

LAUREN FRENZEL SCHUCH

**EXPRESSÃO DAS PROTEÍNAS CHK2, γ H2AX E TP53 EM
CARCINOMA DE CÉLULAS ESCAMOSAS DE BOCA DE
INDIVÍDUOS FUMANTES E NÃO FUMANTES**

**Faculdade de Odontologia
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ATA DA DEFESA DE DISSERTAÇÃO DA ALUNA LAUREN FRENZEL SCHUCH

Aos 20 dias de novembro de 2019, às 14:00 horas, na sala 3403 da Faculdade de Odontologia da Universidade Federal de Minas Gerais, reuniu-se a Comissão Examinadora composta pelos professores Maria Cassia Ferreira de Aguiar (Orientadora) – FO/UFMG, Silvia Ferreira de Sousa – FO/UFMG, Manoela Domingues Martins - UFGRS e Vanessa de Fátima Bernardes – ICB/UFMG, para julgamento da dissertação de Mestrado, área de concentração em Estomatologia, intitulada: **Expressão das proteínas chk2, h2ax e p53 em carcinoma de células escamosas de boca de indivíduos fumantes e não fumantes**. A Presidente da Banca, abriu os trabalhos e apresentou a Comissão Examinadora. Após a exposição oral do trabalho pela aluna e arguição pelos membros da banca, a Comissão Examinadora considerou a dissertação:

Aprovada

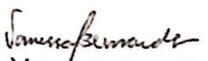
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Finalizados os trabalhos, lavrou-se a presente ata que, lida e aprovada, vai assinada por mim e pelos demais membros da Comissão. Belo Horizonte, 20 dias de novembro de 2019.


Prof(a). Maria Cassia Ferreira de Aguiar


Prof(a). Silvia Ferreira de Sousa


Prof(a). Manoela Domingues Martins


Prof(a). Vanessa de Fátima Bernardes

Lauren Frenzel Schuch

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Orientadora: Profa. Dra. Maria Cássia Ferreira de Aguiar

Coorientadora: Profa. Dra. Vanessa de Fátima Bernardes

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Lou Andreas-Salomé

RESUMO

O câncer de boca corresponde a cerca de 4% das doenças neoplásicas, e o carcinoma de células escamosas representa o tipo mais frequente, englobando cerca de 90 a 95% dos casos. O cigarro é o principal fator etiológico para o câncer de boca, causando danos ao DNA e mutações que se não reparadas levam ao surgimento de lesões. O dano ao DNA associado ao cigarro tem sido estudado em diferentes tipos de câncer, mas ainda pouco explorado em relação ao câncer de boca. *Checkpoint kinase 2* (CHK2) e P53 são proteínas que estão envolvidas no processo de checagem do ciclo celular, sendo responsáveis pelo reparo ao dano ao DNA. A proteína H2AX é uma histona nuclear que sofre fosforilação em resposta aos danos ao DNA, principalmente às quebras da dupla cadeia. O objetivo deste estudo foi avaliar a resposta a danos no DNA através da expressão de CHK2, γ H2AX e TP53 entre fumantes e não fumantes com carcinoma de células escamosas de boca (CCEB). Além disso, foram analisadas associações entre imuno-expressão das proteínas estudadas, dados clínico-patológicos e classificação histopatológica. Foram incluídos 35 indivíduos (18 não fumantes e 17 fumantes) com CCEB de língua e soalho bucal. Reação imuno-histoquímica foi realizada para γ H2AX para identificação de quebras de fita dupla, CHK2 e TP53 para avaliação da indução de parada do ciclo celular. Análises descritivas e estatísticas foram realizadas. A pesquisa foi composta por 22 homens (62,8%) e 13 mulheres (37,2%), com idade média de 63,9 anos. Entre os não fumantes, 50% apresentaram tumores bem diferenciados, enquanto que fumantes mostraram maior número em moderadamente diferenciados e pouco diferenciados (35,3% cada). No geral, 31 (88,6%) casos foram positivos para CHK2, 27 (77,2%) foram positivos para γ H2AX e 23 (65,7%) foram positivos para TP53. Não foi observada associação entre essas proteínas com hábitos de fumar e não fumar ($p > 0,05$). Semelhanças entre os padrões imuno-histoquímicos de CHK2, γ H2AX e TP53 em fumantes e não fumantes com CCEB foram observadas neste estudo, assim como entre os parâmetros clínico-patológicos. De forma geral, os resultados indicaram expressão positiva para essas proteínas no CCEB. Este estudo fornece informações sobre o dano ao DNA na carcinogênese oral.

Palavras-chave: Dano ao DNA. CHK2. Gama-H2AX. P53. Câncer oral. Tabaco.

ABSTRACT

Expression of CHK2, γ H2AX and TP53 proteins in smokers and non-smokers oral squamous cell carcinoma

Oral cancer accounts for about 4% of neoplastic diseases, and squamous cell carcinoma is the most common type, accounting for about 90 to 95% of cases. Cigarette smoking is the main etiological factor for oral cancer, causing DNA damage and mutations that, if not repaired, lead to lesions. Cigarette-associated DNA damage has been studied in different cancers, but is still poorly explored in relation to oral cancer. Checkpoint kinase 2 (CHK2) and P53 are proteins that are involved in the cell cycle checking process and are responsible for repairing DNA damage. The H2AX protein is a nuclear histone that undergoes phosphorylation in response to DNA damage, especially double strand breaks. The aim of this study was to assess the DNA damage response through the expression of checkpoint kinase 2 (CHK2), γ H2A histone family member X (γ H2AX) and TP53 among smokers and non-smokers with oral squamous cell carcinoma (OSCC). In addition, associations amongst immunoexpression of studied proteins, clinicopathologic data and histopathological grading were analyzed. Thirty-five individuals (18 non-smokers and 17 smokers) with OSCC of the tongue and/or floor of the mouth were included. Immunohistochemistry was carried out for γ H2AX for identification of double-strand breaks, CHK2 and P53 for evaluation of the induction of cell cycle arrest. Descriptive and statistical analyses were performed. The survey consisted of 22 males (62.8%) and 13 females (37.2%), with a mean age of 63.9 years. Fifty percent of non-smokers OSCC were well-differentiated tumors, whereas for smokers, OSCC were moderately differentiated and poorly differentiate tumors, equally (35.3% each). Overall, 31 (88.6%) cases were CHK2-positive, 27 (77.2%) were γ H2AX-positive and 23 (65.7%) were TP53-positive. No association among these proteins with smoking and non-smoking habits was observed ($p>0.05$). Similarities in the CHK2, γ H2AX and P53 immunohistochemical staining pattern were observed between smokers and non-smokers with OSCC in this survey, and the immunoexpression was not associated with clinicopathologic parameters. Overall, the results indicated consistent expression of these proteins in OSCC. This study provides information about the DNA damage in oral carcinogenesis.

Keywords: DNA damage. Checkpoint Kinase 2. H2AX protein. Gamma-H2AX protein. Tumor suppressor protein P53. Oral cancer. Smoking.

LISTA DE ABREVIATURA E SIGLAS

CCEB Carcinoma de Células Escamosas de Boca

CHK2 *Checkpoint Kinase 2*

FAO Faculdade de Odontologia

H2AX *Histona Family Member*

IHQ Imuno-histoquímica

INCA Instituto Nacional de Câncer

OMS Organização Mundial da Saúde

OSCC Oral Squamous Cell Carcinoma

SPSS *Statistical Package for the Social Sciences*

UFMG Universidade Federal de Minas Gerais

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1 CONSIDERAÇÕES INICIAIS

O carcinoma de células escamosas de boca (CCEB) possui uma incidência anual de 300.000 casos e está entre as dez malignidades mais comuns do mundo, correspondendo a 90% dos cânceres orais (ALI *et al.*, 2017; CUTILLI *et al.*, 2016; FERLAY *et al.*, 2015). Ainda, as estimativas indicam um aumento global significativo para 19,3 milhões de novos casos anuais até 2025 (D'SOUZA e SARANATH, 2015). Embora avanços recentes relacionados à terapia tenham sido relatados, a resposta à conduta terapêutica, altas taxas de recidivas e a permanência de baixa taxa de sobrevida em 5 anos para alguns pacientes ainda acarretam preocupação aos especialistas (CUTILLI *et al.*, 2016; LEE *et al.*, 2015).

O CCEB representa uma neoplasia maligna de origem epitelial caracterizado histologicamente por uma intensa proliferação de células epiteliais neoplásicas que invadem a lâmina própria subjacente formando ilhas ou cordões. Essas células são caracterizadas por apresentarem pleomorfismo celular e nuclear, hipercromatismo nuclear, nucléolos evidentes, figuras de mitoses atípicas e aumento da relação núcleo-citoplasma. Classificações histopatológicas para os CCEBs surgiram na tentativa de explicar o comportamento biológico discrepante dos tumores. Broders, em 1920, propôs uma gradação histopatológica baseada no grau de diferenciação celular. Contudo, muitos autores questionam o valor dessa classificação e o da proposta pela Organização Mundial de Saúde (OMS), destacando o papel de outras características histopatológicas no comportamento biológico do tumor e propondo novas gradações (ANNEROTH *et al.*, 1986; BARNES *et al.*, 2005; LINDEMANN *et al.*, 2018; WAGNER *et al.*, 2017).

