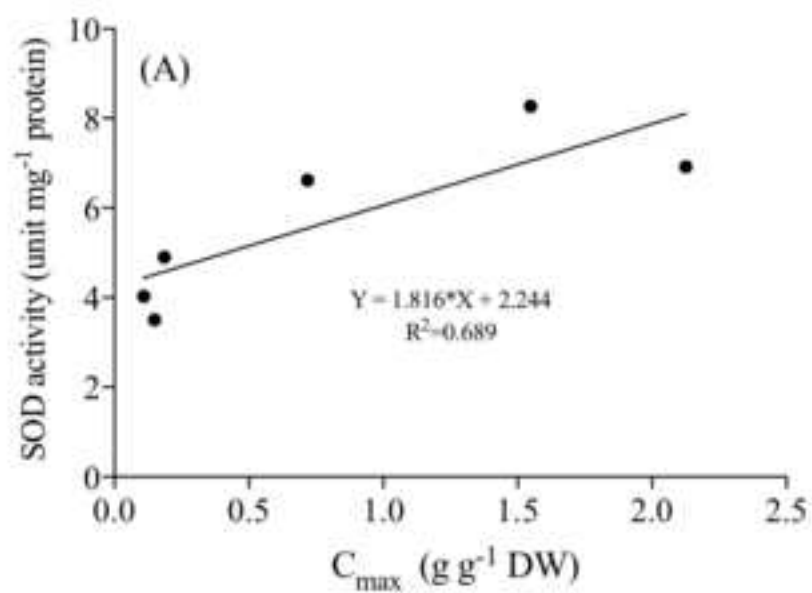
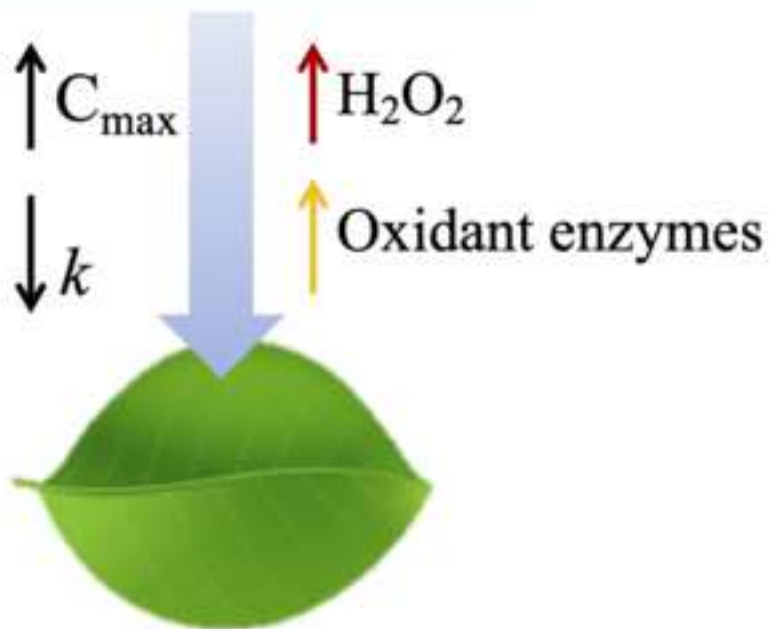


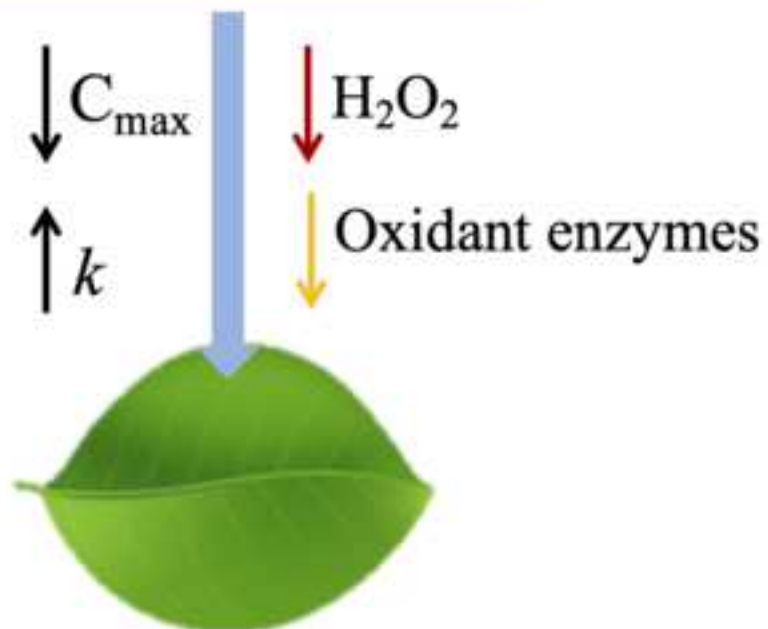
Figure 4  
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**A** More and slowly  
water absorption



**B** Less and quickly  
water absorption



An aerial photograph of a mountain landscape. The mountain is covered in a dense forest of trees, with some trees showing autumnal colors of yellow and orange. Several rocky outcrops are visible on the mountain's surface. A central path or stream bed runs down the mountain, flanked by more trees. The overall scene is a mix of green, brown, and grey tones.

*The influence of fog in the leaf physiology and phenology of two Mediterranean tree species*

1           **The influence of fog in the leaf physiology and phenology of two**  
2                                   **Mediterranean tree species**

3  
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12  
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14  
15           **ABSTRACT**

16  
17           Fog may play a role as an alternative source of water for vegetation, and the leaf absorption  
18           capacity of fog is an important strategy for plants of different terrestrial ecosystems. Thus, our  
19           objective was to evaluate the importance of fog in the physiology of the most common native  
20           species of European forests, *Fagus sylvatica* and *Quercus ilex*, and leaf phenology of *F.*  
21           *sylvatica*, a deciduous species. We evaluated the physiological performance of plantlets  
22           subjected to 3 environmental conditions in a greenhouse experiment: artificial fog, irrigation,  
23           and water deficit. To assess whether the duration of *F. sylvatica* leaves was related to the  
24           presence of fog, the 11-year remote sensing data from four regions of a Mediterranean-  
25           temperate ecotone were analyzed. Fog significantly improve the physiology of both species,

26 especially *F. sylvatica*, preventing or delaying the occurrence of photoinhibition, and  
27 increasing the water status of plants. Vegetative indices were responsive to fog only for *F.*  
28 *sylvatica*, whose phenology has a significant relationship with the presence of fog. Short-  
29 duration fog events accelerates leaf falling, therefore, fog not only improves the physiological  
30 parameters of the two studied species, but also plays an important role on *F. sylvatica*  
31 phenology.

32

33 **Key words:** *Fagus sylvatica*; *Quercus ilex*; Foliar water uptake; Photosynthesis

34

35 **List of abbreviation:**

36  $C_{\max}$  - maximum water content in leaf lamina

37  $k$  - water leaf uptake speed

38  $Fv/Fm$  - potential quantum yield of photosystem II

39 PRI - photochemical reflectance index

40 NDVI - Normalized Difference Vegetation Index

41

42 **Author contributions**

43

44 D.B., J.P., R.S.O. and M.G.C.F. designed the research. D.B., J.L., R.O. and I.F. collected the  
45 data for the research, and D.B. and analyzed the data. D.B., R.M.S.I. and M.G.C.F wrote the  
46 manuscript.

47

48 **HIGHLIGHTS**

49

- 50 • Leaf physiology was closely related to fog in *Fagus sylvatica* and *Quercus ilex*
- 51 • Fog prevented or delayed the occurrence of photoinhibition in plants
- 52 • Leaf phenology of *F. sylvatica* has a significant relationship with fog presence
- 53 • 11-years remote sensing data showed decrease in both, fog events and leaf duration

54

## 55 **Introduction**

56

57 Fog is an important meteorological phenomenon for the hydrological cycle in several  
58 terrestrial ecosystems (Holder 2004, 2006; Prada et al., 2009), especially at altitudes above  
59 800 meters (Mc Jannet et al., 2007). Its formation can occur through the encounter of hot and  
60 cold air masses, when the cooling forms fog and low clouds (Burgess and Dawson 2004). The  
61 air mass stability, environment topography, and the flow and direction of humidity are  
62 conditions which may induce fog formation (Money 2000). In addition to contributing to the  
63 hydrology of ecosystems, fog may play a relevant ecological role as an alternative source of  
64 water for vegetation. First, it can decrease the vapor pressure deficit of the atmosphere,  
65 reducing evaporation and transpiration on plant surfaces (Juvik and Nullet 1995). If wind  
66 directs fog flow through the canopy, much of the fog water may be intercepted by the  
67 vegetation surfaces, where it would remain for some time, returning to the atmosphere  
68 through evaporation, or wetting the soil by the water runoff (Hutley et al., 1997). Also, there  
69 is the possibility of water absorption through plant leaves (Martin and von Willert 2000).

