

UNIVERSIDADE FEDERAL DE MINAS FERAIS
Instituto de Ciências Biológicas
Programa de Pós-Graduação em Biologia Vegetal

Isabelle Cerceau Brandão

**POLINIZAÇÃO, FUNCIONAMENTO FLORAL E AMEAÇAS À CONSERVAÇÃO
DE CACTOS MELITÓFILOS DO RIO GRANDE DO SUL**

Belo Horizonte

2020

Isabelle Cerceau Brandão

**POLINIZAÇÃO, FUNCIONAMENTO FLORAL E AMEAÇAS À CONSERVAÇÃO
DE CACTOS MELITÓFILOS DO RIO GRANDE DO SUL**

Tese apresentada ao Programa de Pós-Graduação em Biologia Vegetal da Universidade Federal de Minas Gerais como requisito parcial para a obtenção do título de doutor em Ciências Biológicas – Biologia Vegetal

Orientador: Prof. Dr. Clemens Schlindwein

Belo Horizonte

2020

043 Brandão, Isabelle Cerceau.
Polinização, funcionamento floral e ameaças à conservação de cactos melitófilos do Rio Grande do Sul [manuscrito] / Isabelle Cerceau Brandão. – 2020.

142 f. : il. ; 29,5 cm.

Orientador: Prof. Dr. Clemens Schlindwein.

Tese (doutorado) – Universidade Federal de Minas Gerais, Instituto de Ciências Biológicas. Programa de Pós-Graduação em Biologia Vegetal.

1. Cactaceae. 2. Polinização. 3. Abelhas. 4. Conservação dos Recursos Naturais. I. Schlindwein, Clemens. II. Universidade Federal de Minas Gerais. Instituto de Ciências Biológicas. III. Título.

CDU: 581



Programa de Pós-Graduação em Biologia Vegetal
Universidade Federal de Minas Gerais
ICB - Departamento de Botânica

UFMG

Tese defendida por Isabelle Cerceau Brandão em 17 de dezembro de 2020 e aprovada pela Banca Examinadora constituída pelos professores:

Dr. Clemens Schindwein (UFMG)

Dr. Christian Westerkamp (Universidade Federal do Cariri)

Dra. Daniela Zappi (Universidade de Brasília)

Dra. Andréa Cardoso de Araujo (Universidade Federal de Mato Grosso do Sul)

Dr. Paulo Milet-Pinheiro (Universidade de Pernambuco)

AGRADECIMENTOS

Agradeço ao meu orientador Clemens Schlindwein por todos ensinamentos durante este período de formação. Ele me ensinou a observar os pequenos detalhes para compreender os sistemas mais complexos. A presença do Clemens em campo foi essencial para meu aprendizado e admiro muito o olhar dele sobre a natureza.

Agradeço ao Samuel, meu companheiro de vida e colega de laboratório, que me acompanhou durante todos os trabalhos de campo e sem a ajuda dele (em casa, no trabalho e na vida) tudo seria mais difícil.

A minha família, em especial minha mãe Rose, meu pai Jacinto, minha irmã Caroline e minha avó Maria que me apoiam sempre.

A Ana Laura, colega de laboratório que se tornou uma grande amiga. Passamos longos meses juntas no campo, compartilhando muitas experiências (boas e ruins!) que serão lembradas por toda a vida.

A Reisla Oliveira pela ajuda em campo, discussões e aprendizados que contribuíram muito na minha formação.

Aos amigos do Laboratório PLEBEIA, Fernanda Figueiredo, Priscilla Araújo, Letícia Pataca, Adriano Valentin, Ana Luísa Cordeiro, que sempre estiveram juntos no dia-a-dia na universidade e compartilharam aprendizados.

Ao João Larocca e toda a equipe do Jardim Botânico de Porto Alegre (Rosana Singer, Ricardo Aranha, Ari Delmo Nilson, Leandro Dal Ri, Josy Matos, Andréa Carneiro) que desenvolveram junto conosco o trabalho de conservação dos cactos do Rio Grande do Sul. Durante nossas expedições de campo descobrimos novas populações de cactos e nos deparamos com desafios para a conservação das espécies, como a coleta ilegal e a criação de animais. Organizamos eventos sobre cactáceas na Universidade Federal do Pampa – Caçapava do Sul e no Jardim botânico de Porto Alegre, que contribuíram muito para minha formação.

Agradeço aos moradores de Minas do Camaquã por todo o apoio logístico durante três anos de trabalho de campo. Matheus e Claudia Pohren, que nos transportaram de trator até as áreas de trabalho, Marlise e família que nos ajudaram a encontrar casas para alugar, Vânia que disponibilizou sua casa durante uma temporada de campo, Tico-Tico que nos ajudou com pneus furados e nos forneceu todos os produtos gaúchos caipiras, será eternamente lembrado pelo bom humor. Agradeço a seu Álvaro que nos deixou trabalhar em suas terras. Agradeço a Marta, diretora da escola de Minas do Camaquã, que nos autorizou a realização de oficinas com os

alunos, onde ensinamos sobre os cactos e abelhas da região. A Mabel que foi uma companheira de campo durante dois anos.

Aos professores, colegas e funcionários do Programa de Pós Graduação em Biologia Vegetal.

Aos professores Eduardo Borba, Vinícius Brito e Cristiane Martins, membros da banca de qualificação, que contribuíram na construção do primeiro artigo dessa tese.

Aos professores Christian Westerkamp, Daniela Zappi, Andrea Cardoso de Araujo e Paulo Milet Pinheiro que participaram da banca de defesa e contribuíram muito para a avaliação e melhoria deste trabalho.

Aos membros da banca de defesa, pela disponibilidade de ler e avaliar esse trabalho.

Ao Grupo Boticário de Proteção à Natureza, que financiou grande parte das expedições de campo, CAPES que concedeu a bolsa durante o doutorado e ICMBIO pelas licenças de coleta concedidas.

RESUMO

Cactaceae é uma família altamente diversa, com cerca de 1450 espécies distribuídas predominantemente no continente americano. O Brasil é um dos centros de diversidade da família e grande proporção das espécies estão ameaçadas de extinção. A alteração dos habitats, a coleta ilegal e a introdução de animais exóticos nas áreas de ocorrência dos cactos induzem o declínio populacional de várias espécies. O conhecimento sobre a biologia das cactáceas em habitat natural pode auxiliar na execução de ações para a conservação de espécies. Dessa maneira, este trabalho teve como objetivo ampliar o conhecimento sobre a biologia de cactáceas melitófilas através do diagnóstico reprodutivo e identificação das principais ameaças em habitat natural. O trabalho foi desenvolvido no Pampa do Rio Grande do Sul, um dos centros de diversidade de cactáceas no Brasil. *Parodia neohorstii* (S.Theun.) N.P.Taylor e *Frailea gracillima* (Lem.) Britton & Rose foram estudadas na perspectiva da biologia da polinização e funcionamento floral. Ambas as espécies apresentam floração sazonal, longevidade floral curta e polinização por abelhas especialistas em pólen (oligoléticas). As abelhas sincronizaram a atividade de voo com o horário de abertura das flores e depositam grandes quantidades de pólen nos estigmas, o que resulta na produção eficiente de frutos e sementes. *Frailea gracillima* apresenta hercogamia temporal, ou seja, quando as flores se abrem, estigma e anteras estão espacialmente separados. Ao longo da antese, lentos movimentos autônomos dos estames e dos lobos estigmáticos diminuem a separação espacial entre as estruturas reprodutivas, o que possibilita a autopolinização autônoma tardia durante o fechamento das flores. Já *P. neohorstii* é uma espécie auto incompatível e suas flores exibem movimentos tigmomásticos dos estames, induzidos pelo contato físico entre as abelhas e os filetes. Os movimentos dos estames escondem momentaneamente parte do conteúdo polínico da flor (75%) e atua como um mecanismo de particionamento de pólen. Mostramos experimentalmente que somente a execução dos movimentos dos estames induzem a senescência floral em *P. neohorstii*, e não a polinização. Na ausência de contato físico com os filetes, os estames não se movimentam de forma autônoma, a longevidade floral é estendida por até 5 dias.

Para a espécie *P. neohorstii*, avaliamos ainda os fatores que impactam a manutenção e ameaçam a conservação de *P. neohorstii* em habitat natural. Avaliamos 3677 indivíduos em 9 localidades de ocorrência. Concluímos que a principal ameaça à espécie é a criação de caprinos e bovinos, 16,9% dos indivíduos amostrados foram impactados por essa atividade. Os principais impactos foram o deslocamento e a remoção dos cactos do substrato, e danos físicos causados

pelo pisoteio e pelo consumo parcial das plantas, o que resultou em 41% de mortalidade. O pastoreio dos animais de criação impactou não somente as espécies foco do estudo, mas também outras populações de cactáceas simpátricas. Para a conservação das comunidades de cactos melitófilos em habitat natural, é necessário a manutenção dos polinizadores especialistas e a redução do acesso de animais de criação nas áreas de ocorrência das espécies.

Palavras-chave: Cactaceae, polinização, tigonastia, interação planta-polinizador, abelhas oligolépticas, movimento de estames, conservação.

ABSTRACT

Cactaceae is a highly diverse family, with 1450 species distributed predominantly on the American continent. Brazil is one of the cacti diversity centers and huge proportion of species are threatened with extinction. Habitat alteration, illegal collection and the introduction of exotic animals induce population decline of several cactus species. Knowledge about the biology of cacti in natural habitat can assist in the execution of conservation actions for the threatened species. Thus, this study aimed to expand knowledge about the biology of melittophilous cacti through reproductive diagnosis and identification of the main threats in natural habitat. The study was developed in the Pampa of Rio Grande do Sul, one of the centers of cactus diversity in Brazil. *Parodia neohorstii* (S.Theun.) N.P.Taylor and *Frailea gracillima* (Lem.) Britton & Rose were studied from the perspective of pollination biology and floral functioning. Both species show seasonal flowering, short floral longevity and pollination by pollen specialists (oligolectic bees). The bees synchronized the flight activity with the opening time of the flowers and deposit huge amounts of pollen on the stigmas, which results in efficient fruit and seed set. *Frailea gracillima* presents temporal herkogamy, that is, when the flowers open, stigma and anthers are spatially separated. Throughout the anthesis, slow autonomous movements of the stamens and stigmatic lobes decrease the spatial separation between reproductive structures, which allows for delayed self-pollination during flower closure. *Parodia neohorstii* is a self-incompatible species and its flowers exhibit thigmonastic stamen movements, induced by physical contact by bees on the stamens. The movements of the stamens momentarily hide part of the flower's pollen content (75%) and act as a pollen partitioning mechanism. We showed experimentally that only the induction of stamen movements induces floral senescence in *P. neohorstii*, and not pollination. In the absence of physical contact, the stamens do not move autonomously and the floral longevity is extended for up to 5 days.