O desenvolvimento de CCEB ocorre através de um processo multifatorial que requer o acúmulo de múltiplas alterações, influenciado tanto pela influência genética do paciente, quanto por fatores ambientais - tabaco, álcool, inflamação crônica e infecção viral (APPAH *et al.*, 2018; SCULLY e BAGAN, 2009; TSANTOULIS *et al.*, 2007). O cigarro continua a ser o principal fator etiológico do câncer de boca e o mecanismo de ação envolve danos diretos ao DNA. O tabaco pode causar alteração epigenética das células epiteliais orais, inibir múltiplas funções imunológicas sistêmicas do hospedeiro e, por meio de seus metabólitos tóxicos, causar estresse oxidativo nos tecidos para induzir o CCEB (JIANG *et al.*,

2019). Nos últimos anos, foi possível um entendimento maior dos componentes cancerígenos contidos nos produtos de tabaco, bem como seu papel na geração de tumores específicos (JETHWA e KHARIWALA, 2017).

Diversas alterações genéticas ocorrem tanto em células normais quanto em células displásicas, incluindo mutações pontuais, deleções, translocações, amplificações, metilações, instabilidade de microssatélites e perda de heterozigosidade (SINEVICI *et al.*, 2016). A superexpressão de genes que promovem crescimento, sobrevivência e disseminação de células pode levar ao desenvolvimento de câncer (SCULLY e BAGAN, 2009). No entanto, a identificação dessa superexpressão de oncogenes representa um desafio. Diante disso, uma demanda por biomarcadores clinicamente confiáveis levou a um aumento da produção científica na pesquisa de possíveis alvos em níveis genéticos, mRNA, proteicos e metabólicos, sendo a grande maioria intrinsecamente ligada ao desenvolvimento de células neoplásicas malignas (D'SOUZA e SARANATH, 2015; SINEVICI *et al.*, 2016).

O ciclo celular, regulado e monitorado por pontos de checagem, é caracterizado por uma sequência de eventos que ocorrem em uma célula resultando na sua replicação ou divisão (ABREU E HOWARD, 2015; ALI *et al.*, 2017). Esses pontos de verificação são importantes pois fornecem uma parada temporária controlada em um estágio específico do ciclo que permite que a célula corrija possíveis defeitos (TAO *et al.*, 2009). Danos ao DNA ocorrem de forma corriqueira, tanto por causas exógenas quanto endógenas, desencadeando respostas que combinam o reparo do DNA com a parada temporária do ciclo celular, evitando efeitos deletérios ou morte celular (DAVIDSON *et al.*, 2018).

A tirosina quinase CHK2 tem papel importante nesse metabolismo, uma vez que está envolvida na via de reparo ao dano do DNA, capaz de ativar o P53, modular a reparação e bloquear o ciclo celular (CARRASSA & DAMIA, 2011; GUFFANTI *et al.*, 2016). A primeira evidência de que a alteração genética em CHK2 pode predispor ao câncer foi relatada por Bell e colaboradores (1999), em mutações germinativas raras no gene em famílias com síndrome de Li-Fraumeni, caracterizada por múltiplos tumores em idade precoce, com predomínio de câncer de mama e sarcomas, e está frequentemente associada a mutações germinativas no gene P53. CHK1 e CHK2 regulam funções celulares fundamentais, como replicação do DNA e progressão do ciclo celular, reestruturação da cromatina e

apoptose, representando dois mensageiros críticos dos pontos de verificação da integridade do genoma, além do seu envolvimento na evolução do câncer humano (BARTEK & LUKAS, 2003). Na ausência de P53 funcional, a interrupção do ciclo celular e o reparo do DNA dependem da função do CHK1/2 para o reparo e replicação do DNA funcional (LINDEMANN *et al.*, 2018). Enquanto que a literatura defende que o CHK1 promove o reparo tumoral, sendo altamente encontrado numa variedade de tumores humanos, incluindo câncer de mama, cólon, fígado, gástrico e carcinoma nasofaríngeo, poucos estudos tem pesquisado sobre o desfecho do CHK2 em neoplasias de cabeça e pescoço (ZHANG & HUNTER, 2013).

A histona variante H2AX pertence à família histona H2A e, como outras variantes histonas, é altamente conservada e desempenha funções celulares críticas (WEYEMI *et al.*, 2018). O gene H2AX possui um papel essencial no reparo da quebra da fita dupla do DNA e na estabilidade do genoma, sendo considerado um gene supressor de tumor (MEADOR *et al.*, 2008; WEYEMI *et al.*, 2016). Em um estudo realizado por Osterman e colaboradores (2014), ao investigar a ligação entre o dano ao DNA e o câncer de pâncreas, ficou bem estabelecido que a H2AX desempenha um papel importante na disseminação do sinal ao dano, e que a H2AX fosforilada (γ H2AX) é necessária para a estabilização de numerosos fatores de resposta ao dano nas lesões do DNA. Assim, observou-se ausência de expressão de γ H2AX nas amostras de tecido pancreático normal, enquanto a expressão de γ H2AX foi altamente positiva tanto no ducto quanto no tecido ao seu redor na amostra do tumor. Poucos estudos tem relatado CCEB e H2AX. Chou e colaboradores, em 2011, demonstraram a proteína em amostras de CCEB, que foi considerada um importante biomarcador de resposta ao dano ao DNA em estágio inicial da carcinogênese.

O gene supressor de tumor TP53, essencial na regulação da progressão do ciclo celular, diferenciação e reparo de DNA, é um gene bem documentado na literatura por estar associado não só ao carcinoma de boca, mas também a vários outros tipos de câncer (WHYTE *et al.*, 2002; SINEVICI, 2016). Cerca de 50% dos pacientes diagnosticados com CCEB possuem mutação no gene TP53 (SINEVICI, 2016). A ativação do TP53 é decorrência do estresse celular, o que inclui dano ao DNA, hipóxia e privação nucleotídea (LINDEMANN *et al.*, 2018). Tal proteína desempenha papel crítico para reprimir a invasão do câncer e a progressão

metastática (SINEVICI *et al.*, 2016; YANG *et al.*, 2015; ZEDAN *et al.*, 2015). Estudos na literatura mostraram que os CCEBs com superexpressão ou mutação de P53 proliferam mais rapidamente e possuem natureza mais agressiva, bem como um mau prognóstico (LEE *et al.*, 2015). Corroborando tal informação, Cutilli e colaboradores (2016), em um estudo avaliando a relação do P53 com o CCEB, observaram que os tumores com superexpressão de TP53 maior que 50% apresentaram uma taxa de sobrevida, em um período de 24 meses, de 47,4%, enquanto que tumores com expressão inferior a 50% tiveram uma taxa de sobrevida de 80,8%.

Embora a associação de TP53 esteja bem estabelecida quando relacionada ao CCEB, as proteínas CHK2 e H2AX ainda foram pouco exploradas no que se refere à expressão destas no câncer oral. A avaliação de possíveis danos que podem causar ao DNA é importante para a melhor compreensão da patogênese do CCEB associada ao tabaco, com impactos no estabelecimento de medidas preventivas e terapêuticas. Diante disso, o objetivo deste estudo foi avaliar a expressão imuno-histoquímica (IHQ) das proteínas CHK2, H2AX e P53, associadas ao processo de proteção e reparo ao DNA, comparando a expressão destas em carcinomas de indivíduos fumantes e não fumantes e verificando se há associação entre as mesmas, além de investigar a associação com características clínicas e histopatológicas da lesão.

1.1 Objetivos da pesquisa

1.1.1 Objetivos gerais

O objetivo deste trabalho foi avaliar a expressão imuno-histoquímica das proteínas CHK2, γ H2AX e TP53 em carcinoma de células escamosas de boca de indivíduos fumantes e não fumantes.

1.1.2 Objetivos específicos

- a) Investigar a expressão imuno-histoquímica das proteínas CHK2, γ H2AX e TP53 em amostras de CCEB de indivíduos fumantes e não fumantes;
- b) Verificar se existe associação entre a expressão destas proteínas;

c) Verificar por meio de análise estatística se algum fator clínico (idade, sexo, fumo e informações da lesão) ou histopatológico (grau de diferenciação) está associado à expressão destas proteínas.

2 METODOLOGIA EXPANDIDA

2.1 Aspectos éticos

O presente trabalho foi aprovado pelo Comitê de Ética em Pesquisa da Universidade Federal de Minas Gerais (COEP/UFMG), sob o número CAAE: 03012618.1.0000.5149; parecer: 3.293.055 (Anexo A), obedecendo ao exigido pela legislação brasileira, conforme as resoluções CNS nº 466/12 e 340/04 do Conselho Nacional de Saúde, sobre Diretrizes e Normas Regulamentadoras de Pesquisas Envolvendo Seres Humanos.

2.2 Delineamento do estudo

Trata-se de um estudo transversal, do tipo retrospectivo, com população-alvo constituída por indivíduos com diagnóstico histopatológico de carcinoma de células escamosas de boca cujo material estava armazenado no arquivo do laboratório de Patologia Bucomaxilofacial da Faculdade de Odontologia da Universidade Federal de Minas Gerais.