70 Foliar water uptake (FWU) occurs when water crosses leaf surface in response to the  
71 potential gradient diffusing into the internal plant tissues through the cuticle (Yates and  
72 Hutley 1995; Gouvra and Grammatikopoulos 2003), and/or absorbent trichomes (Benzing et  
73 al., 1978) and/or through hydathodes (Martin and von Willert 2000). FWU can occur during  
74 and after various forms of precipitation, such as rain and fog. Several studies have been

75 carried out to evaluate the capacity of fog FWU, and its consequences for the ecology and  
76 ecophysiology of different plant species (Holder 2006; Simon et al., 2009). Although FWU  
77 represents a small portion of water entering the leaves in relation to the total entry of water by  
78 the roots, such absorption immediately increases leaf hydration and water potential (Ewing et  
79 al., 2009; Boanares et al., 2019). Nevertheless, it may take more than a month to be  
80 transported from roots into leaves in large species such as *Sequoia sempervirens* (Cassana et  
81 al., 2006; Eller et al., 2013).

82 Fog FWU may play an important role in the survival and growth of plant species occurring  
83 in places with water deficit (Limm et al., 2009). *Fagus sylvatica* L. (European beech) and  
84 *Quercus ilex* L. (Sessile oak), the most common native species of European forests (Corcuera  
85 et al., 2004; Petritan et al., 2014), have differences in growth response to air temperature and  
86 fog presence on a Mediterranean mountain (Barbeta et al., 2019). The growth of *Fagus*  
87 *sylvatica*, a deciduous species, is more responsive to fog presence than *Quercus ilex*, an  
88 evergreen species (Barbeta et al., 2019). Models predict the fog decrease and this will  
89 possibly have consequences to structure and functioning of ecosystems (Johnstone and  
90 Dawson, 2010; LaDochy and Witiw, 2011). A decrease in fog events in this region resulted in  
91 a decrease in growth of *F. sylvatica* along the years (Barbeta et al., 2019).

92 The influence of fog events is well known in tropical environments and little in temperate  
93 forests subject to the Mediterranean climate (Barbeta et al., 2019). The Mediterranean climate  
94 is characterized by wet winters, and hot and dry summers, during which low water availability  
95 severely limits carbon gain (Ackerly et al., 2002). Fog in high altitudes may interfere with the  
96 physiology of coexisting species by reducing water stress in periods when plants experience  
97 water deficit.

98 Our objective was to study physiological responses of *Fagus sylvatica* and *Quercus ilex* to  
99 fog events, and the interference of fog on the phenology of *F. sylvatica*. Measurements of the

100 effect of fog on photosynthesis, water potential, and plant reflectance were conducted in  
101 greenhouse conditions, where the plants were subjected to three environmental conditions,  
102 artificial fog, irrigation, and water deficit. The reflectance indices can be used to estimate  
103 physiological status of plants that estimate of photosynthetic efficiency. Additionally, the  
104 duration of *F. sylvatica* leaves was evaluated by using remote sensing images from 11-years  
105 data in a Mediterranean-temperate ecotone.

106

## 107 **Materials and methods**

108

### 109 *Study species*

110

111 Natural vegetation of *Fagus sylvatica* and *Quercus ilex* coexists in Mediterranean-temperate  
112 ecotone in Montseny Natural Park (Catalonia, Spain, Northeastern Iberian Peninsula). *F.*  
113 *sylvatica* is a deciduous species and dominates the forests in the highest altitude range in the  
114 Montseny Mountains (750-1690 m a.s.l.), and *Q. ilex* is an evergreen species, and occurs in  
115 lower altitudes of the same mountains (300-1300 m a.s.l.) (Bolòs, 1983). Plantlets with  
116 approximately 1-1.20 meters used for this study were obtained from the Vivers Tortadès  
117 nursery in Saint Hilari Sacalm, Spain.

118

### 119 *Greenhouse experiment*

120

121 To evaluate the species responses to fog, plantlet of each species, 6 individuals for each  
122 treatment were subjected to different hydric conditions: artificial fog, soil irrigation and soil  
123 drought in the greenhouse of the Autonomous University of Barcelona (Barcelona, Spain).

124 Artificial fog was created by an air humidifier installed inside a chamber made with pallets  
125 and plastic (1.5 x 3.0 x 2.0 m), and was applied only overnight. The soil in pots had been  
126 sealed with parafilm and plastic bags to prevent fog from contacting the soil (Eller et al.,  
127 2004). Plants hydration status was guaranteed by everyday irrigation. Water deficit was  
128 induced by submitting the plants to a progressive drought stress, suppressing irrigation, all  
129 along the experiment for 30 days.

130

### 131 *Physiological performance*

132

133 Measurements of the potential quantum yield ( $F_v/F_m$ ) were performed at 9:00 am, using a  
134 modulated fluorescence meter (MINI-PAM Waltz, Germany), to identify the occurrence of  
135 photoinhibition. Such measurements were taken at the first day of the experiment with  
136 artificial fog (day 1), after 4 days (day 4), 8 days (day 8), 18 days (day 18), and 24 days (day  
137 24). After adaptation of leaf to the dark for about 30 min, the potential quantum yield ( $F_v/F_m$ )  
138 of photosystem II (PSII) (Rascher et al., 2000) were measured. In order to calculate the  $F_v/F_m$   
139 ( $F_v = F_m - F_0$ ), the  $F_v$  (variable fluorescence) was measured in dark adapted leaves,  $F_m$   
140 (maximum fluorescence) was measured in dark adapted leaves after a pulse of saturating  
141 light, and  $F_0$  is the basal fluorescence produced by dark adapted leaves.

142

### 143 *Measurements of water potential*

144

145 The water potential was measured on leaves of three individuals of each treatment at 10:00  
146 am, the day before the beginning of the treatment with artificial fog (day 0), one day after the  
147 beginning of the treatment (day 1), and at day 4, day 8, day 18, and day 24, with a Scholander  
148 type pressure chamber (Scholander, 1964).

149

## 150 *Reflectance Measurements*

151

152 Spectral reflectance of leaves was measured in situ using a portable field spectroradiometer  
153 (GER1500, Geophysical & Environmental Research, Spectra Vista Corp., Poughkeepsie, NY,  
154 USA). Reflectance was calculated after standardization by canopy irradiance using a  
155 reference spectral panel (Spectralon, Labsphere, North Sutton, NH, USA) serving as a  
156 Lambertian reflector. All spectral measurements were from a nadir view angle approximately  
157 0.1 m above the plant. Three scans were quickly recorded (around one second per record) in  
158 different positions of each plant as replicates after measuring the white standard spectrum. The  
159 photochemical reflectance index (PRI) and Normalized Difference Vegetation Index (NDVI)  
160 were calculated from the reflectance data ( $R_x$  implies reflectance in  $x$  nm) respectively as  $PRI$   
161  $= (R_{531} - R_{570}) / (R_{531} + R_{570})$  (Gamon et al., 1992; Peñuelas et al., 1995) and  $NDVI =$   
162  $(R_{900} - R_{680}) / (R_{900} + R_{680})$  (Tucker, 1979).