For the species *P. neohorstii*, we also evaluated the factors that impact the specie in natural habitat. We evaluated 3677 individuals in 9 locations. We conclude that the main threat to the species is livestock farming of goats and cattle, 16.9% of the individuals sampled were impacted by this activity. The main impacts were the displacement and removal of cacti from the substrate, and physical damage caused by trampling and partial consumption of plants, which resulted in 41% mortality. The grazing impacted not only the species that were the focus of the study, but also other populations of sympatric cacti. For the conservation of

melittophilous cactus communities in natural habitat, it is necessary to maintain specialist pollinators and reduce access for farm animals in the areas where the species occur.

Keywords: Cactaceae, pollination, thigmonasty, plant-pollinator interaction, oligolectic bees, stamen movement, conservation.

SUMÁRIO

Introdução geral	12
Capítulo 1 - The cost of fidelity: foraging oligolectic bees gather huge amounts of pollen in a highly specialized cactus–pollinator association	28
Capítulo 2 – Stimulation of thigmonastic stamens by bees induce floral senescence in cactus flowers	60
Capítulo 3 – Ameaças à conservação de <i>Parodia neohorstii</i> (Cactaceae), espécie criticamente ameaçada de extinção	86
Capítulo 4 – Reproduction of <i>Frailea gracillima</i> (Cactaceae) in natural habitat: Pollination by solitary bees and mechanism of delayed self-pollination	113
Considerações finais	140

INTRODUÇÃO GERAL

Cactaceae é uma família predominantemente americana e altamente diversa, composta por 1450 espécies distribuídas em 4 subfamílias, Pereskioideae, Opuntioideae, Maihuenioideae e Cactoideae (Anderson, 2001; Hunt *et al.*, 2006, 2013). As espécies são amplamente distribuídas em quatro principais centros de diversidade, sendo eles: 1) México e a região Sul dos Estados Unidos; 2) região central da cordilheira dos Andes, principalmente na Bolívia e Peru; 3) região Leste do Brasil; 4) região Sul do Brasil e áreas adjacentes do Uruguai (Oldfield, 1997; Ribeiro-Silva *et al.*, 2011). Apesar da maior diversidade de espécies de Cactaceae ser encontrada em regiões áridas e semiáridas (Oldfield, 1997; Anderson, 2001), suas espécies ocorrem em diferentes habitats, como florestas tropicais úmidas e campos arbustivos (Martin & James, 2009; Anceschi & Magli, 2018).

No Brasil, ocorrem 261 espécies pertencentes a 39 gêneros, distribuídos em todos os biomas (Zappi, 2020). No bioma Mata Atlântica ocorrem predominantemente cactos epífitos, enquanto que no Pampa existe alta diversidade de cactos rupícolas globosos e cilíndricos (Ribeiro-Silva *et al.*, 2011; Carneiro *et al.*, 2016) na Caatinga e Cerrado, os cactos colunares e arborescentes são os mais abundantes (Oldfield, 1997). No Pantanal e Chaco, os cactos colunares e globosos são os mais representativos (Gomes & Araújo 2015, Zappi *et al.*, 2018). Como nas demais regiões do mundo, uma grande proporção das espécies brasileiras são endêmicas e têm distribuição restrita (Forzza *et al.*, 2010).

O bioma Pampa, que ocupa a porção sul do Rio Grande do Sul é um dos principais centros de diversidade de cactáceas no Brasil, com 65 espécies pertencentes a 11 gêneros, onde destacam-se espécies dos gêneros *Parodia* Speg. e *Frailea* Britton & Rose (Ribeiro-Silva *et al.*, 2011; Carneiro *et al.*, 2016; Saraiva *et al.*, 2014). A região é marcada por grande heterogeneidade ambiental e alta disponibilidade de habitats favoráveis à ocorrência das espécies. Uma característica marcante dos cactos gaúchos é o endemismo, que chega a 16% nas espécies da região (Carneiro *et al.*, 2016). Os cactos do Rio Grande do Sul são coletados e estudados desde a primeira metade do século XIX por naturalistas europeus (Pontes *et al.*, 2017a). Apesar disso, até os dias atuais pouco se sabe sobre a distribuição geográfica, o tamanho e o estado de conservação das populações da maioria das espécies (Ribeiro-Silva *et al.*, 2011). Além disso, existem poucos estudos sobre a biologia reprodutiva e da polinização das espécies desse importante centro de diversidade de cactáceas no país (Schlindwein & Wittmann, 1995, 1997).

34

35 *Breve histórico sobre os cactos do Rio Grande do Sul*

36

37 A singularidade e a diversidade de formas das espécies de cactos atraíram a atenção de
38 botânicos e naturalistas europeus que visitaram o continente americano no início do século XIX
39 (Engelmann, 1856; Marshall & Bock, 1941). Em longas expedições de campo, registradas a
40 partir de 1818, diversas espécies de cactos globosos foram descobertas e coletadas no Rio
41 Grande do Sul e em áreas adjacentes do Uruguai e Argentina. Esses exemplares foram enviados
42 à Europa e serviram como material tipo para a descrição de diversas espécies (Pontes *et al.*,
43 2017a), que posteriormente serviram de subsídio para a publicação de livros e chaves de
44 identificação (Förster & Rümpler, 1886; Schumann, 1890; Schumann & Hirscht, 1899;
45 Spegazzini, 1905; Pontes *et al.*, 2017a).

46 A partir de 1950, cidadãos gaúchos, principalmente descendentes de alemães, passaram
47 a se interessar pelos cactos da região e a estabelecer coleções com espécimes coletados em
48 expedições ao interior do estado (Pontes *et al.*, 2017b). Com conhecimento acumulado sobre
49 espécies novas, os brasileiros forneciam informações e enviavam sementes e cactos para a
50 Europa em troca de literatura especializada, além de serem guias em expedições no Brasil
51 (Pontes *et al.*, 2017b). A partir dos anos 1960, iniciou-se uma grande exportação de cactos para
52 Europa com finalidade comercial (Pontes *et al.*, 2017b). Apesar do ponto negativo da
53 exploração comercial dos cactos no RS, esse período foi marcado por grande quantidade de
54 descrição de espécies e um aumento do conhecimento sobre a diversidade das cactáceas no sul
55 do Brasil (Pontes *et al.*, 2017b). No início dos anos 80, leis ambientais começaram a restringir
56 a exportação de espécies nativas da flora (Pontes *et al.*, 2017b) e nos anos 90, a Convenção
57 sobre Comércio Internacional de Espécies Ameaçadas publicou um documento proibindo o
58 comércio internacional de diversas de espécies de cactáceas, o que resultou em um aumento das
59 apreensões de cactos enviados para o exterior nos anos subsequentes (Hunt, 1992).

60 Apesar de grandes expedições em busca de novas espécies, pouco se sabia sobre a
61 biologia dos cactos em habitat natural. Os primeiros registros sobre a história natural dos cactos
62 do Rio Grande do Sul datam do início da década de 1990 com estudos da polinização de
63 espécies melitófilas (Schlindwein & Wittmann, 1995). O levantamento detalhado das espécies
64 de abelhas da região de Caçapava do Sul demonstrou uma rica apifauna com diversas abelhas
65 especialistas em pólen (oligoléticas), muitas delas ainda não descritas (Schlindwein, 1998). As
66 extensas horas de observação da floração dos cactos em habitat natural revelou interações

67 especializadas com polinizadores (Schlindwein & Wittmann, 1995, 1997). Apesar da grande
68 riqueza de interações entre cactos e polinizadores, até o momento poucos estudos foram
69 desenvolvidos com as espécies em habitat natural.

70 As expedições em busca de novas espécies e populações continuaram, assim como a
71 constante atualização da taxonomia e sistemática do grupo (Carneiro *et al.*, 2016; Pontes *et al.*,
72 2017c; Anceschi & Magli, 2018). Atualmente, no Rio Grande do Sul são registradas 65
73 espécies pertencentes a 11 gêneros, sendo *Parodia* o maior gênero (Carneiro *et al.*, 2016). A
74 descrição mais recente de uma espécie de *Parodia* do Rio Grande do Sul, *Parodia gaucha* M.
75 Machado & Larocca é do ano de 2008 (Machado & Larocca, 2008).

76 Apesar da grande riqueza de espécies do estado, muitas estão ameaçadas de extinção e
77 grande proporção está em declínio populacional (Carneiro *et al.*, 2016; Anceschi & Magli,
78 2018; IUCN, 2020). Um grande esforço para a conservação *ex situ* no Brasil é a coleção de
79 cactáceas do Jardim Botânico de Porto Alegre, onde estudos de germinação e desenvolvimentos
80 de cactos são desenvolvidos com as espécies do Rio Grande do Sul (Carneiro *et al.*, 2016).

81

82 *Conservação de cactos*

83

84 Os cactos são plantas que possuem forte apelo ornamental e por isso muito admiradas e
85 cultivadas em várias parte mundo. A grande procura de cactos para fins ornamentais e a coleta
86 excessiva de indivíduos e sementes para horticultura tornou-se uma grande ameaça para a
87 família (Oldfield, 1997; Ribeiro-Silva *et al.*, 2011). Cerca de 86% dos cactos ameaçados de
88 extinção que estão disponíveis para cultivo são coletados diretamente do habitat natural
89 (Goettsch *et al.*, 2015). Dessa maneira, populações de diversas espécies foram drasticamente
90 reduzidas e até mesmo dizimadas (Anderson, 2001).