2.3 Amostra

Inicialmente, foi conduzido um estudo piloto para determinação do cálculo amostral por meio de teste estatístico, através do programa OpenEpi (versão 3.01) (DEAN et al., 2013). No entanto, não foi possível a utilização da amostra estipulada e partiu-se para uma amostra de conveniência. O estudo foi composto apenas por casos de CCEB, através de um cálculo amostral, que tiveram laudo histopatológico pela FAO-UFMG. Os pacientes foram divididos em dois grupos: fumantes e não fumantes. Foram considerados não fumantes os indivíduos que nunca fumaram ou que fumaram até 0,5 maço/ano nos últimos 10 anos (PAULIN *et al.*, 2015). Como fumantes, aqueles que usam cigarro de forma contínua ou esporádica.

2.4 Coleta de dados

Foram coletados, da ficha de biópsia, os seguintes dados: sexo, idade, hábitos de fumo e álcool, sintomatologia, localização anatômica e tipo de amostra (incisional ou excisional). Ainda, foi feita uma análise histológica para determinar o grau do tumor, de acordo com a Organização Mundial da Saúde (OMS) 2017 (EL-NAGGAR *et al.*, 2017), em bem diferenciado, moderadamente diferenciado e mal diferenciado (QUADRO 1).

Quadro 1 - Sistema de gradação histológica para carcinoma de células escamosas de boca proposto pela OMS (2017)

Parâmetros	Características
Bem diferenciado	Arquitetura tecidual semelhante ao padrão normal do epitélio escamoso
Moderadamente diferenciado	Certo grau de pleomorfismo nuclear e atividade mitótica; Pouca ceratinização
Pouco diferenciado	Predomínio de células imaturas; Numerosas mitoses típicas e atípicas; Mínima ceratinização

Fonte: EL-NAGGAR *et al.*, 2017.

2.5 Análise imuno-histoquímica

Os cortes histológicos apresentaram espessura de 4 μm e foram montados em lâminas de vidro StarFrost® (Waldemar Knittel Glasbearbeitungs GmbH, Braunschweig, Alemanha). Foi realizada a avaliação IHQ das proteínas CHK2, H2AX e P53, e os anticorpos estão descritos na Tabela 1.

Tabela 1 - Anticorpos primários, fabricantes, soluções para recuperação antigênica, concentrações e sistema de detecção.

Anticorpo (Clone)	Fabricante	Recuperação antigênica	Concentração	Sistema de detecção
Anti-Phospho-Chk2 (Thr68)	Cell Signaling Technology	<i>Trilogy</i> pH 7.5 90°C 20min	1:50	Envision/HRP
Anti-H2AX (Ser139)	Cell Signaling Technology	<i>Trilogy</i> pH 7.5 90°C 20min	1:200	Envision/HRP

Anti-P53 (DO7)	DAKO	Ác. cítrico pH 6.0 90°C 20min	1:50	Envision/HRP
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Fonte: elaborada pela autora, 2019.

2.5.1 Protocolo de IHQ para CHK2 e γ H2AX

- I. Desparafinização, hidratação e recuperação antigênica: reagente de “*trilogy*” (Rocklin, USA);
- II. Dois banhos “*trilogy*” em água destilada;
- III. Cubas em panela no vapor até a temperatura do banho atingir 90°C;
- IV. Lavagem em 5 banhos de água destilada;
- V. Etapa de bloqueio com metanol + H₂O₂ (10v);
- VI. Lavagem em 5 banhos de água destilada;
- VII. Lavagem em tampão TRIS- HCl 20 mM (3 banhos de 5 minutos);
- VIII. Incubação do anticorpo primário diluído na solução diluente (DAKO; Glostrup Municipality, Denmark), em câmara úmida, *overnight* – 4°C. Lavagem 3 vezes na solução de TRIS-HCl 20 mM;
- IX. Incubação do anticorpo secundário (*DAKO ENVISION + DUAL LINK-SYSTEM HRP*; Glostrup Municipality, Denmark);
- X. Três lavagens na solução de TRIS-HCl 20 mM (5 minutos cada banho);
- XI. Lavagem das lâminas em água destilada;
- XII. Lavagem das lâminas em água corrente por 10 minutos (fluxo forte);
- XIII. Incubação dos cortes com a hematoxilina de Mayer filtrada (3 minutos);
- XIV. Passar as lâminas rapidamente pela solução de hidróxido de amônio 10%;
- XV. Mergulhar as lâminas em água corrente (fluxo forte) por 5 minutos;
- XVI. Desidratação em cadeia ascendente de etanol (70%, 90% e três vezes a de 100%), diafanização em três banhos de xilol, sendo as lâminas montadas ao final do procedimento com lamínulas de vidro e Tissue-Mount™ (TissueClear® based medium. Alphen aan den Rijn, Holanda).

2.5.1 Protocolo de IHQ para TP53

- I. Bateria de hidratação em cadeia descendente de etanol e três banhos em xilol;
- II. Dois banhos em Ácido Cítrico pH 6.0;
- III. Cubas em panela no vapor até a temperatura do banho atingir 90°C;
- IV. Lavagem em 5 banhos de água destilada;
- V. Etapa de bloqueio com metanol + H₂O₂ (10v);

- VI. Lavagem em 5 banhos de água destilada;
- VII. Lavagem em tampão TRIS- HCl 20 mM (3 banhos de 5 minutos);
- VIII. Incubação do anticorpo primário diluído na solução diluente (DAKO; Glostrup Municipality, Denmark), em câmara úmida, *overnight* – 4°C. Lavagem 3 vezes na solução de TRIS-HCl 20 mM;
- IX. Incubação do anticorpo secundário (*DAKO ENVISION + DUAL LINK-SYSTEM HRP*; Glostrup Municipality, Denmark);
- X. Três lavagens na solução de TRIS-HCl 20 mM (5 minutos cada banho);
- XI. Lavagem das lâminas em água destilada;
- XII. Lavagem das lâminas em água corrente por 10 minutos (fluxo forte);
- XIII. Incubação dos cortes com a hematoxilina de Mayer filtrada (3 minutos);
- XIV. Passar as lâminas rapidamente pela solução de hidróxido de amônio 10%;
- XV. Mergulhar as lâminas em água corrente (fluxo forte) por 5 minutos;
- XVI. Desidratação em cadeia ascendente de etanol (70%, 90% e três vezes a de 100%), diafanização em três banhos de xilol, sendo as lâminas montadas ao final do procedimento com lamínulas de vidro e Tissue-Mount™ (TissueClear® based medium. Alphen aan den Rijn, Holanda).

2.6 Análise dos resultados

A análise dos dados foi realizada no software *Statistical Package for the Social Sciences* (SPSS), versão 23.0 (SPSS Inc., Chicago, IL, EUA). As variáveis desfecho foram avaliadas quanto à normalidade por meio dos testes de Shapiro-Wilk, nos grupos fumantes e não fumantes ($p < 0,05$). Testes paramétricos ou não-paramétricos, quando indicados, foram desenvolvidos para comparar os valores das proteínas CHK2, γ H2AX e TP53, entre grupos fumantes e não fumantes, considerando o nível de significância $p < 0,05$.

3 ARTIGO

Os resultados foram escritos em língua inglesa na forma de artigo científico.

3.1 Artigo Científico

Artigo submetido ao periódico internacional: Archives of Oral Biology (Qualis A1 - Fator de impacto: 1.663; em 2019). Encontra-se aguardando resposta dos revisores.

Immunoexpression of CHK2, H2AX and P53 is similar in oral squamous cell carcinoma samples from smoking and non-smoking patients

Running title: Immunoexpression of CHK2, H2AX and P53

Highlights

- Similarities in CHK2, H2AX and P53 staining were observed in smokers and non-smokers with OSCC.
- There is a consistent expression of CHK2, H2AX and P53 proteins in OSCC.
- Pathway of DNA damage as an early and advanced event in oral carcinogenesis.

Abbreviations

CHK2: checkpoint kinase 2

DDR: DNA damage response

DSBs: double-strand breaks

H&E: hematoxylin and eosin

H2AX: H2A histone family member X

OSCC: oral squamous cell carcinoma

UFMG: Universidade Federal de Minas Gerais

Abstract

Objective: To assess the DNA damage response through the expression of checkpoint kinase 2 (CHK2), H2A histone family member X (H2AX) and P53 among smokers and non-smokers with oral squamous cell carcinoma (OSCC). In addition, associations between the immunoexpression of the studied proteins and clinicopathologic data and histopathological grading were analyzed.

Design: Thirty-five individuals (18 non-smokers and 17 smokers) with OSCC of the tongue and/or floor of the mouth were included. Immunohistochemistry for H2AX was carried out for the identification of double-strand breaks, CHK2 and P53 in order to evaluate the induction of the cell cycle. Descriptive and statistical analyses were performed.