163

## 164 *Remote sensing of Montseny Natural Park vegetation*

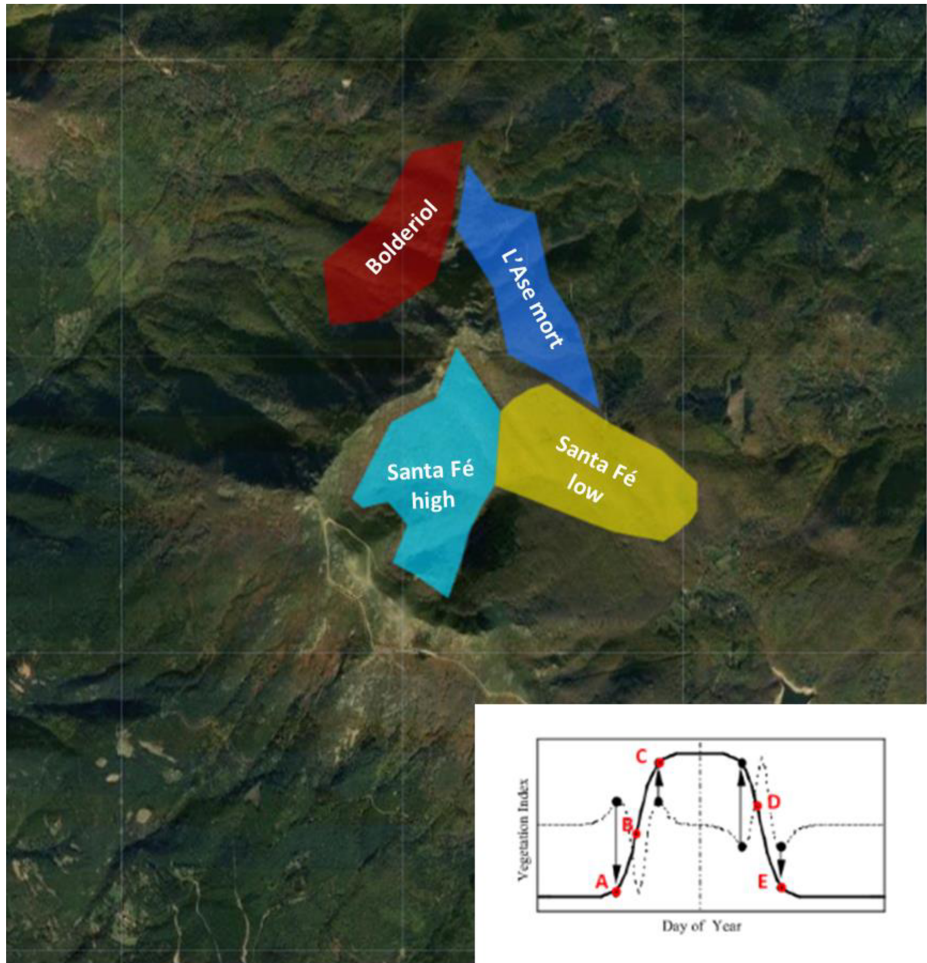
165

166 The annual cycle of *F. sylvatica* vegetation phenology was characterized using remote sensing  
167 using data from enhanced vegetation index (EVI) (Huete et., 2002) in four regions of  
168 Montseny Natural Park (Catalonia, Spain, northeastern Iberian Peninsula; 41°49'28''N,  
169 2°24'20''E). The regions were: Borderiol, l'Ase mort and Santa Fe with two altitudes (low  
170 and high altitudes) (Fig. 1). The data are from the period 2001 to 2014. Data based on MODIS  
171 - MCD12Q2 Land Cover Dynamics Product were obtained from the four moments that define  
172 the main phenological phases of vegetation dynamics at annual time scales (Zhang et al.,  
173 2003). The moments were: beginning of the photosynthetic activity; maturity, in which the

174 plant has its maximum leaf area; senescence, where photosynthetic activity and green leaf  
175 area begin to decrease; and dormancy, which is when the physiological activity is close to  
176 zero (Zhang et al., 2003). To verify leaf duration, the ratio between the period of senescence  
177 (D) and the maximum of the photosynthetic activity (C) was calculated.

178 To assess the direct relationship between the presence of fog in the green canopy duration from  
179 2001 to 2014, a linear regression was used. In order to make this regression, it was necessary to  
180 extrapolate the data of foggy days/year since the data obtained in the weather station captured data  
181 only in the period from 1950 to 2000. These data captured in these years were used to make a linear  
182 regression. From the generated equation, it was possible to estimate the data for the years 2001 to  
183 2014. Thus, time with the obtained data, another linear regression between the foggy days/year and the  
184 green canopy duration was performed.

185



186

187 **Fig. 1.** Aerial photograph of the studied sites in a Mediterranean-temperate forest. The colors  
 188 in the panel show the location of each site, which was used from remote sensing based on  
 189 MODIS - MCD12Q2 Land Cover Dynamics Product data for *Fagus sylvatica*. The lower  
 190 right panel shows the moments of the annual cycle vegetation phenology. Beginning of the  
 191 photosynthetic activity (A and B); maturity, in which the plant is with the maximum leaf area  
 192 (C); senescence (D), where photosynthetic activity and green leaf area begin to decrease; and  
 193 dormancy, which is when the physiological activity is close to zero (E)

194

195 *Data analyses*

196

197 General linear models (GLMs) with Gaussian errors were built to evaluate if potential  
198 quantum yield, Photochemical Reflectance Index (PRI), Normalized Difference Vegetation  
199 Index (NDVI), and water potential (dependent variables) are influenced for the different  
200 treatments (with artificial fog; soil irrigation; soil drought) (independent variables) over the  
201 two species. The analyses were performed in the R platform (R Development Core Team  
202 2015).

203

## 204 **Results**

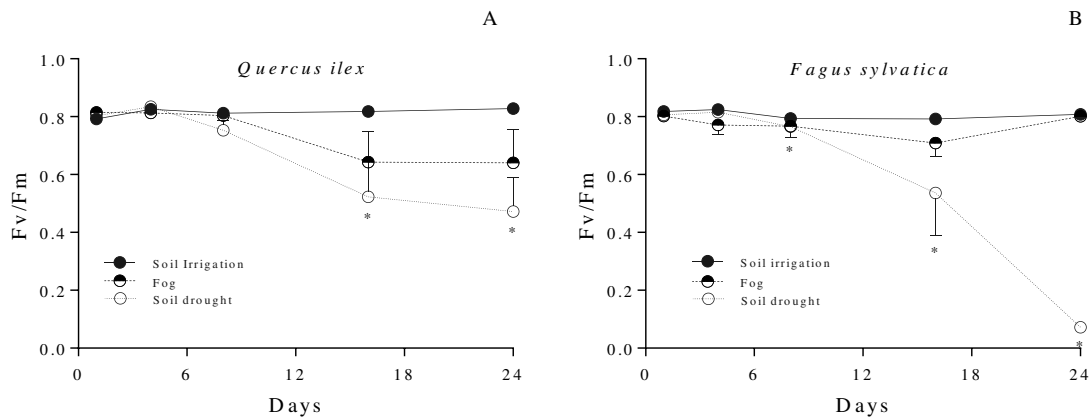
205

### 206 *Photoinhibition measurements*

207

208 The different performances under the three water regimes treatments in greenhouse for *F.*  
209 *sylvatica* and *Q. ilex*, had verified. The potential quantum yield ( $F_v / F_m$ ) of *Q. ilex* was not  
210 influenced by the fog, since the values were considered equal between the drought of the soil  
211 and the fog treatments and different from irrigation soil treatment ( $F_{2,87}=5.680$ ;  $P \leq 0.05$ ) (Fig.  
212 2A). Contrastingly, *F. sylvatica* showed was more responsive in the presence of fog, after  
213 all, there was no difference in the values of  $F_v / F_m$  between the soil irrigation and fog  
214 treatment but were different from soil drought treatment ( $F_{2,87}=7.853$ ;  $P \leq 0.05$ ) (Fig. 2B). The  
215 plants submitted to treatments with artificial fog and soil irrigation had the highest values of  
216 potential quantum yield when compared to the plants submitted to soil drought. There was a  
217 potential of quantum yield decreasing during the experiment with water stress, mainly in *F.*  
218 *sylvatica*, denoting an evident photoinhibition effect. Under water stress, the  $F_v/F_m$  values of  
219 *F. sylvatica* were lower than the values of *Q. ilex*, evidencing *F. sylvatica* higher sensitiveness  
220 to water deficit.

221



222

223 **Fig 2.** Measurements of PSII chlorophyll fluorescence parameters of seedlings submitted to  
224 soil irrigation, fog, and soil drought treatments. Potential quantum yield (Fv/Fm) of (A) *Q.*  
225 *ilex* and (B) *F. sylvatica*. Periods: before treatment with artificial fog (day 0), days 1, 4, 8, 16  
226 and 24. Mean values, and standard deviation, n = 6 species per treatment. General linear  
227 models (GLMs) with Gaussian errors,  $P < 0.05$ , followed by contrast analysis. Different letters  
228 denote significant difference among treatments.