91 Naturalmente, os cactos são vulneráveis a alterações em seu habitat devido distribuição
92 restrita, dependência de polinizadores para reprodução e de agentes facilitadores para
93 germinação e estabelecimento (Godínez-Alvarez *et al.*, 2003; Ortega-Baes & Godínez-Alvarez,
94 2006; Mandujano *et al.*, 2010; Saraiva & Souza, 2012). Esses fatores somados à pressão
95 antropogênica tornaram Cactaceae uma das famílias de plantas mais ameaçadas do planeta
96 (Goettsch *et al.*, 2015; IUCN, 2020). A criação de animais impacta fortemente as populações,
97 seja pelo consumo direto dos cactos por cabras ou pelo pisoteio por gado, que causa altas taxas
98 de mortalidade (Schlosser, 1979; Warrick & Krausman, 1989; Machado *et al.*, 2005; Machado,
99 2007; Anceschi & Magli, 2018). Além disso, a conversão de habitat para silvicultura e outras

100 lavouras diminuiu drasticamente as áreas de ocorrência de diversas espécies (Goettsch *et al.*,
101 2015; Carneiro *et al.*, 2016; Anceschi & Magli, 2018).

102 As mudanças climáticas impactam as interações entre planta-polinizador, uma vez que
103 variações na temperatura média entre os anos podem alterar a fenologia das plantas e o período
104 de atividade de polinizadores (Wall *et al.*, 2003; Memmott *et al.*, 2007; Petanidou *et al.*, 2014).
105 Essas alterações são mais graves para as espécies de cactos auto incompatíveis e que mantêm
106 interações com polinizadores especialistas, que normalmente tem períodos restritos de atividade
107 de voo ao longo do ano (Simpson & Neff, 1987; Schlindwein *et al.*, 1995; McIntosh, 2005;
108 Blair & Williamson, 2008; Egli & Giorgetta, 2017).

109 Em nível global, cerca de 31% das espécies de cactos estão ameaçadas de extinção, e
110 dessas, apenas 6% estão amplamente protegidas dentro de unidades de conservação (Goettsch
111 *et al.*, 2015, 2019). O Brasil é um dos países com maior proporção de espécies de cactáceas
112 endêmicas e ameaçadas de extinção (Ortega-Baes & Godínez-Alvarez, 2006; Ortega-Baes *et*
113 *al.*, 2010), além de ser um país com grande quantidade de espécies fora de áreas protegidas
114 (Oldfield, 1997; Goettsch *et al.*, 2019). Assim, é considerado um país prioritário para a
115 conservação de cactos (Ortega-Baes & Godínez-Alvarez, 2006).

116 Visando a conservação das espécies brasileiras, o Plano de Ação Nacional para
117 Conservação das Cactáceas propõe ações para conservação, incluindo estudos demográficos,
118 reprodutivos, genéticos, entre outros (Ribeiro-Silva *et al.*, 2011).

119 Estudos sobre as espécies são importantes para subsidiar as ações de conservação
120 direcionadas. O levantamento de dados fenológicos, reprodutivos e de interação com
121 polinizadores são importantes para compreender a capacidade das espécies de se reproduzirem
122 *in situ*, principalmente aquelas que não se reproduzem vegetativamente. A baixa produtividade
123 de frutos e sementes, além da baixa viabilidade de sementes, pode influenciar negativamente
124 no recrutamento de novos indivíduos e conseqüentemente afetar a estrutura populacional das
125 espécies (Godínez-Alvarez *et al.*, 2003). Dados sobre a distribuição espacial e abundância de
126 indivíduos em habitat natural podem auxiliar na escolha de áreas prioritárias para a conservação
127 das espécies.

128

129 *Interações entre cactos melitófilos e polinizadores especialistas*

130

131 As espécies da família Cactaceae são polinizadas por diferentes grupos funcionais de
132 visitantes florais, como beija-flores, morcegos, abelhas e mariposas (Porsch, 1938; Grant &

133 Grant, 1979; Rowley, 1980; Anderson, 2001; Barthlott *et al.*, 2015). A diversidade de atributos
134 florais nas cactáceas, como cor, forma, odor, tipo de recursos e tamanho estão diretamente
135 relacionados com os grupos de polinizadores que interagem (Grant & Grant, 1979; Mandujano
136 *et al.*, 2010). As flores podem medir de 5 milímetros a 40 centímetros de diâmetro, com estames
137 numerosos, grande quantidade de grãos de pólen e néctar (Anderson, 2001). Muitas flores
138 apresentam forma de disco ou sino, com recursos florais acessíveis aos visitantes florais (Grant
139 & Grant, 1979). Porém, apesar das flores apresentarem morfologia aparentemente generalista
140 com recursos abundantes, podem exibir mecanismos de restrição ao acesso dos recursos florais
141 pelos visitantes, resultando em interações especializadas com polinizadores (Schlindwein &
142 Wittmann, 1997).

143 As interações cactos-polinizadores e aspectos da biologia reprodutiva foram estudadas
144 em aproximadamente 5% das espécies em nível global e 10% das espécies brasileiras
145 (Mandujano *et al.*, 2010; Ribeiro-Silva *et al.*, 2011). Porém, muitas vezes estes estudos foram
146 conduzidos sem observação de visitantes florais e polinizadores em habitat natural. A maioria
147 das espécies de cactos apresentam características florais que favorecem a polinização cruzada,
148 como hercogamia, dicogamia e mecanismos de auto incompatibilidade, tornando-as
149 completamente dependentes de polinizadores para se reproduzirem (Anderson, 2001;
150 Mandujano *et al.*, 2010). Portanto, estudos detalhados sobre o comportamento dos
151 polinizadores são importantes, uma vez que nem todos os visitantes florais contribuem para
152 formação de frutos e sementes (Ollerton *et al.*, 1996; Alves-dos-Santos *et al.*, 2016).

153 Flores melitófilas, polinizadas por abelhas (Faegri & van der Pijl, 1979), são as mais
154 comuns dentre as cactáceas (Anderson, 2001) e diversas espécies podem manter interações
155 especializadas com abelhas oligoléticas (Simpson & Neff, 1987; Schlindwein *et al.*, 1995;
156 McIntosh, 2005; Blair & Williamson, 2008; Eggli & Giorgetta, 2017). A oligoetia corresponde
157 a especialização da dieta polínica de abelhas, na qual as fêmeas coletam pólen exclusivamente
158 em plantas de um mesmo gênero ou família (Robertson, 1925). No Brasil, a oligoetia ocorre
159 em espécies de pelo menos 12 tribos de abelhas, que se relacionam com pelo menos 16 famílias
160 de plantas (Schlindwein, 2004). Devido a especialização na coleta de pólen, as abelhas
161 oligoléticas podem apresentar adaptações morfológicas, fisiológicas e comportamentais
162 relacionadas a interação com sua planta hospedeira (Linsley, 1958; Danforth *et al.*, 2019a).
163 Uma das adaptações é a sincronia temporal entre a emergência das abelhas dos ninhos e a época
164 de floração de suas plantas hospedeiras, além da sincronia da atividade de voo diária com o
165 momento da abertura das flores (Linsley, 1958; Wcislo & Cane, 1996; Schlindwein &

166 Wittmann, 1997; Danforth *et al.*, 2019a). A sincronia de voo com a antese floral é importante
167 na interação com cactáceas, uma vez que maioria das flores são efêmeras e a longevidade floral
168 pode ser de apenas algumas horas (Schlindwein & Wittmann, 1995; Mandujano *et al.*, 2010;
169 Valverde *et al.*, 2015). Outra característica que abelhas oligoléticas podem apresentar são
170 estruturas adaptadas ao manuseio e transporte de grãos de pólen com tamanho grande (Linsley,
171 1958; Thorp, 1979; Danforth *et al.*, 2019a), uma característica recorrente do pólen de cactáceas
172 (Leuenberger, 1976). O tamanho do pólen pode impedir a coleta dos grãos por abelhas que não
173 apresentam estruturas especializadas (Roberts & Vallespir, 1978; Thorp, 1979).

174 Estudos sobre o comportamento de forrageio de abelhas oligoléticas demonstram altas
175 taxas de visitação e constância na coleta de pólen, contribuindo para a polinização efetiva das
176 suas plantas hospedeiras e formação de frutos e sementes com sucesso (Schlindwein *et al.*,
177 2005; Milet-Pinheiro & Schlindwein, 2010; Siriani-Oliveira *et al.*, 2018). A eficiência de coleta
178 de pólen está diretamente relacionada ao sucesso reprodutivo das abelhas oligoléticas, uma vez
179 que quanto mais pólen for coletado por unidade de tempo, maior a quantidade de células de cria
180 para provisão larval (Strickler, 1979; Schlindwein *et al.*, 2005). Ninhos de abelhas
181 oligoléticas em flores de *Opuntia* mostram elevada pureza no conteúdo polínico, cerca de 99.5
182 a 100% é composto por pólen de cactáceas (Schlindwein & Wittmann, 1997). Apesar das
183 grandes quantidades de pólen coletadas pelas abelhas oligoléticas e transportadas para o ninho,
184 a deposição de pólen no estigma é alta e suficiente para atingir a produção máxima de sementes.
185 Muitas abelhas utilizam o estigma dos cactos como plataforma de pouso, o que favorece a
186 deposição de pólen xenógamo previamente depositado na superfície do corpo do polinizador
187 (Schlindwein & Wittmann, 1997; Blair & Williamson, 2008; Lenzi & Orth 2011).

188 Abelhas oligoléticas podem exibir comportamento reprodutivo atrelado a flores de
189 cactos (Schlindwein & Wittmann, 1997; Oliveira, 2015). Ao aguardarem dentro das flores,
190 machos de diversas linhagens de abelhas solitárias têm a oportunidade de acasalar com fêmeas
191 que visitam flores em busca de recursos (Danforth *et al.*, 2019b). Os machos podem se camuflar
192 entre estames para esperar pelas fêmeas (Schlindwein & Wittmann, 1995; Oliveira, 2015) ou
193 patrulhar uma sequência de flores em busca de parceiras e defender territórios de machos
194 invasores (Oliveira & Schlindwein, 2010). Cópulas podem ocorrer dentro das flores, o que
195 potencializa o contato entre as abelhas e as estruturas reprodutivas florais e contribui para o
196 fluxo polínico entre fores coespecíficas (Grant & Hurd, 1979; Schlindwein & Wittmann, 1995,
197 1997; McIntosh, 2005).