Results: The sample consisted of 22 males (62.8%) and 13 females (37.2%), with a mean age of 63.9 ± 11.8 years. The OSCC of non-smokers were well-differentiated tumors in 50% of cases, and those of smokers were equally distributed into moderately differentiated and poorly differentiated tumors (35.3% each). Overall, 31 (88.6%) cases were CHK2-positive, 27 (77.1%) were H2AX-positive and 23 (65.7%) were P53-positive, with no difference between smokers and non-smokers ($p > 0.05$). No association was found between proteins and clinicopathologic data ($p > 0.05$).

Conclusions: Similarities in the CHK2, H2AX and P53 immunohistochemical staining pattern were observed between smokers and non-smokers with OSCC in this survey, and the immunoexpression was not associated with clinicopathologic parameters. Overall, the results indicated consistent expression of these proteins in OSCC. This study provides information about the pathway of DNA damage as an early and advanced event in oral carcinogenesis.

Keywords: DNA Damage, Checkpoint Kinase 2, H2AX protein, Gamma-H2AX protein, Tumor Suppressor Protein p53, Oral cancer.

1. Introduction

Oral squamous cell carcinoma (OSCC) is a global problem with an annual incidence of 300,000 cases (D’Cruz, Vaish, & Dhar, 2018). Genetic, epigenetic and environmental factors are involved in this multifactorial disease (Chi, Day, & Neville, 2015; Ali et al., 2017). Cigarette smoking is the main etiologic factor of oral cancer and its mechanism of action involves direct DNA damage (Jethwa & Khariwala, 2017). During carcinogenesis, some pathways may be altered, modifying tumor proliferation and apoptosis, and inducing cell transformation and clonal expansion of tumor cells (Monteiro et al., 2018). DNA damage is a recurrent phenomenon in metabolism that can be induced by exogenous and endogenous agents. When these factors accumulate, they can cause genomic instability, which eventually results in the carcinogenesis process (Nikitakis et al., 2018).

Cigarette smoke contains reactive oxidants which may cause macromolecular damage to exposed cells. Thus, the production of reactive oxygen species may damage DNA, occurring in the form of mutations, deletions, changes in sugar bases, cytosine halogenation or oxidation, and methylation (Murugan, Munirajan, & Tsuchida, 2012; De Oliveira, Da Silva, Mariz, Pereira, & De Oliveira, 2015). This fact suggests that the levels of specific proteins related to DNA damage such as checkpoint kinase 2 (CHK2), H2A histone family member X (H2AX) and P53 in smokers might serve as a measure of their cancer risk (Valinluck & Sowers, 2007).

The DNA damage response (DDR) represents the cell’s ability to restore genomic changes caused by endogenous or exogenous mutagens (Nikitakis et al., 2018). H2AX phosphorylation is a key step in the DDR, playing a role in signaling and initiating the repair of double-strand breaks (DSBs) (Sedelnikova & Bonner, 2006; Bonner et al., 2008; Palla et al., 2017). DSBs must be repaired quickly and precisely to avoid cell death, chromosomal aberrations, mutations and, in certain cases, initiation of pathological effects

such as cancer. The rapid phosphorylation of H2AX represents an early cellular response to DSBs (Mah, El-Osta & Karagiannis, 2010).

Moreover, in response to DNA damage, a complex signaling network organizes cell-cycle checkpoints allowing cell cycle arrest and DNA repair, or activates senescence or cell death (Roos & Kaina, 2013). CHK2 is central to transducing the DNA damage signal and has been implicated in the mediation of both G1/S and G2/M cell-cycle arrest in a distinct pathway through P53 (Antoni, Sondha, Collins & Garrett, 2007; Zannini, Delia, & Buscemi, 2014). In addition, P53 is essential in regulating cell cycle progression, differentiation, DNA repair, and apoptosis. Almost 50% of individuals with oral cancer exhibit *P53* gene mutation and the high frequency of P53 mutations in human cancers highlights its role as a tumor suppressor (Sinevici & O'sullivan, 2016).

Nevertheless, the effect of smoking on the expression of CHK2 and H2AX in OSCC has not been well explored (De Oliveira et al., 2015; Nikitakis et al., 2018). In this study, we evaluated the immunoexpression of CHK2, H2AX and P53 proteins associated with the DNA protection and repair process among smokers and non-smokers with OSCC. Additionally, we analyzed the associations of proteins with clinicopathologic data and histopathological grading.

2. Materials and Methods

2.1 Study design and ethical approval

This retrospective and cross-sectional study evaluated 35 paraffin-embedded tissue specimens of OSCC. The specimens were obtained from the archives of the Oral and Maxillofacial Pathology Laboratory of Universidade Federal de Minas Gerais (UFMG), Belo Horizonte, Brazil. The study was approved by the Institutional Ethics Committee (No.

03012618.1.0000.5149) and the patient's identity remained anonymous according to the Declaration of Helsinki.

2.2 Patients and samples

Individuals with OSCC were analyzed according to sex, age, anatomical location (tongue and/or floor of the mouth), alcohol consumption (yes or no), and smoking habit. Never smokers with a ≤ 0.5 pack-years smoking history, as well as smokers with a >10 pack-years smoking history were considered (Paulin et al., 2015).

The histopathological diagnosis was confirmed by two independent oral and maxillofacial pathologists (V.F.B. and M.C.F.A.), who were blinded to the clinicodemographic data, by reviewing the sections stained with hematoxylin and eosin (H&E) retrieved from the files. Disagreements were jointly reviewed to reach a consensus.

All samples were graded as well differentiated, moderately differentiated or poorly differentiated according to the World Health Organization (El-Naggar, Chan, Grandis, Takata, & Slootweg, 2017). The exclusion criteria ruled out OSCC individuals submitted to radiotherapy, chemotherapy or other treatments before the surgical approach.

2.3 Immunohistochemical staining

For the immunohistochemical study, 4- μm thick sections were obtained from paraffin-embedded tissue blocks and mounted on polarized slides (StarFrost[®], Waldemar Knittel Glasbearbeitungs GmbH, Braunschweig, Germany). Monoclonal antibodies, including anti-phospho-CHEK2 (clone Thr68; Rabbit; Cell Signaling Technology, Danvers, MA, US; 1:50), anti-H2AX (clone Ser19; Rabbit; Cell Signaling Technology, Danvers, MA, US; 1:200), and anti-P53 (clone DO7, Mouse; Dako, Carpinteria, CA, US; 1:50) were used. The antigen-retrieval step was performed using the TRILOGY[™] Concentrate (Cell

Marque, Rocklin, CA, US; 1:100) at a temperature of 96°C in a digital water bath (DeLeo, Porto Alegre, RS, Brazil) for 30 minutes. Next, the sections were treated with the EnVision+ Dual Link System-HRP (Dako, Carpinteria, CA, US). 3,3'-Diaminobenzidine was used as the chromogen (Dako, Carpinteria, CA, US).

2.4 Immunohistochemical assessment

All cases were evaluated by one observer (L.F.S.) using an eyepiece grid coupled to a light microscope (Zeiss Axiostar, Ser. 48824, Oberkochen, Germany). Immunostaining was evaluated using a semiquantitative analysis of representative regions of each specimen. The slides were analyzed with a light microscope at a final magnification of 400×. Immunostaining was scored by counting the percentage of cells expressing the above-mentioned proteins in at least 10 to 15 different fields. Semi-quantitative analysis was performed based on the proportion of positive neoplastic cells relative to all neoplastic cells throughout the tissue section (Soares et al., 2017). For each brown stain, CHK2 expression was defined as nuclear, and cytoplasmic immunoreactivity (Lee et al., 2014), H2AX (Nikitakis et al., 2018) and P53 (Nagao et al., 2017) were defined as nuclear immunoreactivity. The immunoreactivity expression was considered negative (0%), normal (<50%) and overexpression ($\geq 50\%$) (Lassus, Leminen, Lundin, Lehtovirta & Butzow, 2003; Karpathiou et al., 2006).

2.6 Data analysis

Statistical tests were carried out using by the Statistical Package for the Social Sciences (SPSS) software (IBM Corp., version 23.0, Armonk, USA). Outcome variables were assessed for normality using the Shapiro-Wilk test for the smoker and non-smoker groups ($p < 0.05$). The Mann-Whitney test was used to compare the values of CHK2, H2AX,

and P53 proteins according to the clinical features (smoking habits, alcohol consumption, and anatomical location). The level of significance was set at $p < 0.05$.

3. Results

3.1 Demographic and clinicopathological data

The demographic and clinicopathological characteristics of the sample are depicted in Table 1. Thirty-five OSCC were included. The sample consisted of 22 males (62.8%) and 13 females (37.2%), with a male-to-female ratio of 1.7:1. The mean age of the sample as a whole was 63.9 years (range: 47 to 86 \pm 11.8 years). Of the study subjects, 18 (51.4%) were non-smokers and 17 (48.6%) were smokers. Among the smokers, males (n=14, 82.3%) in the sixth decade of life (n=10, 58.9%) were most affected. Regarding non-smokers, females (n=10, 55.6%) in the eighth decade of life (n=6, 33.3%) were most affected. Alcohol consumption was reported by 70.6% (n=12) of smokers and 29.4% (n=5) of non-smokers.

As regards histological grading, 40% (n=14) were well-differentiated tumors, 31.4% (n=11) were moderately differentiated, and 28.6% (n=10) poorly differentiated. Fifty percent of non-smokers' OSCC were well-differentiated tumors, whereas for smokers, OSCC were moderately differentiated and poorly differentiated tumors in equal proportions (n=6, 35.3% each).