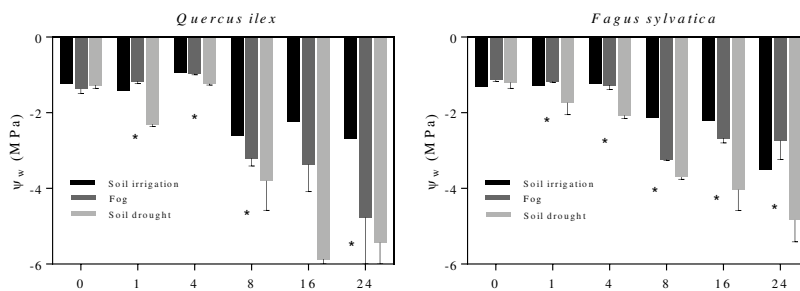
229

### 230 *Water potential*

231

232 Leaf water potential did not vary significantly between plants subjected to artificial fog  
233 and soil irrigation treatments but it was significantly lower in plants subjected to soil drought  
234 in both species ( $F_{2,105} = 24.845$ ;  $P \leq 0.05$ ) (Fig. 3). The artificial fog increased the mean values  
235 of leaf water potential both in *F. sylvatica* and *Q. ilex* generating water potential values  
236 similar to those of plants of soil irrigation treatment, mainly the first days. It is important to  
237 emphasize that at the beginning of the experiment, the water potential of the plants did not  
238 show significant difference between the different environmental conditions, demonstrating

239 that all plants presented the same water status ( $F_{5,100}=28.357$ ;  $P\leq 0.05$ ). Water from artificial fog  
 240 was enough to increase the water potential of these two species, improving their water status. This  
 241 increase in water potential demonstrates the strategy of these species to absorb water by the leaves  
 242 thus, the FWU capacity of both species is evident but possibly with different magnitudes.  
 243



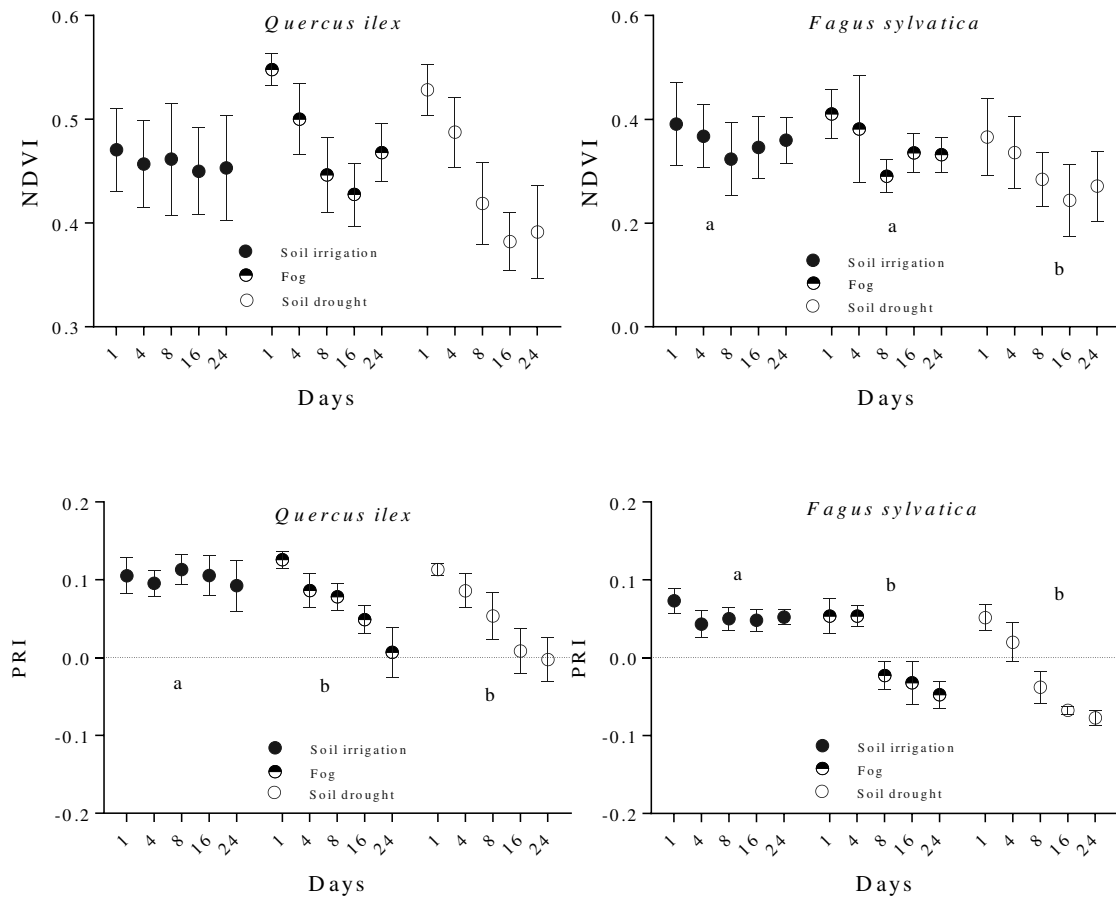
244 **Fig 3.** Leaf water

245 potential ( $\Psi_w$ ) of the branches of the two studied species along the three different treatments:  
 246 soil irrigation, fog, and soil drought. Periods: before treatment with fog (day 0), and 1, 4, 8, 16  
 247 and 24 days after to beginning of the treatments.

248  
 249 *Reflectance index*

250  
 251 The performance of the two studied species regarding the reflectance index is different. The  
 252 plants of *Q. ilex* showed no significant differences among the treatments for NDVI  
 253 ( $F_{2,87}=1.204$ ;  $P\geq 0.05$ ) (Fig. 4). However, there were differences for PRI ( $F_{2,87}=5.863$ ;  $P\leq 0.05$ )  
 254 between plants submitted to the soil irrigation treatment and those submitted of soil drought  
 255 and fog. For *F. sylvatica*, the NDVI values ( $F_{2,87}=7.498$ ;  $P\leq 0.05$ ) were different between  
 256 plants submitted to the water deficit treatment and the plants submitted to the treatments of  
 257 artificial fog and soil irrigation. Plants subjected to fog and the soil irrigation treatments had  
 258 higher values of NVDI than plants submitted to the soil drought treatment, mainly in the  
 259 initial days ( $F_{4,83}=6.744$ ;  $P\leq 0.05$ ). In contrast, the PRI values were similar between plants

260 subjected to the artificial fog and to the soil drought treatments, and PRI values were higher in  
 261 plants submitted to the soil irrigation treatment ( $F_{2,87}=21.123$ ;  $P\leq 0.05$ ). *Q. ilex* showed overall  
 262 higher values of NDVI and PRI than *F. sylvatica*.



263 **Fig. 4.** Variation of the efficiency of the use of radiation in *Q. ilex* and *F. sylvatica*. The  
 264 parameters used were NDVI (Normalized Difference Vegetation Index) and PRI  
 265 (Photochemical Reflectance Index). Measurements occurred in first day to start the treatment  
 266 with fog, 4, 8, 16 and 24 days. Error bars indicate the standard errors of the mean ( $n = 6$  for  
 267 each treatment). The significances of the GLM with *posthoc* test are represented using  
 268 different letters to show differences between treatments ( $P\leq 0.05$ ).  
 269