198 Devido à grande demanda de pólen por abelhas, a coleta excessiva sem transferência
199 polínica entre indivíduos pode ser prejudicial às plantas em algumas interações (Westerkamp,
200 1996; Hargreaves *et al.*, 2009). Abelhas pequenas podem coletar recursos em flores de cactos
201 sem tocar nos estigmas, assim, muitas vezes podem atuar como pilhadoras de pólen (McFarland
202 *et al.*, 1989; Osborn *et al.*, 1988; Schlindwein & Wittmann, 1997). Como flores de cactos
203 produzem grandes quantidades de pólen em numerosos estames (Anderson, 2001; Mandujano
204 *et al.*, 2010), algumas espécies de abelhas podem preencher as escopas em uma única visita
205 floral sem contribuir para polinização, como observado na interação entre *Opuntia phaeacantha*
206 (Engelm.) (Cactaceae) e a abelha oligolética *Perdita texana* (Cresson) (Andrenidae) (Barrows
207 *et al.*, 1976). Em contrapartida, alguns mecanismos florais podem limitar a quantidade de pólen
208 coletado pelos visitantes e favorecer a polinização cruzada (Westerkamp, 1996; Harder &
209 Wilson, 1997; Schlindwein & Wittmann, 1997), por exemplo, através de movimento
210 tigonástico de estames. Os estames sensitivos de cactos, especialmente do gênero *Opuntia*,
211 se movem em direção ao estilete após o contato do visitante floral com os filetes (Schlindwein
212 & Wittmann, 1997; Lenzi & Orth, 2011; Cota-Sánchez *et al.*, 2013). Esse movimento atua como
213 um mecanismo de particionamento na oferta de pólen, uma vez que os estames maiores
214 recobrem os estames menores, escondendo cerca de 80% do recurso disponível de uma flor
215 (Schlindwein & Wittmann, 1997). Com a limitação a quantidade de grãos de pólen coletado a
216 cada visita floral, há um aumento da quantidade de eventos de exportação polínica e
217 consequentemente um aumento do sucesso masculino da planta (Harder & Thomson, 1989;
218 Westerkamp, 1996; Harder & Wilson, 1997).

219 O movimento dos estames em cactos também tem outros papéis importantes na
220 interação com polinizadores. Pode atuar como um mecanismo de deposição de grãos de pólen
221 no corpo de visitantes florais (Toumey, 1899; Lenzi & Orth, 2011), direcionar o movimento
222 dos insetos dentro das flores (Negrón-Ortiz, 1998) e favorecer o uso do estigma como
223 plataforma de pouso (Grant & Hurd, 1979). Esses movimentos ocorrem em pelo menos 10
224 gêneros de cactáceas em duas subfamílias (Jeffries, 1966; Grant & Hurd, 1979; Rauh, 1979;
225 Kiesling, 1982; Jeffries, 1991), mas apenas espécies de *Opuntia* foram estudadas sob ponto de
226 vista da polinização (Schlindwein & Wittmann, 1997; Lenzi & Orth, 2011).

227 Enquanto grande proporção das cactáceas é dependente de polinizadores, uma pequena
228 porção é independente de vetores de pólen para se reproduzir sexualmente (Mandujano *et al.*,
229 2010). Algumas espécies apresentam mecanismos de autopolinização autônoma, que pode
230 garantir a reprodução em ambientes com baixa densidade de polinizadores (Nassar *et al.*, 2007;

231 Mandujano *et al.*, 2010). Pequena proporção de espécies apresenta cleistogamia, uma estratégia
232 reprodutiva que garante independência de vetores de pólen, uma vez que ocorre a
233 autopolinização sem que as flores se abram (Lord, 1981; Richards, 1997). A cleistogamia foi
234 reportada em espécies de quatro gêneros de cactos, sendo mais frequente em *Frailea* (Cota-
235 Sánchez, 2004; Barthlott & Hunt, 1993; Taylor, 1991; Putnam, 1968).

236 Diante do exposto, essa tese teve como objetivo estudar aspectos da biologia da
237 polinização e do funcionamento floral de duas espécies de cactáceas melitófilas do Rio Grande
238 do Sul, *Parodia neohorstii* (S.Theun.) N.P.Taylor e *Frailea gracillima* (Lem.) Britton & Rose,
239 bem como avaliar as principais ameaças a conservação de *P. neohorstii* em habitat natural.

REFERÊNCIAS

- 240
- 241
- 242 **Alves-dos-Santos I, Silva CID, Pinheiro M, Kleinert ADMP.** 2016. Quando um visitante
243 floral é um polinizador? *Rodriguésia* **67**: 295-307.
- 244 **Anceschi G, Magli A.** 2018. A synopsis of the genus *Parodia* Spegazzini s.l. (Cactaceae).
245 *Bradleya* **1**: 70-161.
- 246 **Anderson EF.** 2001. *The cactus family*. Timber Press (OR). 776 pp.
- 247 **Barrows EM, Chabot MR, Michener CD, Snyder TP.** 1976. Foraging and mating behavior
248 in *Perdita texana* (Hymenoptera: Andrenidae). *Journal of the Kansas Entomological*
249 *Society* **49**: 275-279.
- 250 **Barthlott W, Hunt DR.** 1993. Cactaceae. pp 161 – 197. In: **Kubitzki K, Rohwer JG, Bittrich**
251 **V.** (eds) *Flowering Plants Dicotyledons. The Families and Genera of Vascular Plants*, v.2
252 Springer, Berlin, Heidelberg. 653 pp.
- 253 **Barthlott W, Burstedde K, Geffert JL, Ibisch PL, Korotkova N, Miebach A, Rafiqpoor**
254 **MD, Stein A, Mutke J.** 2015. Biogeography and Biodiversity of Cacti. *Schumannia* **7**. 205
255 pp.
- 256 **Blair AW, Williamson, PS.** 2008. Effectiveness and importance of pollinators to the star cactus
257 (*Astrophytum asterias*). *The Southwestern Naturalist* **53**: 423-430.
- 258 **Carneiro AM, Farias-Singer R, Ramos RA, Nilson AD.** 2016. *Cactos do Rio Grande do Sul*.
259 Fundação Zoobotânica do Rio Grande do Sul, Porto Alegre. 224 pp.
- 260 **Carvalho AT, Schlindwein C.** 2011. Obligate association of an oligolectic bee and a seasonal
261 aquatic herb in semi-arid north-eastern Brazil. *Biological Journal of the Linnean Society*
262 **102**: 355-368.
- 263 **Cota-Sánchez JH.** 2004. Vivipary in the Cactaceae: its taxonomic occurrence and biological
264 significance. *Flora-Morphology, Distribution, Functional Ecology of Plants* **199**: 481-490.
- 265 **Cota-Sánchez JH, Almeida OJG, Falconer DJ, Choi HJ, Bevan L.** 2013. Intriguing
266 thigmonastic (sensitive) stamens in the Plains Prickly Pear *Opuntia polyacantha*
267 (Cactaceae). *Flora - Morphology, Distribution, Functional Ecology of Plants* **208**: 381-
268 389.
- 269 **Danforth B, Minckley R, Neff J, Fawcett F.** 2019a. Bees and Plants: Love Story, Arms Race,
270 or Something in Between? In *The Solitary Bees: Biology, Evolution, Conservation*. pp. 289
271 - 317. Princeton; Oxford: Princeton University Press. 448 pp.

- 272 **Danforth B, Minckley R, Neff J, Fawcett F.** 2019b. Alternative male mating tactics: The race
 273 to be first and the race do be the last. In: *The Solitary Bees: Biology, Evolution,*
 274 *Conservation.* pp. 70 - 92. Princeton; Oxford: Princeton University Press. 448 pp.
- 275 **Eggl U, Giorgetta M.** 2017. Flowering phenology and observations on the pollination biology
 276 of South American cacti. 3. Temporally robust sequential “big bang” flowering of two
 277 unrelated sympatric globular cacti in northern Argentina. *Haseltonia* **23**: 97-109.
- 278 **Engelmann G.** 1856. *Cactaceae of the Boundary.* US Government Printing Office. 153 pp.
- 279 **Faegri K, Van der Pijl L.** 1979. *The Principles of Pollination Ecology.* 3rd ed. Pergamon
 280 Press, Oxford. 244pp.
- 281 **Förster CF, Rümpler KT.** 1886. *Handbuch der Kakteenkunde* 2. Leipzig: pp. 563-564.
- 282 **Forzza RC, Baumgratz JFA, Bicudo CEM, Canhos DAL, Carvalho JrAA, Costa AF,**
 283 **Costa DP, Hopkins M, Leitman PM, Lohmann LG, Maia LC, Martinelli G, Menezes**
 284 **M, Morim MP, Nadruz-Coelho MA, Peixoto AL, Pirani JR, Prado J, Queiroz LP,**
 285 **Souza VC, Stehmann JR, Sylvestre L, Walter BMT, Zappi D.** 2010. *Catálogo de*
 286 *plantas e fungos do Brasil.* 2 vols. Andrea Jakobsson Estúdio / Jardim Botânico do Rio de
 287 Janeiro, Rio de Janeiro. 1699 pp.
- 288 **Godínez-Álvarez H, Valverde T, Ortega-Baes P.** 2003. Demographic trends in the Cactaceae.
 289 *The Botanical Review* **69**: 173-201.
- 290 **Goettsch B, Hilton-Taylor C, Cruz-Pinon G, Duffy JP, Frances A, Hernandez HM, Inger**
 291 **R, Pollock C, Schipper J, Superina M, Taylor NP, Tognelli M, Abba AM, Arias S,**
 292 **Arreola-Nava HJ, Baker MA, Barcenás RT, Barrios D, Braun P, Butterworth CA,**
 293 **Burquez A, Caceres F, Chazaro-Basanez M, Corral-Diaz R, Del Valle Perea M, Demaio**
 294 **PH, Duarte de Barros WA, Duran R, Yancas LF, Felger RS, Fitz-Maurice B, Fitz-**
 295 **Maurice WA, Gann G, Gomez-Hinostrosa C, Gonzales-Torres LR, Patrick Griffith M,**
 296 **Guerrero PC, Hammel B, Heil KD, Hernandez-Oria JG, Hoffmann M, Ishihara MI,**
 297 **Kiesling R, Larocca J, Leon-de la Luz JL, Loaiza SC, Lowry M, Machado MC, Majure**
 298 **LC, Avalos JG, Martorell C, Maschinski J, Mendez E, Mittermeier RA, Nassar JM,**
 299 **Negrón-Ortiz V, Oakley LJ, Ortega-Baes P, Ferreira AB, Pinkava DJ, Porter JM,**
 300 **Puente-Martinez R, Gamarra JR, Perez PS, Martinez ES, Smith M, Manuel**
 301 **Sotomayor MDCJ, Stuart SN, Muñoz JL, Terrazas T, Terry M, Trevisson M, Valverde**
 302 **T, Van Devender TR, Veliz-Perez ME, Walter HE, Wyatt SA, Zappi D, Alejandro**
 303 **Zavala-Hurtado J, Gaston KJ.** 2015. High proportion of cactus species threatened with
 304 extinction. *Nature Plants* **1**: 15142.