3.2 Immunoexpression of CHK2, H2AX and P53 positive cells

Overall, most cases exhibited positivity (normal or overexpression) for CHK2, H2AX and P53. Thirty-one (88.6%) cases were CHK2-positive, 27 (77.2%) were H2AX-positive, and 23 (65.7%) were P53-positive. Figure 1 shows the distribution of immunostaining among non-smokers and smokers.

In smokers' OSCC samples, CHK2 staining ranged from 0 to 83.3% of positive cells (Figure 2A). One case was negative, six cases demonstrated normal positivity, and 10 showed overexpression. The expression in non-smokers' OSCC was closely similar (Figure 2B), with an expression index ranging from 0 to 91.2%. Three cases were negative, seven were normal, and eight showed overexpression.

The expression of H2AX was also quite similar in smokers (Figure 2C) and non-smokers (Figure 2D). However, disparities in mean number of positive cells were observed between smokers (0 to 92.9%) and non-smokers (0 to 53.5%). Two OSCC of smokers showed H2AX overexpression, 11 showed normal expression and four were negative. Four OSCC of non-smokers were negative, 11 were considered to have normal expression, and one showed overexpression.

The expression of P53 positive cells ranged from 0 to 91.9% for smokers and from 0 to 92.8% for non-smokers (Figure 2E and 2F). Six cases of each group were negative, and two cases of each group showed normal staining. Nine cases of smokers and 10 cases of non-smokers were considered to show overexpression.

No association was found between protein immunoexpression and the clinicopathological features of smokers and non-smokers ($p > 0.05$). Also, no statistically significant difference was observed between groups (smokers vs. non-smokers) regarding CHK2 ($p = 0.909$), H2AX ($p = 0.807$) and P53 ($p = 0.546$) proteins (Figure 3). No correlation was found among the proteins.

4. Discussion

Tobacco smoking involves a high risk for human malignancies, including oral cancer, because it contains multiple carcinogens that cause genetic instability (Beynon et al., 2018). Therefore, a worse prognosis would be expected for cancer patients who are

smokers. The present study investigated the expression of molecules related to the DNA damage response in smokers and non-smokers with OSCC. The selected markers participate in different stages of the DNA damage response, from identification of DSBs (H2AX) to induction of cell cycle arrest (CHK2 and P53). Herein, the immunoexpression of CHK2, H2AX and P53 was similar in smoking and non-smoking OSCC samples. Likewise, a previous study showed similar carcinogenic pathways and outcomes in oral premalignant lesions of smokers and non-smokers (de la Oliva et al., 2019).

The carcinogenic environment is represented by continuous genomic instability involving the activation of oncogenes and inactivation of tumor suppressor genes (Nikitakis et al., 2018; de la Oliva et al., 2019). As a consequence, the DNA damage response system suffers a breakdown. Moreover, epigenetic regulation, dietary factors, oral homeostasis and environmental pressures may be implicated in OSCC of both non-smokers and smokers (De Oliveira et al., 2015; Sarode, Sharma, Sarode, & Patil, 2019). Nevertheless, this similarity may explain the absence of difference between groups.

CHK2 is a key regulator kinase involved in the DNA-damage response-signaling pathway. Induced by DNA DSBs, ataxia telangiectasia mutated participates in the activation of CHK2, which is subsequently phosphorylated to block the cell cycle and to activate the transcription of repair genes (Squatrito et al., 2010; Matthews, Jones, & Collins, 2013; Tu et al., 2013). Studies have reported that CHK2 is linked to tumor progression in high-grade serous carcinoma and associated with an advanced TNM stage and poor prognosis in gastric malignant tumors (Lee et al., 2014; Davidson et al., 2018). In this study, although CHK2 expression was slightly lower in non-smokers than in smokers, protein immunoexpression did not differ between the two groups and no other association with clinicopathological features was demonstrated. The influence of tobacco on DNA repair and CHK2 action has been demonstrated in other studies (Tanaka et al., 2007; Zhao,

Albino, Jorgensen, Traganos & Darzynkiewicz, 2009; Li et al., 2013). An *in vitro* investigation showed an attenuation of CHK2 phosphorylation in lung epithelial cells treated with nicotine (Nishioka et al., 2011). In the present survey, the similarity between groups could be overridden by a large sample.

The tumor suppressor P53 has been reported to be another key target of CHK2 in response to DNA damage (Stolz, Ertych, & Bastians, 2011). P53 is reported to be mutated by over 80% in all cancers (Harjes, 2019). Loss of this protein prevents cell cycle arrest and apoptosis in the DNA damage scenario (Senturk & Manfredi, 2013). P53 changes are mainly caused by exogenous factors, particularly tobacco carcinogens in cases of OSCC (Hsieh et al., 2011). In the *P53* gene, the frequency of mutations is higher in lung tumors from smokers than in lung tumors from non-smokers. Additionally, the occurrence of *P53* mutations in lung cancer samples from smokers has been observed to be dependent on lifetime cigarette consumption or duration of smoking (Takehima et al., 1993). Regarding prostate tissue, smoking-related carcinogens can alter the expressions of some suppressor genes such as P53, suggesting that changes in these genes in prostate gland epithelia may possibly increase the risk for prostate carcinoma (Boran et al., 2017).

Herein, we observed an overexpression of P53 in 19/35 (54.3%) of cases, with no difference between smokers and non-smokers. Other studies have also found a similar expression of P53 in OSCC of smokers and non-smokers (Zaid, Azar-Maalouf, Barakat, & Chantiri 2018; de la Oliva et al., 2019). In one study, all specimens were P53 positive, independent of tobacco (Nikitakis et al., 2018). The authors stated that the main molecular alterations in oral premalignant lesions and those associated with the progression to OSCC are shared by non-smokers and smokers and appear in the early stages of carcinogenesis.

In recent years, the phosphorylated histone H2AX (γ -H2AX) marker has become a robust tool used to monitor DNA DSBs in cancer research, and has been suggested to play

a potential role in carcinogenesis and early cancer diagnosis (Sedelnikova & Bonner, 2006; Bonner et al., 2008; Palla et al., 2017). Accordingly, many studies have revealed a correlation between high γ -H2AX levels and worse prognosis and reduced disease-free survival in breast cancer (Wang, Zhang, Xia, Jiang, & Wang, 2019), ovarian cancer (Mei et al., 2015), colon carcinoma (Sedelnikova & Bonner, 2006), hepatocellular carcinoma (Xiao et al., 2015), and melanoma (Warters, Adamson, Pond, & Leachman, 2005). Interestingly, an association of H2AX positivity in OSCC with reduced overall survival time has been reported (Oliveira-Costa et al., 2014).

In the present study, no association was observed between clinicopathologic features of smokers and non-smokers and H2AX protein. However, a study has reported that the DNA DSB γ -H2AX marker exists in smoke-exposed placentae and the cessation of smoking reduces DSB DNA damage to the levels associated with non-smokers (Slatter et al., 2014). In addition, available literature data have demonstrated that exposure of A549 (human lung adenocarcinoma) cells to tobacco smoke or of NHBE (normal human bronchial epithelial) cells to smoke condensate induced γ -H2AX (Albino et al., 2004; Albino et al., 2009).

In our study, the majority of non-smokers (70.6%) did not have associated alcohol consumption and were women in the eighth and ninth decades of life. In fact, other OSCC series that included non-smokers also reported similar data (Koo, Barrowman, McCullough, Iseli, & Wiesenfeld, 2013; DeAngelis et al., 2018). Moreover, some studies that discussed changes in the genetic profile of OSCC when comparing smoker and non-smokers have been reported elsewhere with divergent results and the etiopathogenesis of OSCC in the non-smoking population remains unknown (DeAngelis et al., 2018).

While in some studies smoking history did not play a differential role in carcinogenesis (Kato et al., 1998; Kietthubthew et al., 2001; Chaves et al., 2004), the vast

majority have reported that smokers are more subjected to genetic instability (Park et al., 2000; Prior et al., 2006; Sharma et al., 2006; Bau et al., 2017; Chen et al., 2007; Tsai et al., 2009). Although tobacco can cause epigenetic changes in oral epithelial cells, inhibit multiple systemic immune functions of the host, and induce oxidative stress in tissues through its toxic metabolites, leading to OSCC (Jiang, Wu, Wang & Huang, 2019), there is no specific mutation signature associated with smoking for OSCC (Chaves et al., 2004; Pickering et al., 2014; Jethwa & Khariwala, 2017; Jiang, Wu, Wang, & Huang, 2019; Tomar, 2019). The mutation signature from smoking appears to be site-specific and has been defined for lung tumors. Laryngeal squamous cell carcinoma also exhibits the strongest smoking signature among the head and neck squamous cell carcinoma sites, but there is no defined signature related to smoking for tumors at other sites (Brennan, Koenig, Gentles, Sunwoo, & Gevaerta, 2017).