270

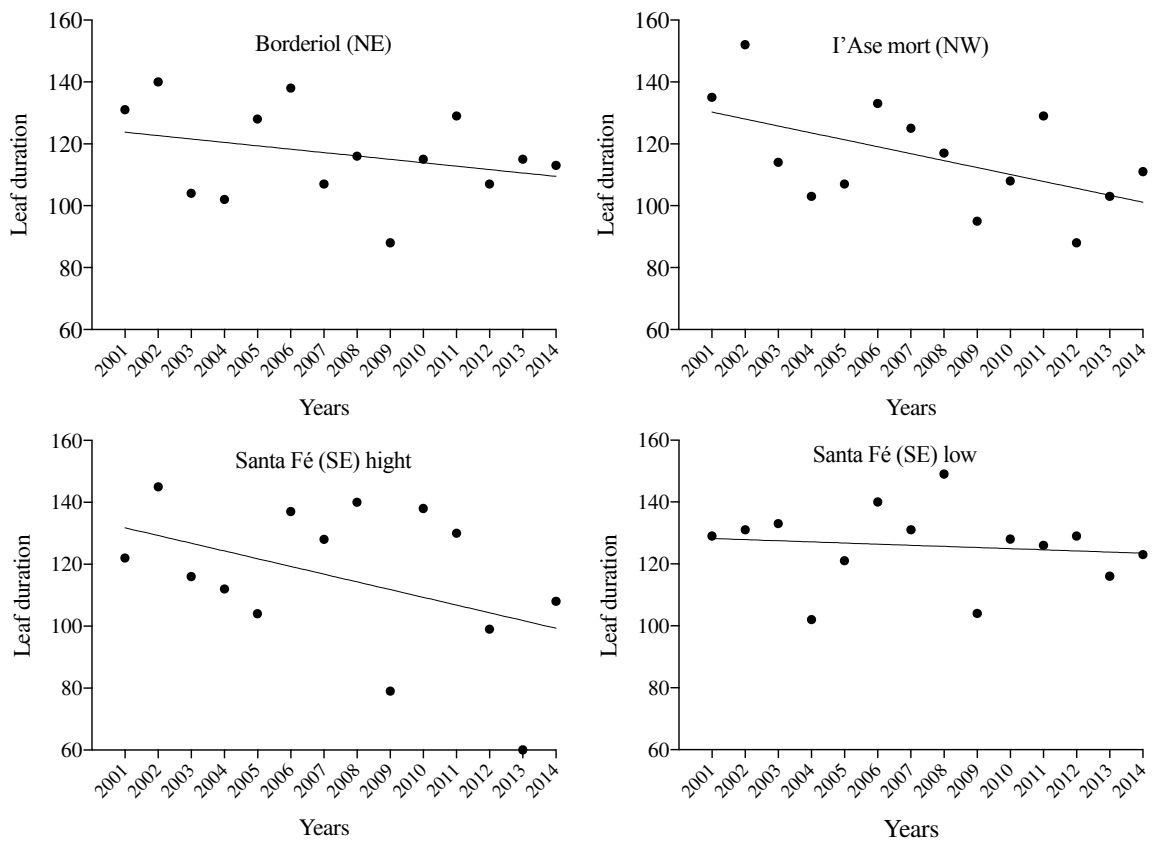
271 *Remote sensing - MODIS*

272

273 There was a trend to extend the duration of *F. sylvatica* leaves with the presence of fog. Thus,  
274 the decrease in fog events negatively interferes in leaf duration from the EVI-MODIS data,  
275 mainly in the high altitude of Santa Fe (SE) (Fig. 5).

276 Using data from 14 years of fog occurrence in the region, there was a direct relationship  
277 between the fog events and the duration of the leaves of *F. sylvatica*. There is a trend in  
278 which, with the increase in the foggy days, there was an increase in the duration of the leaves  
279 from canopy greenness duration (Fig. 6).

280

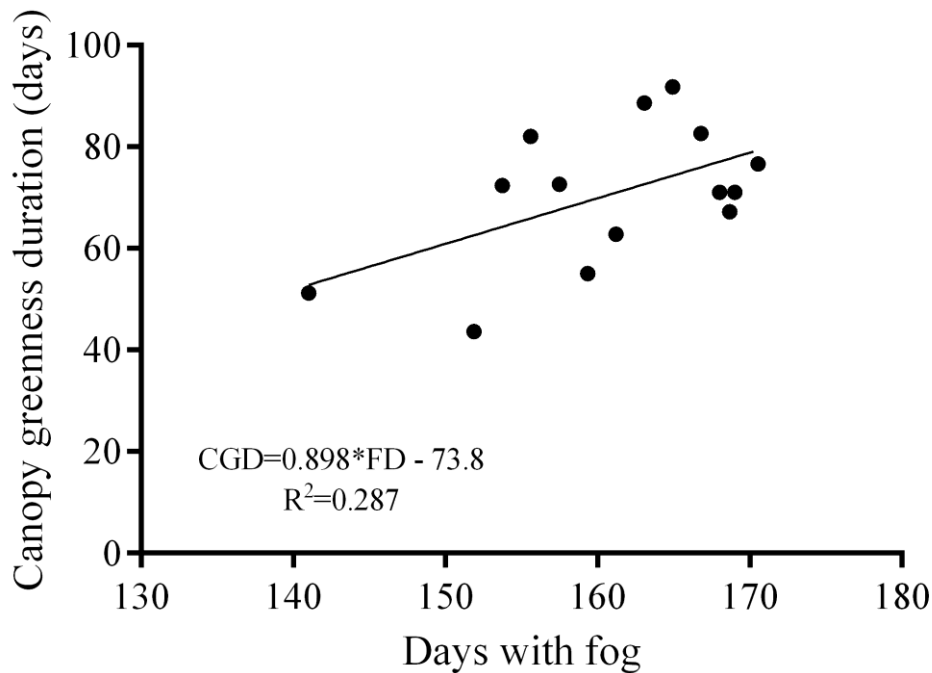


281

282 **Fig. 5.** Linear regression between the years and leaf duration from the product MCD12Q2 of

283 *F. sylvatica* in the three regions of Montseny Park. Linear regression: Bolderiol (NE): EVI=-

284 1.097\*years+2318; L'Ase mort (NW): EVI=-2.242\*years+4616; Santa Fe (SE) high altitude:  
285 EVI=-2.497\*years+5128 and Santa Fe (SE) low altitude: EVI=-0.3648\*years+8583.  
286



287  
288 **Fig. 6.** Linear regression between the days with fog and canopy greenness duration from the  
289 product MCD12Q2 of *F. sylvatica* in the Montseny Natural Park. Data collected from 14  
290 years (2001 -2014). Linear regression: CGD = canopy greenness duration and FD = fog days.  
291  $P < 0.05$ .

292  
293 **Discussion**

294  
295 Our physiological data complemented the understanding of the importance of fog in the  
296 Mediterranean environment for *F. sylvatica* and *Q. ilex*, corroborating that *F. sylvatica* is  
297 more responsive to fog than *Q. ilex* (Barbeta et al., 2019). Furthermore, we have verified, for

298 the first time, the role of fog in the phenology of a deciduous species. Its decrease negatively  
299 affected the duration of *F. sylvatica* leaves.

300 The fog, and its consequent absorption by the leaf, increased the water status of the plants  
301 and then interfered in photosynthetic efficiency. The FWU generated by the artificial fog was  
302 enough to keep the photosynthetic apparatus stability by increasing the potential quantum  
303 yield ( $F_v/F_m$ ) throughout the experiment and preventing possible photoinhibition effect for  
304 both species. Photoinhibition had already been demonstrated in *F. sylvatica* plants cultivated  
305 both under low and high light conditions (Tognetti et al., 1998), corroborating the importance  
306 of fog, since, besides providing an alternative water source for the plant, it also decreases  
307 radiation at the environment. Although *Q. ilex* showed no significant difference between  
308 treatments of drought and subjected to fog, artificial fog improved the photosynthetic  
309 efficiency not at the same intensity of the irrigated treatment, but efficient enough to avoid  
310 photoinhibition. Perhaps the non-significant difference between treatments is due to the  
311 higher tolerance of *Q. ilex* to photoinhibition (Méthy et al. 1996, Oliveira and Peñuelas 2000).  
312 Thus, the artificial fog was not enough to promote a statistical difference between the  
313 treatments of water deficit and fog.

314 The vegetation indices that also indicate the photosynthetic efficiency of the plants were  
315 influenced by the presence of artificial fog. The similarity between the vegetation status index  
316 for the NDVI in the irrigated treatment and in the artificial fog for *F. sylvatica* indicated the  
317 importance of fog for the maintenance of biomass-related biological variables such as leaf  
318 area index, chlorophyll by soil area, intercepted radiation fraction, leaf cover and green  
319 vegetation factor, which means the proportion of green vegetation in the canopy (Gamon et  
320 al., 1995; Rundquist 2002). The PRI index did not show a positive relationship in plants of *F.*  
321 *sylvatica* and *Q. ilex* subjected both to artificial fog and irrigated treatment. Accordingly,  
322 other leaf-scale studies have already seen reduced accuracy of PRI to track the photosynthetic

323 activity of herbaceous and shrubby plants under water deficit (Winkel et al., 2002; Filella et  
324 al., 2004). Then, the reduced accuracy of PRI may also have prevented the detection of fog  
325 influence on the hydration of *F. sylvatica* and *Q. ilex* because these plants were seedlings,  
326 similar sizes to herbaceous plants.

327 A more comprehensive view of the importance of fog in determining the landscape of an  
328 ecosystem was the verification of *F. sylvatica* phenology. The shorter duration of *F. sylvatica*  
329 leaves may be influenced by a decrease in fog events, as it is already proposed (Barbeta et al.,  
330 2019). This phenological change in response to warming processes is a sensitive and easily  
331 observable indicator of climate global changes (Parmesan and Yohe 2003; Root et al., 2003),  
332 which also interfere with the structure and functioning of the ecosystems (Peñuelas and Filella  
333 2001). It is well known that decreasing rainfall can change leaf phenology of drought-  
334 deciduous species, as rainfall may become insufficient to replenish soil water reserves  
335 regularly (Borchert 1998; Enquist and Enquist 2011). Although, the shortening of leaf  
336 duration as a consequence of low fog events is a slight trend for all the studied regions  
337 (Borderiol, l'Ase mort and Santa Fe with two altitudes), *F. sylvatica* behaves as an  
338 opportunistic species, because it maintains its leaves, and consequently, photosynthesis  
339 activity, while some type of water source for hydration is available. Thus, the presence of fog  
340 increase leaf duration, influence the phenology of *F. sylvatica*, by promoting instantaneous  
341 leaf hydration without the need for soil wetting.