- 305 **Goettsch B, Duran AP, Gaston KJ.** 2019. Global gap analysis of cactus species and priority
306 sites for their conservation. *Conservation Biology* **33**: 369-376.
- 307 **Gomes VG, Araújo AC.** 2015. Cacti species from the Brazilian Chaco: floral and fruit traits.
308 *Gaia Scientia* **9**: 1-8.
- 309 **Grant V, Grant KA.** 1979. The pollination spectrum in the southwestern American cactus
310 flora. *Plant Systematics and Evolution* **133**: 29-37.
- 311 **Grant V, Hurd P.** 1979. Pollination of the southwestern Opuntias. *Plant Systematics and*
312 *Evolution* **133**: 15-28.
- 313 **Harder LD, Thomson JD.** 1989. Evolutionary options for maximizing pollen dispersal of
314 animal-pollinated plants. *The American Naturalist* **133**: 323-344.
- 315 **Harder LD, Wilson W.** 1996. Theoretical perspectives on pollination. *Acta Horticulturae* **437**:
316 83-102.
- 317
- 318 **Hargreaves AL, Harder LD, Johnson SD.** 2009. Consumptive emasculation: the ecological
319 and evolutionary consequences of pollen theft. *Biological Reviews* **84**: 259-276.
- 320 **Hunt D.** 1992. CITES Cactaceae checklist. Royal Botanic Gardens.
- 321 **Hunt D, Taylor NP, Charles G.** 2006. *The New Cactus Lexicon*. 2 vols. DH Books, Milborne
322 Port. 900 pp.
- 323 **Hunt D, Taylor NP, Charles G.** 2013. *The New Cactus Lexicon*, Illustrations. 2nd ed. DH
324 Books, Milborne Port. 527 pp.
- 325 **IUCN.** 2020. The IUCN Red List of Threatened Species. Version 2020-2.
326 <https://www.iucnredlist.org>
- 327 **Jeffries L.** 1966. Stamen irritability in Cacti. *The Cactus and Succulent Journal of Great*
328 *Britain* **28**: 78-79.
- 329 **Jeffries L.** 1991. *Pereskia* Flowers have Irritable Stamens: Is this a Feature of all
330 Opuntioideae? *British Cactus and Succulent Journal* **9**: 5.
- 331 **Kiesling R.** 1982. The genus *Pterocactus*. *The Cactus and Succulent Journal of Great Britain*
332 **44**: 51-56.
- 333 **Lenzi M, Orth AI.** 2011. Visitantes florais de *Opuntia monacantha* (Cactaceae) em restingas
334 de Florianópolis, SC, Brasil. *Acta Biológica Paranaense* **40**: 19-32.
- 335 **Leuenberger BE.** 1976. Pollen Morphology of the Cactaceae: An SEM-Survey of exine
336 sculpturing and its tentative implications for taxonomy and phylogeny. *The Cactus and*
337 *Succulent Journal of Great Britain*, **38**: 79-94.

- 338 **Linsley E.** 1958. The ecology of solitary bees. *Hilgardia* **27**: 543-599.
- 339 **Lord EM.** 1981. Cleistogamy: a tool for the study of floral morphogenesis, function and
340 evolution. *The Botanical Review* **47**: 421-449.
- 341 **Machado MC, Nyffeler R, Eggli U, Larocca JF.** 2008. A New Species of *Parodia* (Cactaceae,
342 Notocactaceae) from Rio Grande do Sul, Brazil. *A Journal for Botanical Nomenclature* **18**:
343 214-219.
- 344 **Machado M.** 2007. Fascinating *Frailea*, Part I: General impressions. *Cactus World* **25**: 5-11.
- 345 **Mandujano MdC, Carrillo-Angeles I, Martínez-Peralta C, Golubov J.** 2010. Reproductive
346 biology of Cactaceae. pp. 197-230. In: **Ramawat KG.** Eds *Desert plants: biology and*
347 *biotechnology*. Berlin: Heidelberg, Springer-Verlag. 503 pp.
- 348 **Marshall WT, Bock TM.** 1941. Cactaceae: With Illustrated Keys of All Tribes, Sub-tribes and
349 Genera. Abbey Garden Press. 227 pp.
- 350 **Martin LA, James G.** 2009. Unusual habitats, unusual plants. *Cactus and Succulent*
351 *Journal*, **81**:106-112.
- 352 **McFarland JD, Kevan PG, Lane MA.** 1989. Pollination biology of *Opuntia imbricata*
353 (Cactaceae) in southern Colorado. *Canadian Journal of Botany* **67**: 24-28.
- 354 **McIntosh ME.** 2005. Pollination of two species of *Ferocactus*: interactions between cactus-
355 specialist bees and their host plants. *Functional Ecology* **19**: 727-734.
- 356 **Memmott J, Craze PG, Waser NM, Price MV.** 2007. Global warming and the disruption of
357 plant-pollinator interactions. *Ecology Letters* **10**: 710-717.
- 358 **Milet-Pinheiro P, Schlindwein C.** 2010. Mutual reproductive dependence of distylic *Cordia*
359 *leucocephala* (Cordiaceae) and oligolectic *Ceblurgus longipalpis* (Halictidae, Rophitinae)
360 in the Caatinga. *Annals of Botany* **106**: 17-27.
- 361 **Nassar JM, Ramírez N, Lampo M, González JA, Casado R, Nava F.** 2007. Reproductive
362 biology and mating system estimates of two Andean melocacti, *Melocactus schatzlii* and
363 *M. andinus* (Cactaceae). *Annals of Botany* **99**: 29-38.
- 364 **Negrón-Ortiz V.** 1998. Reproductive biology of a rare cactus, *Opuntia spinosissima*
365 (Cactaceae), in the Florida Keys: why is seed set very low? *Sexual Plant Reproduction* **11**:
366 208-212.
- 367 **Oldfield S.** 1997. Cactus and succulent plants: status survey and conservation action plan.
368 International Union for Conservation of Nature and Natural Resources (IUCN).
- 369 **Oliveira R, Schlindwein C.** 2010. Experimental demonstration of alternative mating tactics of
370 male *Ptilothrix fructifera* (Hymenoptera, Apidae). *Animal Behaviour* **80**: 241-247.

- 371 **Oliveira R.** 2015. Estratégias de acasalamento em abelhas solitárias no Brasil: o que os machos
 372 fazer para se acasalar? pp. 439-457. In: **Aguiar AJC, Gonçalves RB, Ramos KS** eds.
 373 *Ensaio sobre as abelhas da Região Neotropical*. Curitiba: Editora UFPR. 456 pp.
- 374 **Ollerton J.** 1996. Reconciling ecological processes with phylogenetic patterns: the apparent
 375 paradox of plant--pollinator Systems. *Journal of Ecology* **84**: 767-769.
- 376 **Ortega-Baes P, Godínez-Alvarez H.** 2006. Global diversity and conservation priorities in the
 377 Cactaceae. *Biodiversity & Conservation* **15**:817-827.
- 378 **Ortega-Baes P, Sühling S, Sajama J, Sotola E, Alonso-Pedano M, Bravo S, Godínez-**
 379 **Alvarez H.** 2010. Diversity and conservation in the cactus family. pp. 157-173. In *Desert*
 380 *plants*. Springer, Berlin, Heidelberg. 503pp.
- 381 **Osborn MM, Kevan PG, Lane MA.** 1988. Pollination biology of *Opuntia polyacantha* and
 382 *Opuntia phaeacantha* (Cactaceae) in southern Colorado. *Plant Systematics and Evolution*
 383 **159**: 85-94.
- 384 **Petanidou T, Kallimanis AS, Sgardelis SP, Mazari, AD, Pantis JD, Waser NM.** 2014.
 385 Variable flowering phenology and pollinator use in a community suggest future
 386 phenological mismatch. *Acta Oecologica* **59**:104-111.
- 387 **Pontes RC, Marchiori JNC, Neto LW.** 2017. Notas históricas sobre a família Cactaceae no
 388 Rio Grande do Sul (Brasil) e Uruguai. I–Período Clássico (1818-1950): viajantes
 389 naturalistas e botânicos europeus. *Balduinia* **56**: 01-11.
- 390 **Pontes RC, Marchiori JNC, Neto LW.** 2017. Notas históricas sobre a família Cactaceae no
 391 Rio Grande do Sul (Brasil) e Uruguai. II – Período moderno (1950-1980): Colecionadores
 392 e exploradores da região e imigrantes. *Balduinia* **57**: 01-17.
- 393 **Pontes RC, Marchiori JNC, Neto LW.** 2017. Notas históricas sobre a família Cactaceae no
 394 Rio Grande do Sul (Brasil) e Uruguai. III–Período contemporâneo (1980-2017):
 395 Taxonomistas gaúchos, uruguaios e europeus. *Balduinia* **58**: 1-9.
- 396 **Porsch O.** 1938. Deutsche Kakteen-Gesellschaft. *Das Bestäubungsleben der Kakteenblüte*.
- 397 **Putnam EW.** 1968. The Genus *Frailea*. *The National Cactus and Succulent Journal* **23**: 54-
 398 56.
- 399 **Rauh W.** 1979. Kakteen an ihren Standorten. — Berlin, Hamburg: Parey. 230 pp.
- 400 **Ribeiro-Silva S, Zappi DC, Taylor N, Machado M.** 2011. Plano de ação nacional para a
 401 conservação das cactáceas. Série Espécies Ameaçadas, 24.
- 402 **Richards AJ.** 1997. *Plant breeding systems*. Garland Science. 529 pp.