Besides the limited sample, some other restrictions should be considered. The evaluation of tobacco smoking was based on self-reports documented in medical records. Thus, it was not possible to assess the amount of passive smoking of environmental tobacco and other carcinogens to which the patients may have been exposed. Furthermore, we did not consider patient diet, hormonal status, HPV infection or second-hand smoking. Thus, the effect of these factors on the immunoexpression of the studied proteins remains to be investigated.

5. Conclusions

The present study showed similarities in the CHK2, H2AX and P53 immunohistochemical staining pattern between smokers and non-smokers with OSCC. Overall, the results indicated consistent expression of these proteins in OSCC with marked nuclear and/or cytoplasmic labeling. It would be worth investigating if these characteristics

persist in non-smokers with poorly differentiated OSCC. This study provides insights on the pathway of DNA damage as an early and advanced event in oral carcinogenesis.

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Figure legends

Figure 1. Overall distribution of immunostaining in smokers and non-smokers.

Figure 2. CHK2 staining in a smoker **(A)** and non-smoker **(B)**. H2AX staining in a smoker **(C)** and non-smoker **(D)**. P53 staining in a smoker **(E)** and non-smoker **(F)**. Immunohistochemistry, 20 \times .

Figure 3. Immunoexpression of the proteins CHK2 ($p=0.909$), H2AX ($p=0.807$) and P53 ($p=0.546$) between smokers and non-smokers.

Table 1. Demographic data and clinicopathological characteristics of the sample

Variable	n (%)	
	Non-smokers, n=18 (51.4)	Smokers, n=17 (48.6)
Sex, n=35		
Male	8 (44.4)	14 (82.3)
Female	10 (55.6)	3 (17.7)
Age, n=35		
40–49	2 (11.1)	1 (5.9)
50–59	4 (22.2)	10 (58.9)
60–69	1 (5.6)	6 (35.2)
70–79	6 (33.3)	-
80–89	5 (27.8)	-
Alcohol consumption, n=34		
Yes	5 (29.4)	12 (70.6)
No	12 (70.6)	5 (29.4)
Symptoms, n=18		
Yes	7 (58.3)	4 (66.7)
No	5 (41.7)	2 (33.3)
Anatomical location, n=35		
Tongue	14 (77.8)	7 (41.2)
Floor of the mouth	3 (16.7)	8 (47.1)
Tongue + floor of the mouth	1 (5.5)	2 (11.7)
Type of sample, n=35		
Incisional	12 (66.7)	8 (47.1)
Excisional	6 (33.3)	9 (52.9)
Histology/grade, n=35		
Well-differentiated	9 (50.0)	5 (29.4)
Moderately differentiated	4 (22.2)	6 (35.3)
Poorly differentiated	5 (27.8)	6 (35.3)

Figure 1. Overall distribution of immunostaining in smokers and non-smokers.

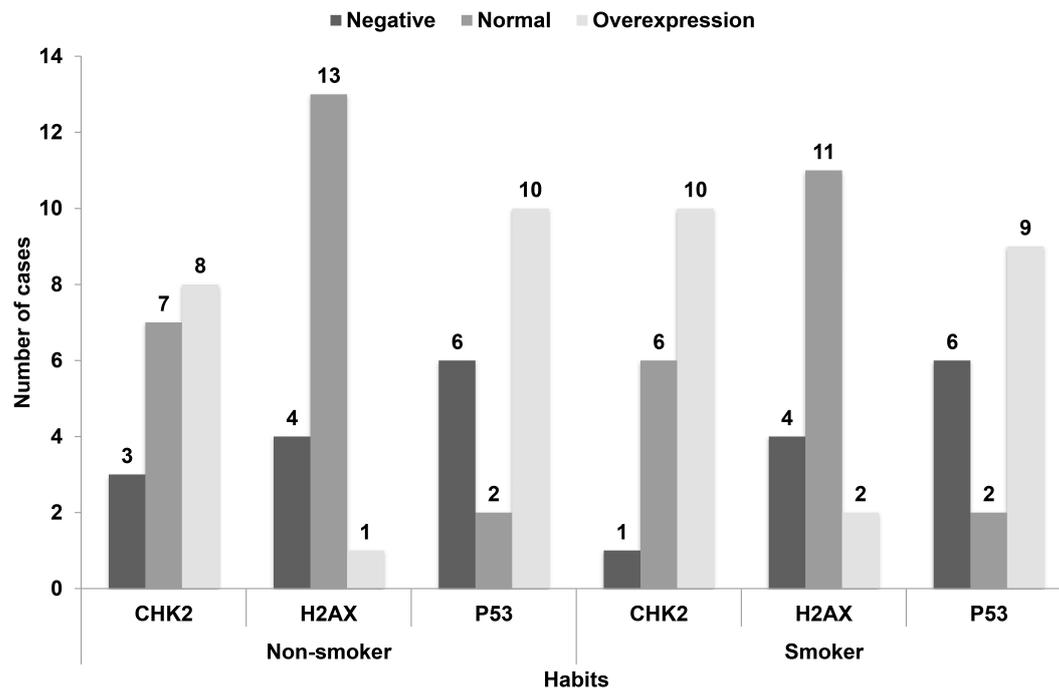


Figure 2. CHK2 staining in a smoker (A) and non-smoker (B). H2AX staining in a smoker (C) and non-smoker (D). P53 staining in a smoker (E) and non-smoker (F). Immunohistochemistry, 20 \times .

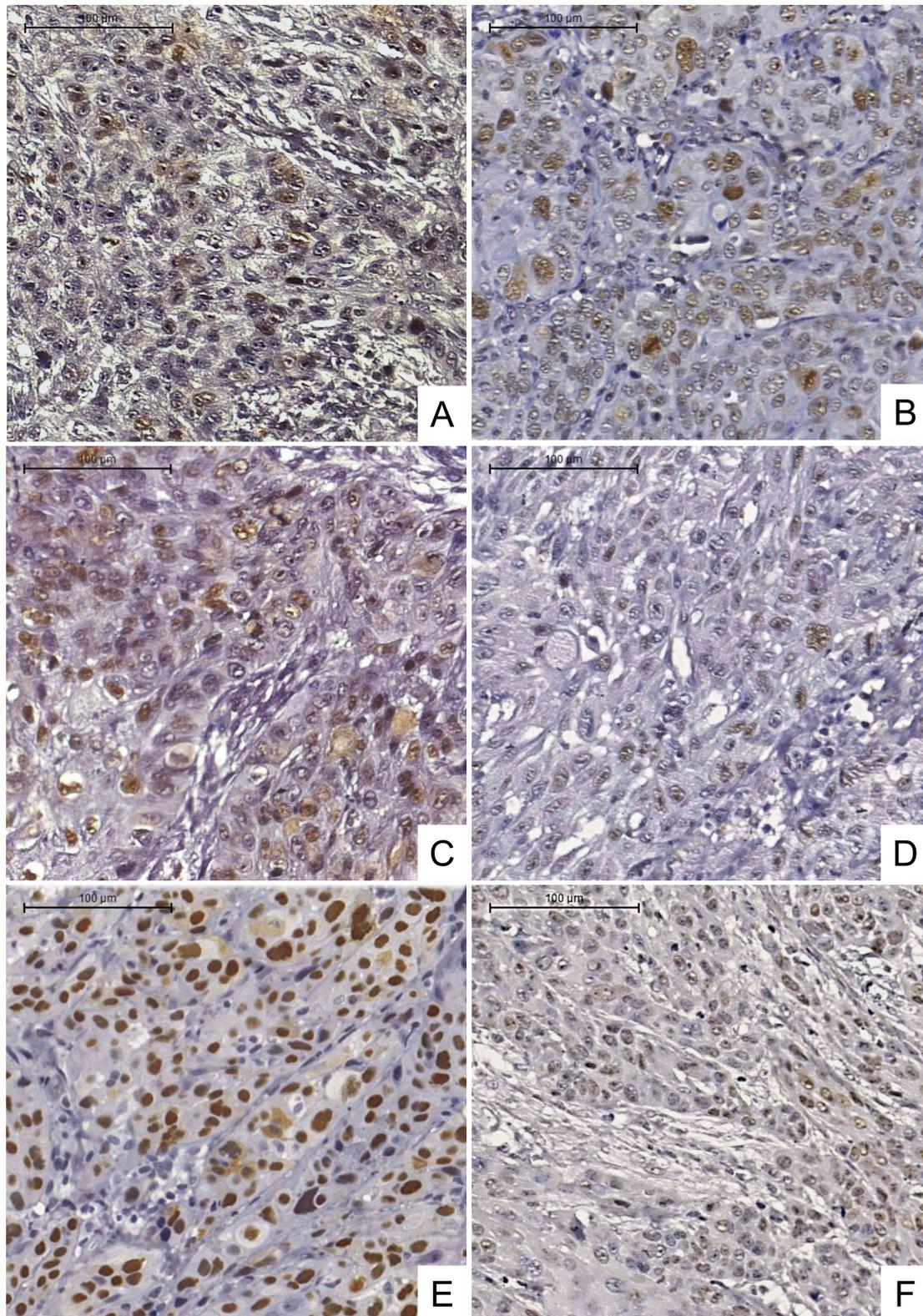
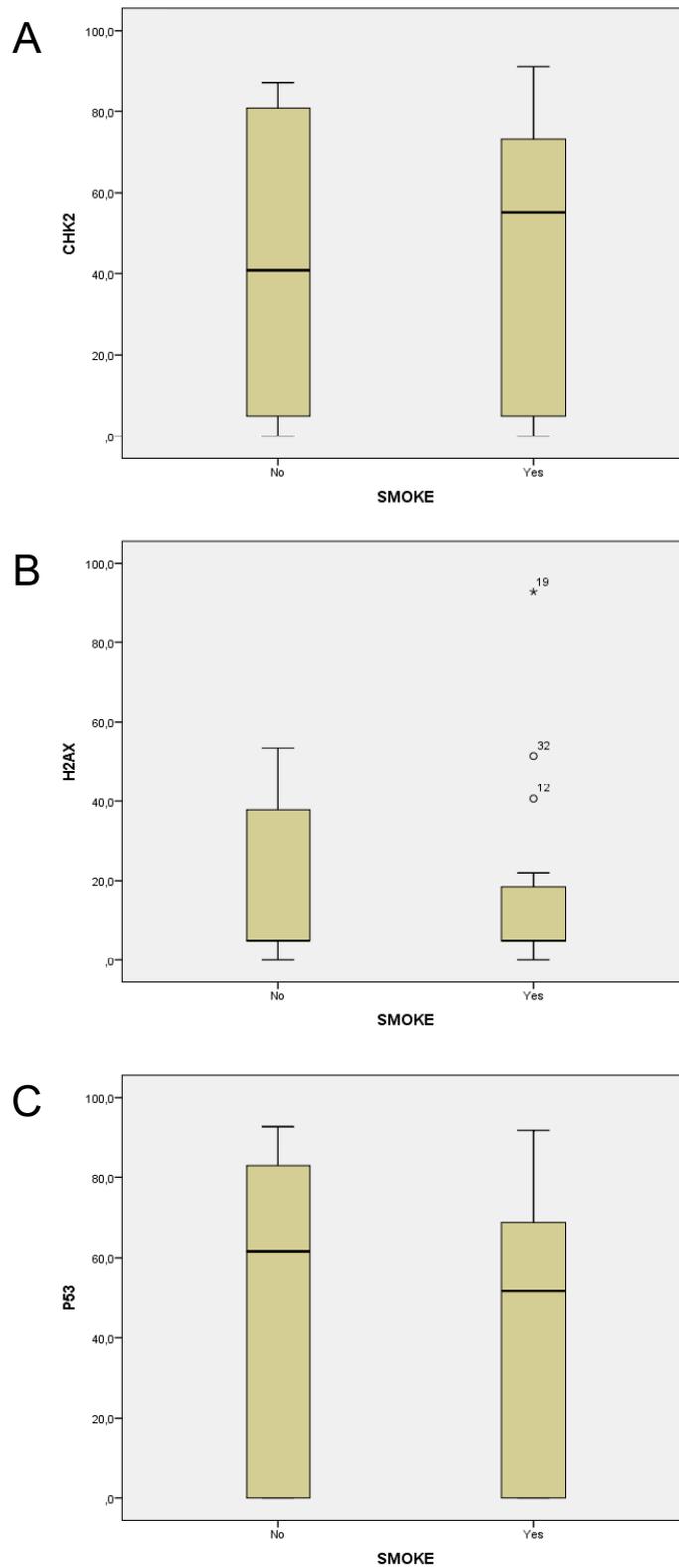