342 Although both species have increased their water potential due to the presence of fog,  
343 there was a difference in behavior between the two species studied. This difference can be  
344 explained by the different phenological and physiological strategies of these species in  
345 relation to the availability of water. *F. sylvatica* is a deciduous species, that is, it has the  
346 strategy of avoiding the pressure imposed by water deficit by losing its leaves and limiting its  
347 physiological activity. On the other hand, *Q. ilex* is a perennial species, which has the strategy

348 of withstanding periods of low water availability without stress, allowing plant survival  
349 during the dry season (Murphy and Lugo 1986; Bowman and Prior 2005; Lock 2006;  
350 Lehmann et al., 2009). Thus, the higher sensitivity of *F. sylvatica* to fog is related to its  
351 phenology, since the presence of fog relieves the stress that the plant would suffer, delaying  
352 leaf falling. Since lower water availability and environmental warming are the two most  
353 important environmental and selective pressures that restrict *F. sylvatica* along its  
354 geographical range (Leuzinger et al., 2005), fog plays a key role in alleviating these  
355 environmental constraints. In fact, the presence of *F. sylvatica* at high altitudes is positively  
356 influenced by cloudiness during the growing season and may even have an impact on the  
357 growth of this species (Rozas et al., 2015; Barbeta et al., 2019).

358 The distinct habitat of *F. sylvatica* and *Q. ilex* may also explain the different responses of  
359 these species to fog. As an evergreen sclerophyll, *Q. ilex* occurs in lower altitude areas, which  
360 consequently have less fog events (Corell et al., 2014), and are often subjected to stressful  
361 environmental conditions including water deficit (Fleck et al., 1998). However, the  
362 relationship between the occurrence of *F. sylvatica* and fog events is traditionally known  
363 (Bolòs 1983; Gutiérrez 1988; Barbeta et al., 2019), which is the reason for the dominance of  
364 this species in higher altitude locations (Peters 1997; Ellenberg 2009), corroborating the view  
365 that *F. sylvatica* is a drought-sensitive tree species. This statement can be reinforced by  
366 observing that the relationship between leaf duration and the presence of fog was in the  
367 highest altitude region, demonstrating the selective pressure of fog in this ecosystem.

368 *Fagus sylvatica* has the behavior of an isohydric species (Arando 2018), that is, during the  
369 dry season this plant species can control the stomatal opening, under ideal or moderate  
370 drought conditions, and consequently prevent a xylem hydraulic conductivity failure (Tardieu  
371 and Simonneau 1998; Peuke et al., 2002). However, under severe drought, this response may  
372 be disadvantageous, for hydraulic system cavitation may occur (Wortemann et al., 2011;

373 Choat et al., 2012; Gleason et al., 2016). The risk that hydraulic irreversibility dysfunction  
374 may occur demonstrates that *F. sylvatica* cannot withstand such a severe drought (Choat et  
375 al., 2012; Gleason et al., 2016). Current data corroborate *F. sylvatica* as drought-sensitive  
376 species (Backes and Leuschner 2000; Aranda et al., 2005; Bréda et al., 2006; Ellenberg, 2009;  
377 Cano et al., 2013; Bahamonde et al., 2018). However, *F. sylvatica* has some relative high  
378 resilience and ability to recover from moderate water stress (Sánchez-Gómez et al., 2013;  
379 Blessing et al., 2016; Bahamonde et al., 2018). Perhaps this resilience can be enhanced by the  
380 presence of fog as an alternative source of water.

381 We conclude *F. sylvatica* and *Q. ilex* had different physiological responses regarding  
382 photoinhibition prevention, water potential increasing, and improvement in vegetative indices  
383 in the presence of fog. Unheard information that the reduction of fog events provides  
384 decreased leaf duration of a deciduous species, *F. sylvatica*, in this case, is of great  
385 importance for understanding the environment functioning. *F. sylvatica* main response to the  
386 fog events is a delay in its leaf falling, which improves its growth (Barbeta et al., 2019), and  
387 physiology and phenology. This information can be useful in predicting the possible behavior  
388 of *F. sylvatica* and *Q. ilex* in a less foggy environment due to global climate change. *F.*  
389 *sylvatica* will be the most negatively affected species in all physiological and phenological  
390 parameters analyzed with decreasing fog events, since it was the most responsive to the  
391 presence of this type of precipitation. Therefore, fog is extremely important for the eco-  
392 hydrology of the Mediterranean mountain, where *F. sylvatica* and *Q. ilex* are the main  
393 species.

394

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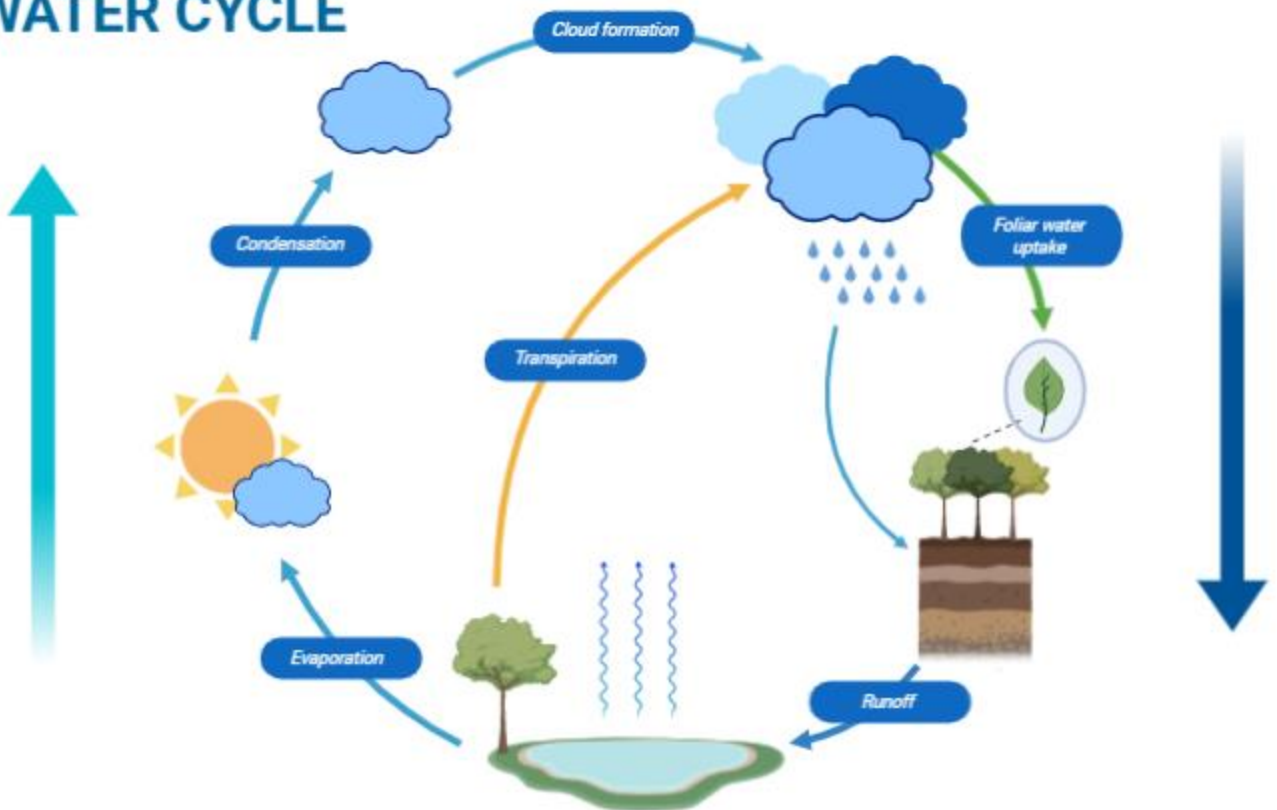
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*Towards a more comprehensive understanding of water flux in plants*

## COMPLETE WATER CYCLE



# 1 **Towards a more comprehensive understanding of water flux in plants**

2

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12

## 13 **ABSTRACT**

14

15 The reverse movement of water on the atmosphere - plant - soil continuum (APSC) driven by foliar water uptake has  
16 important consequences for plant performance and water flow in ecosystems. Here we discuss a novel perspective on  
17 the resistances that control the APSC in contrast to the soil-plant-atmosphere continuum (SPAC).