- 403 **Roberts RB, Vallespir SR.** 1978. Specialization of hairs bearing pollen and oil on the legs of
 404 bees (Apoidea: Hymenoptera). *Annals of the Entomological Society of America*, **71**: 619-
 405 627.
- 406 **Robertson C.** 1925. Heterotropic Bees. *Ecology* **6**: 412-436.
- 407 **Rowley G.** 1980. Pollination syndromes and cactus taxonomy. *The Cactus and Succulent*
 408 *Journal of Great Britain*, **42**: 95-98.
- 409 **Saraiva DD, Souza AF.** 2012. Effects of environmental factors and plantation forests on
 410 endangered cactus diversity and composition in subtropical South American
 411 grasslands. *Perspectives in Plant Ecology, Evolution and Systematics* **14**: 267-274.
- 412 **Saraiva DD, de Sousa KDS, Overbeck GE.** 2015. Multiscale partitioning of cactus species
 413 diversity in the South Brazilian grasslands: Implications for conservation. *Journal for*
 414 *Nature Conservation* **24**:117-122.
- 415 **Schlindwein C, Wittmann D.** 1995. Specialized solitary bees as effective pollinators of South
 416 Brazilian species of *Notocactus* and *Gymnocalycium* (Cactaceae). *Bradleya* **13**: 25-34.
- 417 **Schlindwein C, Wittmann D.** 1997. Stamen movements in flowers of *Opuntia* (Cactaceae)
 418 favour oligolectic pollinators. *Plant Systematics and Evolution* **204**: 179-193.
- 419 **Schlindwein C.** 1998. Frequent Oligolecty Characterizing a Diverse Bee–Plant Community in
 420 a Xerophytic Bushland of Subtropical Brazil. *Studies on Neotropical Fauna and*
 421 *Environment* **33**: 46-59.
- 422 **Schlindwein C.** 2004. Abelhas solitárias e flores: especialistas são polinizadores efetivos?
 423 In *Anais do 55 Congresso Nacional de Botânica* 1-8.
- 424 **Schlindwein C, Wittmann D, Martins CF, Hamm A, Siqueira JA, Schiffler, D, Machado**
 425 **IC.** 2005. Pollination of *Campanula rapuncululus* L. (Campanulaceae): How much pollen
 426 flows into pollination and into reproduction of oligolectic pollinators? *Plant Systematics and*
 427 *Evolution* **250**: 147-156.
- 428 **Schlosser H.** 1979. Allgemeines über Uruguay und einige seiner Kakteen. *Kakteen und andere*
 429 *Sukkulente* 213-217.
- 430 **Schumann K, Hirscht K.** 1899. Gesamtbeschreibung der Kakteen (Monographia
 431 Cactacearum). J. Neumann. 823 pp.
- 432 **Schumann K.** 1890. Cactaceae. In: *Martius C.P, von Eichler AW, Urban I. Flora Brasiliensis*
 433 **4**: 185-322.
- 434 **Simpson BB, Neff JL.** 1987. Pollination Ecology in the Southwest. *Journal of Systematic and*
 435 *Evolutionary Botany* **11**: 417-440.

- 436 **Siriani-Oliveira S, Oliveira R, Schlindwein C.** 2018. Pollination of *Blumenbachia amana*
 437 (Loasaceae): flower morphology and partitioned pollen presentation guarantee a private
 438 reward to a specialist pollinator. *Biological Journal of the Linnean Society* **124**: 479-491.
- 439 **Spegazzini CL.** 1905. Cactacearum Platensium Tentamen. *Anales Del Museo Nacional de*
 440 *Buenos Aires* **11**:477-521.
- 441 **Strickler K.** 1979. Specialization and foraging efficiency of solitary bees. *Ecology* **60**: 998-
 442 1009.
- 443 **Taylor NP.** 1991. The genus *Melocactus* (Cactaceae) in Central and South America. *Bradleya*
 444 **9**: 1-80.
- 445 **Thorp RW.** 1979. Structural, behavioral, and physiological adaptations of bees (Apoidea) for
 446 collecting pollen. *Annals of the Missouri Botanical Garden* 788-812.
- 447 **Toumey JW.** 1899. Sensitive stamens in the genus *Opuntia*. *The Asa Gray Bulletin* **7**: 35-37.
- 448 **Valverde T, Quijas S, López-Villavicencio M, Castillo S.** 2004. Population dynamics of
 449 *Mammillaria magnimamma* Haworth. (Cactaceae) in a lava-field in central Mexico. *Plant*
 450 *Ecology* **170**: 167-184.
- 451 **Valverde PL, Jiménez-Sierra C, López-Ortega G, Zavala-Hurtado JA, Rivas-Arancibia**
 452 **S, Rendón-Aguilar B, Pérez-Hernández MA, Cornejo-Romero A, Carrillo-Ruiz, H.**
 453 2015. Floral morphometry, anthesis, and pollination success of *Mammillaria pectinifera*
 454 (Cactaceae), a rare and threatened endemic species of Central Mexico. *Journal of Arid*
 455 *Environments* **116**: 29-32.
- 456 **Wall MA, Timmerman-Erskine M, Boyd RS.** 2003. Conservation impact of climatic
 457 variability on pollination of the Federally endangered plant, *Clematis socialis*
 458 (Ranunculaceae). *Southeastern Naturalist* **2**:11–24.
- 459 **Warrick GD, Krausman PR.** 1989. Barrel cacti consumption by desert bighorn sheep. *The*
 460 *Southwestern Naturalist* **34**: 483-486.
- 461 **Wcislo WT, Cane JH.** 1996. Floral resource utilization by solitary bees (Hymenoptera:
 462 Apoidea) and exploitation of their stored foods by natural enemies. *Annual review of*
 463 *entomology* **41**: 257-286.
- 464 **Westerkamp CH.** 1996. Pollen in bee-flower relations some considerations on melittophily.
 465 *Botanica Acta* **109**: 325-332.
- 466 **Zappi DC, Taylor NP, Damasceno JGA, Pott VJ, Machado MC.** 2018. Check-list das
 467 Cactaceae do estado do Mato Grosso do Sul, Brasil. *Iheringia. Série Botânica*, **73**: 169-
 468 173.

469 **Zappi D, Taylor N.** 2020. Cactaceae in Flora do Brasil 2020 em construção. Jardim Botânico
470 do Rio de Janeiro. Disponível em: <<http://reflora.jbrj.gov.br/reflora/floradobrasil/FB70>>.
471 Acesso em: 08 set. 2020
472
473

CAPÍTULO 1

474

475

476

477

478

479

480

481 **THE COST OF FIDELITY: FORAGING OLIGOLECTIC BEES GATHER HUGE**482 **AMOUNTS OF POLLEN IN A HIGHLY SPECIALIZED CACTUS–POLLINATOR**483 **ASSOCIATION¹**

484

¹ A publicação original está disponível em <https://academic.oup.com/biolinnean/article-abstract/128/1/30/5520426>. Publicado como: Cerceau, I., Siriani-Oliveira, S., Dutra, A. L., Oliveira, R., & Schlindwein, C. (2019). The cost of fidelity: foraging oligolectic bees gather huge amounts of pollen in a highly specialized cactus–pollinator association. *Biological Journal of the Linnean Society*, 128(1), 30–43.

511 **INTRODUCTION**

512

513 Despite numerous adaptations of flowering plants to improve pollen transfer, only a
514 small portion of the pollen produced by a flower reaches the stigmatic surface of a conspecific
515 individual, while a huge amount is excluded from plant reproduction (Inouye *et al.*, 1994). In
516 bee-pollinated pollen-flowers, the majority of pollen is transported to bee nests as larval food
517 while only a minor portion reaches stigmas (Harder & Wilson, 1996; Schlindwein *et al.*, 2005;
518 Müller *et al.*, 2006). Some plants are pollinated by oligolectic bees, which are pollen specialists
519 that collect pollen exclusively from the same genus or family (Robertson, 1925). In these
520 pollination systems, the bees acquire for larval supply often more than 85% of the pollen
521 produced by a flower (Schlindwein & Martins, 2000; Schlindwein *et al.*, 2005; Carvalho &
522 Schlindwein, 2011; Pick & Schlindwein, 2011). Oligolectic bees exhibit high flower fidelity
523 often with high flower visitation rates and, despite the removal of huge amounts of pollen from
524 host flowers, generally deposit a sufficient amount of conspecific pollen grains on the stigmatic
525 surface for maximal fruit and seed set (Schlindwein *et al.*, 2005; Milet-Pinheiro & Schlindwein,
526 2010; Carvalho & Schlindwein, 2011; Pick & Schlindwein, 2011; Siriani-Oliveira *et al.*, 2018).
527 Pollinator behavior between collecting pollen from a donor flower and arrival at the stigmas of
528 conspecific individuals is crucial, since the fate of pollen grains directly affects reproductive
529 success (Johnson *et al.*, 2005). In self-incompatible plant species that are threatened with
530 extinction, pollinator effectiveness is vital for maintaining viable populations over time (Bond,
531 1994).

532 Cactaceae is the fifth most threatened botanical family, with almost one third of the
533 species being classified as endangered (Goettsch *et al.*, 2015). The majority of studied cacti
534 species are self-incompatible and depend on pollinators to reproduce (Osborn *et al.*, 1988;
535 McFarland *et al.*, 1989; Boyle, 1997; McIntosh, 2002; McIntosh, 2005; Strong & Williamson,
536 2007; Mandujano *et al.*, 2010; Martinez-Peralta *et al.*, 2014; Martins & Freitas, 2018).
537 Melittophilous cacti apparently have phenotypically generalized flowers — they possess floral
538 traits that are attractive and accessible to a wide range of floral visitors (Ollerton *et al.*, 2007),
539 due to radial symmetry, and wide-open bowl-shaped corollas that make abundant pollen
540 rewards accessible. Surprisingly, however, several melittophilous cacti species maintain
541 relationships with specialized pollinators, such as oligolectic bees (Grant & Hurd, 1979;
542 Simpson & Neff, 1987; Osborn *et al.*, 1988; McFarland *et al.*, 1989; Schlindwein & Wittmann,
543 1995, 1997; McIntosh, 2005; Blair & Williamson, 2008; Roig-Alsina & Schlumpberger, 2008)

544 that exhibit foraging and reproductive behaviors in association with cactus flowers, and which
545 have been reported as the main pollinators for several genera (Simpson & Neff, 1987;
546 Schlindwein & Wittmann, 1995, 1997; Roig-Alsina & Schlumpberger, 2008; Eggli &
547 Giorgetta, 2017).