Figure 3. Immunoeexpression of the proteins CHK2 ($p=0.909$), H2AX ($p=0.807$) and P53 ($p=0.546$) between smokers and non-smokers.



4 CONSIDERAÇÕES FINAIS

Este estudo fornece informações sobre algumas proteínas envolvidas no dano ao DNA, importantes no processo de carcinogênese oral. Observou-se semelhanças entre a expressão imuno-histoquímica de CHK2, γ H2AX e TP53 em fumantes e não fumantes com CCEB. Os mecanismos de proteção ao DNA e as alterações relacionadas ao ciclo e checagem permaneceram as mesmas em pacientes fumantes e não fumantes. Porém, nos não fumantes, outras alterações genéticas que não foram abordadas neste trabalho influenciam no desenvolvimento do tumor, podendo levar a lesões mais agressivas. Esses achados não diminuem a importância dos programas de cessação do tabagismo e da ingestão de álcool para reduzir a incidência de câncer oral. No entanto, eles sugerem possíveis novos caminhos para a prevenção ou tratamento desta doença.

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ANEXO A – Aprovação do comitê de ética em pesquisa

UNIVERSIDADE FEDERAL DE
MINAS GERAIS



PARECER CONSUBSTANCIADO DO CEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: PADRÃO DE CONSUMO DE TABACO EM PACIENTES COM CÂNCER DA CAVIDADE ORAL: ESTUDO DAS ASSOCIAÇÕES CLINICOPATOLÓGICAS E IMUNOISTOQUÍMICAS

Pesquisador: Maria Cássia Ferreira de Aguiar

Área Temática:

Versão: 2

CAAE: 03012618.1.0000.5149

Instituição Proponente: PRO REITORIA DE PESQUISA

Patrocinador Principal: Financiamento Próprio

DADOS DO PARECER

Número do Parecer: 3.293.055

Apresentação do Projeto:

A incidência dos casos de câncer da cavidade oral, lábios e orofaringe, representados principalmente pela variante carcinoma de células escamosas (CCE), apesar das variações entre os países, na maioria deles ainda apresenta trajetória ascendente e preocupante. O presente estudo é do tipo prospectivo caso-controle dos casos de câncer da cavidade oral atendidos na Clínica de Patologia, Cirurgia e Radiologia da Faculdade de Odontologia da Universidade Federal de Minas Gerais, que será desenvolvido conjuntamente com um estudo transversal, do tipo retrospectivo, com população-alvo constituída por indivíduos com diagnóstico histopatológico de carcinoma de células escamosas de boca que tiveram laudo gerado pelo Laboratório de Patologia Bucomaxilofacial da Faculdade de Odontologia da Universidade Federal de Minas Gerais.

Objetivo da Pesquisa:

O objetivo principal é avaliar prospectivamente o padrão de consumo de tabaco, incluindo o índice de exposição cumulativa ao alcatrão (IEC), em pacientes com CCEB, e verificar sua relação com características clínicas e histopatológicas da doença, bem como investigar a expressão de algumas proteínas do ciclo celular em amostras de CCEB já coletadas, de pacientes fumantes e não fumantes.

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Telefone: (31)3409-4592

E-mail: coep@prpq.ufmg.br

Continuação do Parecer: 3.293.055

Avaliação dos Riscos e Benefícios:

De acordo com o pesquisador, a pesquisa em suas duas vertentes oferece riscos mínimos aos indivíduos, uma vez que será realizada por meio da aplicação de questionários, após consentimento dos pacientes; e por meio da avaliação imunistoquímica de material biológico previamente coletado. Em relação à aplicação dos questionários, os participantes poderão se sentir cansados ou constrangidos com alguma pergunta. Os pesquisadores irão minimizar este risco cuidando para que a entrevista seja feita de forma reservada, breve e deixando claro que o sigilo quanto à identidade dos participantes será mantido. Os mesmos cuidados quanto à preservação da identidade serão tomados em relação ao uso do material de arquivo. Ressalta-se ainda, que os pacientes não terão seu tratamento e acompanhamento prejudicados.

Como benefícios, é descrito conhecer melhor o padrão de consumo de tabaco entre indivíduos com e sem CCEB, bem como avaliar a relação do uso de tabaco com características clínicas, patológicas e imunistoquímicas dessa neoplasia. Sendo assim, espera-se gerar resultados que subsidiem políticas públicas de combate ao tabagismo e alertem a população sobre a importância do consumo de tabaco no desenvolvimento do câncer de boca.

Comentários e Considerações sobre a Pesquisa:

Constam no processo: Projeto original; Formulário de Informações Básicas; Carta Resposta ao Comitê de Ética; TCLE modificado de acordo com as solicitações do Comitê de Ética; Termo de Compromisso de Utilização de Dados; Declaração de Compromisso do Pesquisador; Parecer Departamental e Orçamento.

Considerações sobre os Termos de apresentação obrigatória:

A pesquisadora atendeu todas as solicitações do Comitê de Ética, enviando carta resposta, TCLE com alterações e TCUD.

Recomendações:

Não há recomendações.

Conclusões ou Pendências e Lista de Inadequações:

S.M.J. julgo aprovado o projeto de pesquisa.

Considerações Finais a critério do CEP:

Tendo em vista a legislação vigente (Resolução CNS 466/12), o CEP-UFMG recomenda aos Pesquisadores: comunicar toda e qualquer alteração do projeto e do termo de consentimento via emenda na Plataforma Brasil, informar imediatamente qualquer evento adverso ocorrido durante o

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Continuação do Parecer: 3.293.055

desenvolvimento da pesquisa (via documental encaminhada em papel), apresentar na forma de notificação relatórios parciais do andamento do mesmo a cada 06 (seis) meses e ao término da pesquisa encaminhar a este Comitê um sumário dos resultados do projeto (relatório final).