## 18 **MAIN TEXT**

19

20 Water movement through the soil-plant-atmosphere continuum (SPAC) has been traditionally seen as a  
21 unidirectional flux. Water flow starts from soil water absorption by plant roots through stems and leaves and diffuses  
22 to the atmosphere, in larger proportion, through transpiration, and to a lesser extent, by evaporation from leaf  
23 surfaces [1]. Water flow reversals induced by foliar water uptake (FWU) lead to an alternative water pathway [1] and  
24 here we propose to call this underappreciated pathway as atmosphere-plant-soil continuum (APSC). The  
25 bidirectional water movement is driven by differences in water potential among the atmosphere, leaves and soil as a  
26 consequence of the vapor pressure gradient generated among these compartments. This vapor gradient density  
27 between atmosphere and leaves is reduced, while relative air humidity between atmosphere and leaves is increased  
28 [1,2]. The objective of this commentary is to emphasize the differences between SPAC and APSC to highlight the  
29 importance of FWU as a major process driving water fluxes in ecosystems. Foliar water uptake starts when water  
30 overruns foliar surface due to vapor air saturation, resulting in full reversal in water potential. The reverse water flow

31 from the atmosphere into the plant leaf tissues requires water to overcome different hydraulic resistances, which are  
32 not exactly the same as those in the SPAC pathway.

33 The first barrier that water encounters is the foliar boundary layer [3], where vapor density tends to be high and  
34 vapor-pressure deficit consequently tends to be low. This gradient of water vapor accordingly hinders the entrance of  
35 water through the foliar boundary layer. The saturation vapor pressure inside and outside the leaves equalizes and  
36 favors water vapor condensation. Liquid water, when reaching the foliar surface, adheres to superficial structures,  
37 such as emergences, trichomes or wrinkles in the cuticle [4], then takes this pathway of lower resistance and enters  
38 the foliar tissues.

39 The foliar surface is thus the second barrier to FWU [5,6], where trichomes, emergences, cuticular and stomatal  
40 structures and even endophytic fungi may contribute to the hydration of the foliar surface. These structures decrease  
41 water surface tension and can be a lower resistance pathway for water influx and to restore leaf hydraulic  
42 conductance [7]. The chemical nature of the cuticle is important to determine leaf surface properties and, constitutes  
43 a barrier to water entrance into foliar tissues [8]. The chemical properties of this layer vary with plant species climate  
44 conditions, and foliar phenology, leading to distinct FWU competences [8], since different proportions in the  
45 hydrophobic and hydrophilic structural compounds can occur [9].

46 After the cuticular barrier has been crossed, water will always flow towards sites of less resistance down the  
47 water potential gradient, either through the apoplastic or the symplastic pathway [3]. To cross the intercellular spaces,  
48 water must overcome gas pressure and the resistance in the middle lamella and cell walls, where pectins and  
49 cellulose [10] influence the permeability of the cell wall and influence FWU [10]. A high proportion of pectins  
50 confers porosity to cell walls and favors rapid water absorption, and a high proportion of cellulose confers  
51 hygroscopicity to cell walls, and favors high water absorption. Both cell-wall components strongly influence FWU  
52 through the cell walls [10]. Water can also enter foliar palisade cells or cells of the spongy parenchyma through  
53 plasmodesmata and aquaporins, integral membrane proteins, in the plasma membrane [2]. Water conductivity  
54 through the plasma membrane can be increased by a higher expression of genes encoding proteins of the aquaporin  
55 channels [2]. The regulation of aquaporin genes influences foliar wetting, which together with high atmospheric  
56 humidity increases FWU [2]. After reaching the symplast, water can be retained in the mucilaginous content of cells  
57 or can diffuse throughout the foliar mesophyll [2] as well as go through paraveinal mesophyll, an important site to  
58 water redistribution and storage [11].

59 Lignified vascular bundle sheaths represent the next barrier before water gets into xylem vessels [3], which  
60 reduces hydraulic conductance in the vascular bundles [11]. Phloem cells are an alternative pathway for water to

61 reach the xylem vessels [2,12]. After water enters the vessels, hydraulic resistance may be influenced by the pits in  
62 the vessel walls parallel to the foliar surface, where occasional transpiration can induce an opposite water flow,  
63 increasing the resistance to water flow in APSS. Shoots and roots grow in opposite directions, so the water basipetal  
64 movement in the shoots, and its acropetal movement in the roots towards the root apices and soil, prevent  
65 transpiration. The soil-plant-atmosphere system flow can be completely reversed due to a favorable gradient of the  
66 soil-water potential attributed to the roots and transpiration suspension [1]. This reversed water flow has been  
67 observed in trunks and branches during precipitation and fog, in which the water absorbed by leaves reached the  
68 roots and occasionally the soil [1,13]. The resistance to water flow can increase if hydraulic conductivity varies due  
69 to the different calibers of the xylem vessels, which may imply an increase or decrease in water flow. In order for  
70 both the upward and downward movements start in roots and leaves, respectively, water flow will encounter  
71 resistance, which interferes with the entire water flow in bidirectional movements. Considering the downward  
72 movement, and all the resistances, especially that conferred by the cuticle, leaves impose the greatest resistance to  
73 FWU. However, water entrance through the leaves is not essentially the same as in the output movement. The  
74 cuticular pores and fissures, which are the main waterways for the APSC have much greater resistance compared to  
75 the ostiole and the boundary layer, when water vapor diffuses through the stomata in the SPAC. In contrast, for the  
76 upward flow, the greatest resistance to water flow from soil to the plant is found on the root endoderm, mainly due to  
77 the Casparian strips that force the water flow from the apoplastic to the symplastic pathway. Nevertheless, water  
78 absorption by the roots goes towards the driving forces of transpiration, driven by the established gradient of water  
79 potential, and propels the water into the atmosphere, causing the water upward flow to be greater than the downward  
80 movement.

81 About 9-70% of the water lost by transpiration can be recovered by FWU, depending on the species and  
82 environmental conditions [13-15]. FWU, although reduced for most species, plays a very important role in plant  
83 water balance, because it can rapidly increase water potential, reducing water deficit and supplying water to  
84 mesophyll cells or the vascular system [3,15]. The velocity of water absorbed, however, will depend on the traits of  
85 the foliar surface, mainly the composition and structure of the cuticle, which provide the major resistance to FWU.  
86 After this resistance is overcome, the ability of leaves to store water in their tissues can also interfere with the amount  
87 of water absorbed [4]. Most of the levels of hydraulic resistance observed during water flow from leaves to soil must  
88 overcome the biochemical composition of the leaves and the structural and anatomical traits of the entire plant body,  
89 i.e. leaves, stems and roots. These traits are genetically established but can also be strongly influenced by the  
90 environment, especially in plants with high phenotypic plasticity. Differences in environmental factors, such as solar

91 radiation and precipitation (fog, rain and dew), modify the efficiency of FWU due to changes in molecular,  
92 anatomical and physiological plant traits. Plant plasticity enables distinct anatomical strategies, such as greater  
93 thickness of foliar tissues and presence of cells with mucilage, which affect FWU capacity [4]. The different FWU  
94 strategies: quickly and lower water absorption or slower and higher water absorption were characterized in some  
95 plant species [4]. That happens as a response to water deficits in individuals of a given vegetation type related to  
96 individuals of the same species from a site with less evaporative demand [4,15]. The flow of water in the APSC in  
97 recent years it has received a lot of attention with the aim of complementing the SPAC, since this bidirectional  
98 movement is more common than imagined [16].. We thus recommend considering the importance of resistances  
99 imposed to water flow in multiple directions and also an integrative morphological, structural, biochemical and  
100 physiological approach to obtain a better understanding of the entire water flow in plants.