548 The genus *Parodia* contains 62 species, 33 of which are native to Rio Grande do Sul,
549 the southernmost state of Brazil (Anceschi & Magli, 2018). This region was identified as a
550 hotspot for threatened cactus species (Goettsch *et al.*, 2015), most of which grow outside of
551 protected areas (Goettsch *et al.*, 2018). Oligolectic bees were found to be effective pollinators
552 of three species of *Parodia* Speg. in the region (cited as *Notocactus* (K.Schum.) Frič)
553 (Schlindwein & Wittmann, 1995), and recorded as floral visitors of other *Parodia* species not
554 yet studied (Schlindwein, 1998).

555 We studied the relationships between the endemic red-listed cactus *Parodia neohorstii*
556 (S.Theun.) N.P.Taylor and oligolectic bees in the Pampa region of South Brazil. *Parodia*
557 *neohorstii* is classified as Critically Endangered (IUCN) due to its restricted distribution,
558 population decline and fragmented populations (Larocca, 2013). We focused on the role of
559 specialist bees for the reproduction of this species and asked: 1) What is the breeding system of
560 *P. neohorstii*? 2) Are oligolectic bees more effective pollinators and more efficient pollen
561 collectors than other floral visitors? 3) If so, what are the set of floral traits and feeding
562 behaviors of oligolectic bees that mediate this relationship? 4) What is the fate of pollen during
563 dispersal by flower visitors (stigmatic deposition, pollen removal by bees or removal failure)?
564 5) Is there pollinator limitation and, as a consequence, an insufficient supply of pollen to the
565 stigmas that contributes to threatening the survival of the species?

566 To answer these questions, we characterized the floral biology and breeding system of
567 *P. neohorstii*, analyzed the behavior of its flower visitors, assessed the fate of its pollen and
568 quantified the deposition of heterospecific pollen grains on the stigmatic surface.

569

570 MATERIAL AND METHODS

571

572 *Study site and plant species*

573

574 The study was conducted in the Serra do Sudeste in the municipalities of Santana da Boa
575 Vista and Caçapava do Sul in the state of Rio Grande do Sul, Brazil, from October to December
576 in 2016 and 2017. The study sites are located in the region of Guaritas near the mining town

577 Minas do Camaquã (30°53'36.3"S 53°25'14.3"W), where steep blocks of sandstone and
578 conglomerate of up to 100 meters in height are common (Fig. 1A). The region is part of the
579 Pampa domain, while the local vegetation is characterized by a grassland matrix with shrubs
580 and small trees and a rich flora of herbaceous species. The local bee fauna is diverse and
581 contains a high proportion of oligolectic species (Schlindwein, 1998). The climate is sub-
582 temperate and humid (Maluf, 2000). The area has an average annual precipitation of 1509 mm
583 and an average annual temperature of 18.7 °C, with frost occurring occasionally in the winter,
584 while summer is hot (INMET, 2018). *Parodia neohorstii* is a local endemic cactus restricted to
585 the region of Guaritas. When the study was undertaken, only two populations were known for
586 the species, with one being chosen for study based on the number and accessibility of
587 individuals. The cacti grow on sun-exposed rock substrate almost without soil with their roots
588 fixed between crevices and small loose stones (Fig. 1B, C). Neighboring vegetation is limited
589 to terrestrial bromeliads (*Dyckia maritima* Baker) and agglomerations of bryophytes. The cacti
590 grow as solitary individuals, and do not branch or form clones. The body of the cactus is
591 globular when young and vertically cylindrical at advanced ages, reaching up to 11 cm height.
592 It possesses 18-26 obtuse ribs and areoles with central straight stiff spines (Anceschi & Magli,
593 2018). At apex of the cactus, long spines and apical lanosity surround the flowers and protect
594 ovaries and fruits, which remain hidden inside the wool. The nectarless flowers have bright
595 yellow petals with a pinkish inner floral tube and produce numerous stamens.

596 Vouchers of the species were registered at BHCB herbarium - Universidade Federal de
597 Minas Gerais, Belo Horizonte, Brazil (BHCB 194347 and 194073).

598

599 *Anthesis and flowering*

600

601 The studied population contained 248 individuals, 93 of which were in reproductive
602 stage. We numbered all reproductive stage individuals (n = 93 plants) with small aluminum
603 tags, and documented flower production over time and the time of flower opening and closure.
604 During 25 days of the same period in 2016 and 2017 (October 27 to November 20), which
605 corresponded to the flowering period of *P. neohorstii*, we recorded the number of open flowers
606 daily to characterize the temporal pattern of flowering. To measure flowering intensity and
607 flowering synchronization among individuals, we counted the number of flowering individuals
608 per day in relation to the total number of reproductive individuals of the population. We related
609 the flowering pattern to the amount of precipitation over time, which was measured daily with

610 a pluviometer. Time of anther opening was determined by directly examining flower buds and
611 flowers.

612

613 *Breeding system*

614

615 To determine the breeding system of *P. neohorstii* we performed controlled pollination
616 experiments with individuals with one open flower per day as follows: (1) hand self-pollination
617 — flowers were hand-pollinated with their own pollen (n = 31); (2) autonomous self-pollination
618 — flowers were bagged without manipulation (n = 33); (3) hand cross-pollination — flowers
619 were cross-pollinated with pollen from other individuals (n = 20); and (4) natural pollination
620 — non-manipulated marked flowers were maintained accessible to flower visitors (n = 31).

621 Flowers used for hand self-pollination, spontaneous self-pollination, and hand cross-
622 pollination were bagged in the bud stage and remained bagged after the treatments until
623 senescence. We determined fruit set by calculating the percentage of formed fruits in relation
624 to the flowers used in each treatment. We determined seed set by calculating the percentage of
625 seeds formed in relation to the average number of ovules per flower, which was determined
626 from 32 flowers of 32 individuals.

627

628 *Floral visitors*

629

630 We recorded the frequency of floral visitors of 30 cactus flowers of different individuals
631 throughout the entire process of anthesis (75 hours of observation) in 2016. We noted the
632 following during 30-min intervals throughout anthesis: (1) bee species; (2) bee sex; (3) landing
633 place on arrival at flower (stamens, petals or stigma); (4) whether or not the bee touched the
634 stigma during the visit; (5) whether or not the bee crossed the bundle of stimulated thigmonastic
635 stamens to access hidden pollen at the base of the flower; and (6) visit duration. The 30-min
636 intervals were used to characterize the frequency of visitation over time.

637 To evaluate pollinator effectiveness, we considered whether the bees touched the
638 stigmas and the frequency of visitors. Bee behavior was also analyzed from video recordings
639 (6 hours) made with a camcorder (Panasonic HC-X929) and digital camera (Canon EOS T5
640 and 5D) mounted on a tripod. Specimens of bees were deposited in the Entomological
641 Collection of UFMG (Centro de Coleções Taxonômicas da UFMG, Universidade Federal de
642 Minas Gerais, Belo Horizonte, Brazil).

643

644 *Impact of the absence of the main pollinator on cactus reproduction*

645

646 At the beginning of the flowering season in 2017, when the main pollinator, *Arhysosage*
647 *cactorum* Moure 1999 (Andrenidae, Calliopsini), was still inactive, we measured fruit and seed
648 set in 33 flowers to assess the impact of this species on the pollination of *P. neohorstii*. These
649 flowers were all accessible to other flower visitors, which were identified through observations
650 of flowers for 10 hours.

651

652 *Pollen fate*

653

654 To characterize pollen fate (and pollen flow promoted by flower visitors), we counted
655 pollen grains inside the flower at different stages of anthesis: (1) “pre-anthesis” — flower buds
656 one hour before opening; (2) “early stage anthesis” — period of high frequency of flower visits,
657 one hour after opening; and (3) “post-anthesis” — recently closed flowers, approximately three
658 hours after opening.

659

660 Bee movements inside flowers deposited pollen grains on various flower parts during
661 visits. Thus, we counted the pollen grains adhering to (a) anthers and petals and (b) the stigmatic
662 surface for the three previously described stages. To quantify pollen grains adhering to the
663 stigma surface, we collected the styles of 10 flowers in each stage of anthesis mentioned above
664 (N = 30 styles total). Each style was transferred to a microscope slide containing glycerine
665 gelatin stained with alcoholic fuchsine solution. The gelatin was melted by heat with an alcohol
666 flame, covered with a coverslip and sealed with paraffin wax. We analyzed the samples under
667 a microscope by identifying and counting all the pollen grains present.

668

669 To measure the quantity of cross-pollen and self-pollen deposited on the stigmatic
670 surface, we emasculated 10 flowers and compared the number of pollen grains adhering to the
671 stigma of emasculated and intact flowers at the end of anthesis. Because pollen is deposited on
672 the stigma after it is touched by bees, we compared the number of stigma touches by floral
673 visitors to emasculated flowers with that of intact flowers (3 hours of observation for pairs of
674 emasculated and intact flowers). The number of conspecific pollen grains adhering to the stigma
675 of emasculated flowers was considered cross-pollen flow because emasculated flowers are only
676 pollen receptors. The difference in mean number of pollen grains on stigmas in emasculated
677 flowers and stigmas in intact flowers was considered self-pollen flow. Differences in mean

676 pollen content of flowers throughout anthesis (pre-anthesis, early stage anthesis, post-anthesis)
677 were considered as the quantity of both pollen collected by flower visiting bees and pollen
678 remaining in the flowers at different time intervals.

679 For flowers and flower buds that had styles removed, we washed petals and stamens
680 with 70% ethanol solution and separated the pollen grains present with a nylon sieve (mesh size
681 70 μm). The solution with the pollen grains was centrifuged for five minutes at 4500 RPM. The
682 suspended solution was withdrawn and 500 μl of 3:1 glycerol lactic acid was added followed
683 by 2 minutes of shaking with a vortex shaker. We extracted an aliquot from this solution for
684 counting pollen grains in a Neubauer chamber (Maêda, 1985).

685

686 *Statistical analyses*

687

688 We performed Student's t-test to compare: (1) the number of seeds produced in natural
689 pollination and hand cross-pollination; (2) number of seeds produced in natural pollination with
690 presence and absence of main pollinator; and (3) number of stigma touches received by
691 emasculated and intact flowers. We used the non-parametric Mann-Whitney Rank Sum Test to
692 compare: (1) the amount of pollen deposited on the stigmatic surface between one hour after
693 flower opening and at the end of anthesis; and (2) the amount of pollen deposited on the
694 stigmatic surface of emasculated and intact flowers. All statistical tests were performed with
695 SigmaStat 3.5 software for Windows.