Este parecer foi elaborado baseado nos documentos abaixo relacionados:

Tipo Documento	Arquivo	Postagem	Autor	Situação
Informações Básicas do Projeto	PB_INFORMAÇÕES_BÁSICAS_DO_PROJETO_1229178.pdf	22/03/2019 13:02:31		Aceito
Outros	TCUD.pdf	22/03/2019 13:00:56	Karine Duarte da Silva	Aceito
Projeto Detalhado / Brochura Investigador	Projeto_corrigido.pdf	22/03/2019 12:59:11	Karine Duarte da Silva	Aceito
Outros	Carta_resposta_COEP.pdf	22/03/2019 12:58:56	Karine Duarte da Silva	Aceito
Declaração de Manuseio Material Biológico / Biorepositório / Biobanco	Autorizacao_novo2.pdf	22/03/2019 12:58:21	Karine Duarte da Silva	Aceito
TCLE / Termos de Assentimento / Justificativa de Ausência	TCLE_novo2.pdf	22/03/2019 11:27:42	Karine Duarte da Silva	Aceito
TCLE / Termos de Assentimento / Justificativa de Ausência	dispensa.pdf	08/10/2018 08:30:44	Karine Duarte da Silva	Aceito
Outros	Parecer_departamento.pdf	08/10/2018 08:29:56	Karine Duarte da Silva	Aceito
Orçamento	orcamento.pdf	08/10/2018 08:29:00	Karine Duarte da Silva	Aceito
Declaração de Pesquisadores	Declaracao.pdf	08/10/2018 08:27:51	Karine Duarte da Silva	Aceito
Cronograma	Cronograma.pdf	08/10/2018 08:27:32	Karine Duarte da Silva	Aceito
Folha de Rosto	Folha_de_rosto.pdf	08/10/2018 08:26:59	Karine Duarte da Silva	Aceito

Situação do Parecer:

Aprovado

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Continuação do Parecer: 3.293.055

Necessita Apreciação da CONEP:

Não

BELO HORIZONTE, 29 de Abril de 2019

Assinado por:
Eliane Cristina de Freitas Rocha
(Coordenador(a))

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ANEXO B – Informações da revista

Revista: Archives of Oral Biology – A1

Editors-in-Chief:

Professor G B Proctor, London, UK

Professor S W Cadden, Dundee, Scotland

Archives of Oral Biology is an international journal which aims to publish papers of the highest scientific quality reporting new knowledge from the orofacial region including:

- developmental biology
- cell and molecular biology
- molecular genetics
- immunology
- pathogenesis
- microbiology
- biology of dental caries and periodontal disease
- forensic dentistry
- neuroscience
- salivary biology
- mastication and swallowing
- comparative anatomy
- paeleodontology

Archives of Oral Biology will also publish expert reviews and articles concerned with advancement in relevant methodologies. The journal will consider clinical papers only where they make a significant contribution to the understanding of a disease process.

These guidelines generally follow the Uniform Requirements for Manuscripts Submitted to Biomedical Journals

Types of Contribution

Original papers and review articles are welcomed. There will be no differentiation on the basis of length into full or short communications. Editorial commentaries will also be considered but only by invitation. All submissions will be refereed.

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You should use this list to carry out a final check of your submission before you send it to the journal for review. Please check all relevant sections in this Guide for Authors for more details.

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One author has been designated as the corresponding author with contact details:

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All necessary files have been uploaded:

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- Ensure all figure and table citations in the text match the files provided
- Indicate clearly if color should be used for any figures in print *Graphical Abstracts* (where applicable)

Highlights (where applicable)

Supplemental files (where applicable)

Further considerations

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- All references mentioned in the Reference List are cited in the text, and vice versa
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- Declarations of authors' contributions have been made if there are four or more authors
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be indicated, and where appropriate, the influence (or association) of sex on the results of the study.

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If there are four or more authors, then each is required to declare his or her individual contribution to the article: all authors must have materially participated in the research and article preparation, so roles for all authors should be described. The statement that "All authors have read and approved the final article" should be true and included in the disclosure.

Authorship

All authors should have made substantial contributions to all of the following: (1) the conception and design of the study, or acquisition of data, or analysis and interpretation of data, (2) drafting the article or revising it critically for important intellectual content, (3) final approval of the version to be submitted.

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Article structure

Manuscript Structure

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Introduction

This should be a succinct statement of the problem investigated within the context of a brief review of the relevant literature. Literature directly relevant to any inferences or argument presented in the Discussion should in general be reserved for that section. The introduction may conclude with the reason for doing the work but should not state what was done nor the findings.

Materials and Methods

Enough detail must be given here so that another worker can repeat the procedures exactly. Where the materials and methods were exactly as in a previous paper, it is not necessary to repeat all the details but sufficient information must be given for the reader to comprehend what was done without having to consult the earlier work.

Authors are requested to make plain that the conditions of animal and human experimentation are as outlined in the "Ethics" and "Studies on Animals" sections above

Results or Findings

These should be given clearly and concisely. Care should be taken to avoid drawing inferences that belong to the Discussion. Data may be presented in various forms such as histograms or tables but, in view of pressure on space, presentation of the same data in more than one form is unacceptable.

Discussion

This should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is occasionally appropriate. Avoid extensive citations and discussion of published literature. ***Conclusions***

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion section.

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- ***Title.*** Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.
- ***Author names and affiliations.*** Please clearly indicate the given name(s) and family name(s) of each author and check that all names are accurately spelled. You can add your name between parentheses in your own script behind the English transliteration. Present the authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-

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As titles frequently stand alone in indexes, bibliographic journals etc., and indexing of papers is, to an increasing extent, becoming computerized from key words in the titles, it is important that titles should be as concise and informative as possible. Thus the animal species to which the observations refer should always be given and it is desirable to indicate the type of method on which the observations are based, e.g. chemical, bacteriological, electron-microscopic, histochemical, etc. A "running title" of not more than 40 letters and spaces must also be supplied. A keyword index must be supplied for each paper.

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Highlights are mandatory for this journal as they help increase the discoverability of your article via search engines. They consist of a short collection of bullet points that capture the novel results of your research as well as new methods that were used during the study (if any).

Highlights should be submitted in a separate editable file in the online submission system. Please use 'Highlights' in the file name and include 3 to 5 bullet points (maximum 85 characters, including spaces, per bullet point).

Structured abstract

The paper should be prefaced by an abstract aimed at giving the entire paper in miniature. Abstracts should be no longer than 250 words and should be structured as per the guidelines published in the Journal of the American Medical Association (JAMA 1995; 273: 27-34). In brief, the abstract should be divided into the following sections: (1) Objective; (2) Design - if clinical, to include setting, selection of patients, details on the intervention, outcome measures, etc.; if laboratory research, to include details on methods; (3) Results; (4) Conclusions.

Keywords

Immediately after the abstract, provide a maximum of 6 keywords, using British spelling and avoiding general and plural terms and multiple concepts (avoid, for example, 'and', 'of'). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

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As Archives of Oral Biology is a journal with a multidisciplinary readership, abbreviations, except those universally understood such as mm, g, min. u.v., w/v and those listed below, should be avoided if possible. Examples of abbreviations which may be used without definition are: ADP, AMP, ATP, DEAE-cellulose, DNA, RNA, EDTA, EMG, tris. Other abbreviations used to improve legibility should be listed as a footnote on the title page as well as being defined in both the abstract and the main text on first usage. Chemical symbols may be used for elements, groups and simple compounds, but excessive use should be avoided. Abbreviations other than the above should not be used in titles and even these should be avoided if possible.

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Artwork

Image manipulation

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- Use a logical naming convention for your artwork files.

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Examples:

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Reference to a website:

Cancer Research UK. Cancer statistics reports for the UK. (2003). <http://www.cancerresearchuk.org/aboutcancer/statistics/cancerstatsreport/> Accessed 13 March 2003.

Reference to a dataset:

[dataset] Oguro, M., Imahiro, S., Saito, S., Nakashizuka, T. (2015). *Mortality data for Japanese oak wilt disease and surrounding forest compositions*. Mendeley Data, v1. <https://doi.org/10.17632/xwj98nb39r.1>.

Reference to a conference paper or poster presentation:
Engle, E.K., Cash, T.F., & Jarry, J.L. (2009, November). The Body Image Behaviours Inventory-3: Development and validation of the Body Image Compulsive Actions and Body Image Avoidance Scales. Poster session presentation at the meeting of the Association for Behavioural and Cognitive Therapies, New York, NY.

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- Use of parametric tests when non-parametric tests are required
- Inconsistencies between summary statistics and statistical tests such as giving means and standard deviations for data which were analysed with non-parametric tests.
- Multiple comparisons undertaken with multiple t tests or non-parametric equivalents rather than with analysis of variance (ANOVA) or non-parametric equivalents.
- Post hoc tests being used following an ANOVA which has yielded a non-significant result.
- Incomplete names for tests (e.g. stating "Student's t test" without qualifying it by stating "single sample", "paired" or "independent sample")

- n values being given in a way which obscures how many independent samples there were (e.g. stating simply $n=50$ when 10 samples/measurements were obtained from each of 5 animals/human subjects).
- Stating that $P=0.000$ (a figure which is generated by some computer packages). The correct statement (in this case) is $P<0.0005$.