101

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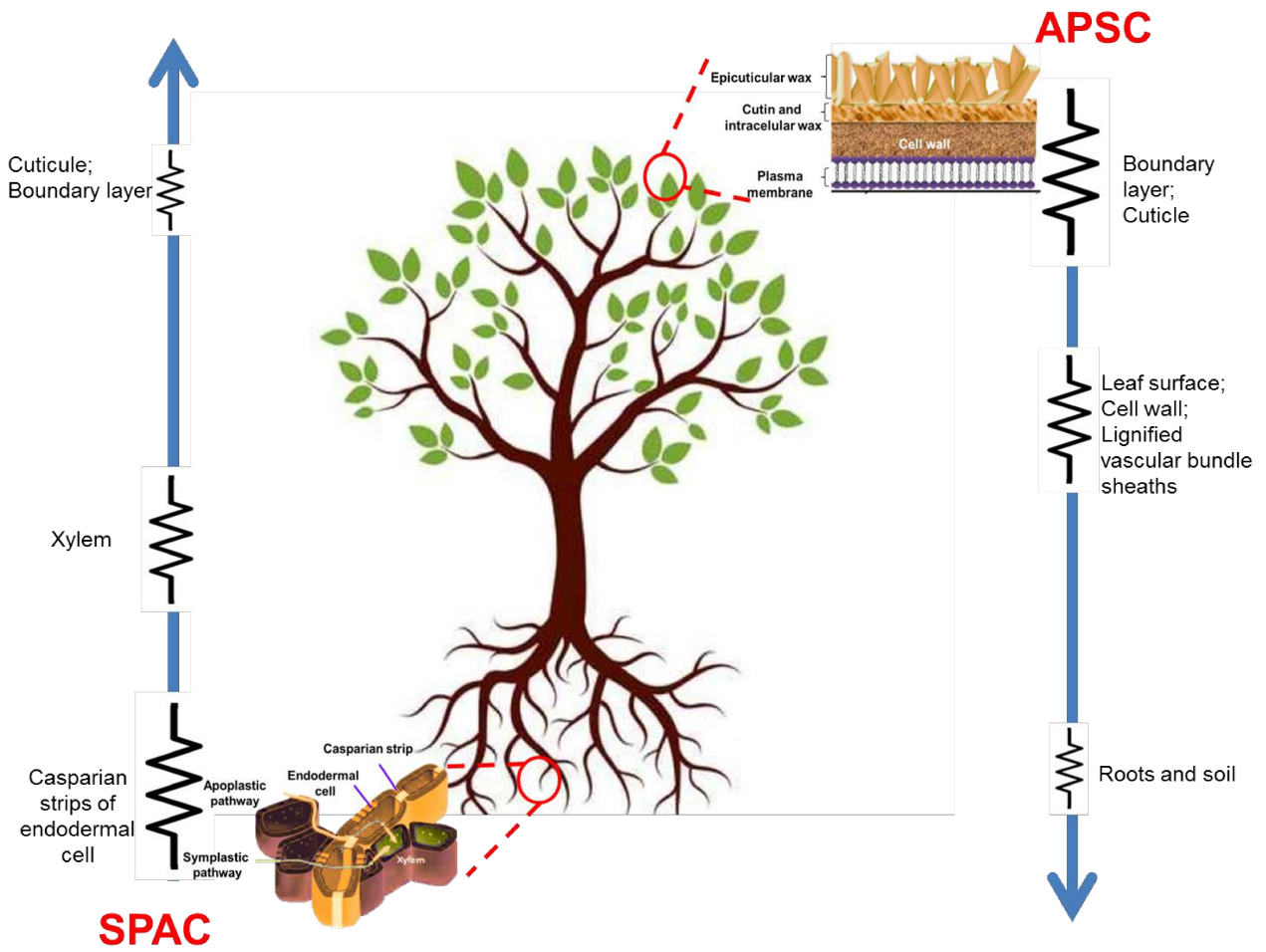
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138 **Figure caption**



139

140 Scheme of resistances to ascending (SPAC) soil-plant-atmosphere continuum) and descending (APSC, atmosphere-  
 141 plant-soil continuum) water flux. The size of resistance symbol represents the proportional resistances found  
 142 respectively in SPAC and APSS water flux.

## CONSIDERAÇÕES FINAIS

As plantas têm capacidades distintas de absorver água pelas folhas (foliar water uptake – FWU) e essa característica decorre de atributos particulares de cunho morfológico, anatômico e ultraestrutural. Esses atributos, conseqüentemente, contribuem para as diferentes respostas ecofisiológicas, de caráter bioquímico e fenológico. A possibilidade de absorver água pelas folhas pode influenciar na sobrevivência de espécies de um determinado ambiente, principalmente mediante as mudanças no regime de precipitação preditas nos modelos climáticos globais. Utilizando dois ecossistemas (Campo rupestre ferruginoso e ecótono-mediterrâneo-temperado) e também sete espécies com diferentes capacidades de absorção foliar de água podemos verificar a influência da FWU em todos os atributos descritos acima e a importância da neblina para a ocorrência destas espécies nestes dois ambientes.

Quanto à ultraestrutura celular, foi mostrado que existe relação entre as ornamentações de cera epicuticular, composição química da cera cuticular e da parede celular com as diferentes capacidades de FWU. Verificou-se que a proporção dos componentes da cera e da parede celular interfere nas diferentes estratégias de absorver água rápido, porém em menor quantidade ou mais lentamente, mas em maior quantidade. Como perspectivas, o isolamento da cera cuticular para análises mais aprofundadas da permeabilidade cuticular permitirá o maior entendimento do fluxo reverso de água na epiderme das folhas.

Embora a FWU tenha interferido negativamente na assimilação de CO<sub>2</sub>, essa interferência foi breve e evitou a perda de água através da transpiração, aumentando o status hídrico da planta na estação seca, onde a única forma de precipitação é a neblina. A neblina demonstrou ter um papel fundamental na melhoria do status hídrico das espécies na estação chuvosa e principalmente na estação seca na presença de neblina.

Verificou-se que entre as estratégias de FWU, as espécies que absorvem lentamente uma quantidade maior de água (*T. laniflora*, *P.heteromallum*, *M. corallina*, e *E. erythropappus*) tiveram melhor desempenho fotossintético com maior aumento da termotolerância e diminuição

e/ou desaparecimento da fotoinibição comparado com as espécies que absorvem água mais rapidamente, mas em menor quantidade (*O. pulchella*, *B. variabilis* e *L. australis*). O metabolismo oxidativo também foi associado às estratégias de FWU. As espécies que apresentam a estratégia de maior velocidade de absorção foram também as mais sensíveis, já que houve aumento de espécies reativas ao oxigênio comparado com aquelas que possuem a estratégia de absorver mais lentamente maior quantidade de água.

Já foi demonstrado indiretamente em alguns destes parâmetros (trocas gasosas, eficiência fotossintética, termotolerância e metabolismo oxidativo) e diretamente através de medidas de potencial hídrico das plantas na estação seca com neblina, a importância da neblina no campo rupestre ferruginoso. Assim, visando avaliar a importância da neblina em outro ecossistema, foram utilizadas duas espécies do ecótono mediterrânico e temperado, visto que pesquisas da Universidade Autônoma de Barcelona, verificaram diferentes respostas no crescimento dessas espécies em relação à neblina. Como esperado, foram mostrados respostas fisiológicas diferentes em relação à prevenção de fotoinibição e melhora dos índices vegetativos na presença de neblina. Além disso, foi visto a interferência da neblina na fenologia de *Fagus sylvatica* até então nunca verificado, ou seja, a manutenção das folhas de uma espécie decídua em condições de estresse hídrico minimizado pela ocorrência de neblina. Com a diminuição nos eventos de neblina, ocorre uma aceleração da queda foliar. Para finalizar, compilamos todas as principais vias de entrada de água a partir das folhas para aprimorar o modelo o ciclo da água no sistema atmosfera-planta-solo.

Em conclusão, as plantas que apresentaram a estratégia de absorver lentamente maior quantidade de água tiveram maior sucesso nos parâmetros fotossintéticos, na termotolerância e no metabolismo oxidativo comparado com as espécies que absorvem água mais rapidamente mas em menor quantidade. Assim, o conhecimento de como cada estratégia interfere nestes atributos ajudará na escolha de espécies mais adaptadas aos futuros cenários climáticos para a revegetação destes ecossistemas, e também contribuirá na previsão do cenário das possíveis mudanças na paisagem e quais as espécies serão mais susceptíveis ou tolerantes a essas

mudanças. Foi demonstrado que a neblina tem um papel importante para todas as espécies estudadas nos distintos ecossistemas (*T. laniflora*, *P.heteromallum*, *M. corallina*, *E. erythropappus*, *O. pulchella*, *B. variabilis*, *L. australis*, *F. sylvatica* e *Q. ilex*), sendo um fenômeno meteorológico importante e necessário para a sobrevivência destas plantas.

## EPÍLOGO

*Voar com asa ferida?*

*Abram alas quando eu falo.*

*Fiz, pequeno, quando o tempo estava todo do meu lado e o que se chama passado,  
passatempo, pesadelo só me existia nos livros.*

*Feliz, depois, dono de mim, quando tiver que escolher entre um abismo, o começo, e  
essa história sem fim.*

*Asa ferida, asa ferida,*

*Meu espaço, meu herói. A asa arde. Voar, isso não dói.*

Paulo Leminski

*A maior riqueza do homem é sua incompletude.*

*Nesse ponto sou abastado.*

*Palavras que me aceitam como sou*

*— eu não aceito.*

*Não aguento ser apenas um sujeito que abre  
portas, que puxa válvulas, que olha o  
relógio, que compra pão às 6 da tarde, que vai  
lá fora, que aponta lápis, que vê a uva etc. etc.*

*Perdoai.*

*Mas eu preciso ser Outros.*

*Eu penso renovar o homem  
usando borboletas.*

Manoel de Barros

E valeu a pena!!!