696

697 **RESULTS**

698

699 *Anthesis and flowering*

700

701 *Parodia neohorstii* blossomed predominantly in November. We identified flowering
702 synchronization among individuals in the population during 2016 and 2017. We identified four
703 flowering peaks each year in the same period, between October 27 and November 20, when up
704 to 61% of the individuals set flowers on the same day (Fig. 2). Flowering seems to have been
705 triggered by rainfall since in both years flowers opened on average 2.6 ± 1.0 (N = 11) days after
706 rain, but only on days with high solar intensity (Fig. 2). Flowering, thus, was not continuous
707 and concentrated to just a few days; there were no cactus flowers present during 56% of the
708 flowering period (N = 25 days) in 2016 and 39% (N = 41 days) in 2017.

709 Flowers opened synchronously in the population at around 12:44h and closed 3 hours
 710 later (188 ± 18 min; $N = 41$). There was little variation in the time of flower opening within the
 711 population (on average 30 ± 13 minutes) ($N = 7$ days). Under natural conditions, all flowers
 712 accessible to pollinators opened for only one day ($N = 136$). Bagged flowers without pollen
 713 deposition on the stigmas opened for two (55.4%) or three days (44.6%) ($N = 56$). Flowering
 714 individuals had on average 1.5 ± 0.8 open flowers per day; 65.8% had one, 25.1% two, 6.1%
 715 three, and 3.0% four to seven simultaneously open flowers per day ($N = 700$). Individual plants
 716 produced from 1 to 26 flowers (5.3 ± 4.3 , $N=332$) per year.

717 When petals unfolded, all anthers were already dehisced and the stigma receptive.
 718 Hermaphrodite flowers had on average 206 ± 20 ($N = 10$) sensitive stamens inserted in 10 or
 719 11 layers ($N = 20$) along the floral tube. When floral visitors contacted the stamens, the stamens
 720 responded to the mechanic stimulus by bending inward until they attached to the style. Thus,
 721 the layer of the uppermost stamens formed by a set of 53 ± 4 stamens ($N = 20$), covered the rest
 722 of the stamens that were inserted in the lower part of the floral tube. In the stimulated condition,
 723 only 25% of the total pollen content of the flower was freely accessible to floral visitors.

724

725 *Breeding system*

726

727 The studied population of *P. neohorstii* was predominantly xenogamous. All marked
 728 non-bagged flowers accessible to pollinators, as well as all hand cross-pollinated flowers, had
 729 similar fruit and seed set ($t = -0.513$, d.f. = 49, $P = 0.610$, $N = 51$, Table 1). Fruit set was low
 730 for self-pollination treatments, (32% after hand self-pollination and 10% after autonomous self-
 731 pollination), while seed set was minute (2 seeds per fruit on average in both treatments) (Table
 732 1).

733

734 *Floral visitors*

735

736 We recorded a total of 2284 bee visits to flowers of *P. neohorstii* representing seven
 737 species of four families, all of which were solitary species (Table 2). Only two species were
 738 frequent flower visitors: *Arhysosage cactorum* with 81.2% of the visits, and *Anthrenoides*
 739 *micans* Urban 1955 (Andrenidae, Protandrenini) with 16.9% (Fig 3, Video 1). Other bee species
 740 were sporadic flower visitors and together accounted for only 1.9% of the visits (Table 2).

741 Bees landed on flowers at three sites: stigmatic surface, petals and stamens. Females of
742 *A. cactorum* used the stigmatic surface as their main landing platform while *A. micans* landed
743 mainly on petals (Table 3). The flowers were also searched by male bees of both species, who
744 mainly used the stigmatic surface to land. The bees synchronized their flight activity with
745 flower opening and actively collected pollen from the first minutes opening onward (Fig. 4A).
746 Females of *A. cactorum* forced their entry when the petals unfolded before the stamens were
747 visible. Males of both bee species patrolled flowers in search of females, and flowers served as
748 mating sites. Males of *A. cactorum* occasionally hid between the stamens waiting for females.
749 When females arrived in flowers in search for pollen, males seized them with their mandibles
750 and tried to copulate. Couples in copulatory position flew between flowers while the females
751 continued foraging for pollen even *in copula*, which resulted in high frequencies of pollen
752 transfer and stigma touches (Fig. 3A). Couples contacted stigmas repeatedly during almost all
753 flower visits (Table 3). Females of *A. cactorum* touched stigmas more frequently than females
754 of *A. micans*. Visits of males of *A. micans* were rare because they frequently flew over the
755 flowers without touching them; of 13 flyovers the flower was touched only once (N = 413), and
756 most of these contacts were to the stigma surface (Table 3). Couples of *A. micans* frequently
757 touched the stigma, but they were very rare in the flowers (0.4% of the visits).

758 The sensitive stamens moved to the center of the flower after mechanic stimuli such as
759 body contacts of bees during pollen collection. The dense layer of the stimulated uppermost
760 stamens hindered the bee access to the pollen of the lower stamens, thus forming a pollen
761 chamber containing 75% of the pollen grains of a flower. Females of *A. cactorum* crossed the
762 stamen barrier in stimulated flowers during 94% of the visits (N = 83) and continued to collect
763 the hidden pollen, while females of *A. micans* reached the pollen chamber during only 39% (N
764 = 98) of the visits to flowers with stimulated stamens. Larger bees of *Cephalocolletes rugata*
765 Urban 1995 and *Ancyloscelis romeroi* Holmberg 1903, which visited the flowers only
766 sporadically, accessed the hidden pollen on all of their recorded visits. Small bees, like those of
767 *Dialictus* sp., never crossed the bundle of stamens.

768 Fifty-two percent of all visits occurred during the first hour of anthesis. The frequency
769 of female bees decreased drastically thereafter, and about two hours after flower opening
770 females were practically absent. Males of *A. cactorum*, on the other hand, continued patrolling
771 flowers in search of females even when female flower visits had ceased (Fig. 4B). Visits were
772 very infrequent during the last 30 minutes of anthesis and represented only 1.4% of the total.

773

774 *Impact of the absence of the main pollinator on cactus reproduction*

775

776 In the beginning of the flowering season in 2017, the most frequent visitor, *A. cactorum*,
 777 was completely absent for 13 days. During this period only 33% of the flowers set fruits (N =
 778 33). Seed set differed dramatically between the presence and absence of this main pollinator (t
 779 = -8.199, d.f = 40, P = <0.001, N = 42) with only 18% of ovules setting seed in its absence. In
 780 addition, flower longevity increased to 2.25 ± 0.44 days (N = 16). The flowers were visited by
 781 bees of *A. micans* (72.1%), *Dialictus* sp. (24.5%), *C. rugata* (1.7%), and *Callonychium* sp.
 782 (1.7%) (N = 61 visits).

783

784 *Pollen fate*

785

786 Flowers of *P. neohorstii* produced on average $100,200 \pm 13,614$ (N = 10) pollen grains.
 787 Bees collected 58% of the total pollen from the flowers during the first hour of anthesis (Fig.
 788 5A), after which the stigmatic surface was already covered with pollen grains, with an average
 789 of $1,721 \pm 695$ grains (N = 10) adhered to the stigmas. There was no difference in the number
 790 of pollen grains adhering to the stigma in the first hour and at the end of anthesis (U = 55,000,
 791 P = 0.734, N = 20, Fig. 5B, Fig. 6). Heteroespecific pollen grains (i.e., from other species)
 792 represented 0.34% of the total pollen deposition on the stigma (5.9 ± 6.9 ; N = 10). This pollen
 793 content came from Asteraceae, Poaceae, Iridaceae and Euphorbiaceae (*Croton* sp.).
 794 Autonomous pollen deposition (i.e., by the flower on its own stigma) was minimal. Only $106 \pm$
 795 97 (N=10) grains were deposited on stigmas of fresh open flowers.

796 Emasculated and intact flowers received a similar number of stigma touches by bees (t
 797 = -1.294, d.f. = 34, P = 0.204, N = 36), which promoted the same chance of pollen deposition
 798 on the stigma surface in both treatments. Emasculated flowers received on average 2.2 ± 1.2
 799 stigma touches per 10-minute interval, while intact flowers received on average 1.8 ± 0.7 (N =
 800 18 pairs). The number of pollen grains deposited on the stigmatic surface of the two treatments,
 801 however, differed (U = 84.000, P = 0.011, N = 20), with emasculated flowers having on average
 802 929 ± 577 (N = 10) pollen grains on the stigmatic surface while intact flowers had on average
 803 $1,730 \pm 747$ (N = 10). The difference between treatments was considered autogamous pollen
 804 deposition during the movement of bees inside the flower (801 ± 314 grains, N = 10).

805 Only 1.7% of the pollen produced by a flower was deposited on a stigmatic surface by
 806 the end of anthesis (Fig. 7). Thus, bees collected 86.5% of the total pollen produced by flowers.

807 At flower closure, 11.8% of the total pollen content ($11,890 \pm 3.813$ grains, $N = 10$) remained
808 adhering to floral parts inside the flower.

809

810 **DISCUSSION**

811

812 *Pollination of Parodia neohorstii*

813

814 *Parodia neohorstii* depends on specialized pollinators to reproduce and only sets fruit
815 and seeds after cross-pollination. The oligolectic bee *Arhysosage cactorum* is by far its most
816 important pollinator due to the following attributes: (1) high rate of stigma touches; (2) effective
817 pollen transfer among individuals; (3) frequent floral visits; (5) high flower constancy during
818 foraging flights; and (6) high fruit and seed set only in its presence.

819 The use of the stigmatic surface as a landing platform dramatically increases cross-
820 pollen transfer because xenogamous pollen adhering to a bee's body from previously visits is
821 deposited directly on the stigma before the bee makes contact with autogamous pollen. When
822 floral visitors land on petals or stamens, and directly collect pollen before touching the stigma,
823 autogamous pollen grains are deposited on their body, adhering to the superficial layers and
824 covering xenogamous pollen. According to layering models (Harder & Wilson, 1996), such
825 covering decreases the chance of xenogamous pollen deposition on the stigma because it is
826 located under layers of autogamous pollen on the bee's body. Other cactus-oligolectic bees have
827 also been reported to land on stigmas (Schlindwein & Wittmann, 1997; Blair & Williamson,
828 2008; Lenzi & Orth, 2011).

829 Besides *A. cactorum* being the most frequent floral visitor, females of the species have
830 additional stigma touches during movements within the flowers and touch the stigmatic surface
831 twice as much as females of *A. micans* and four times more than females of *Dialictus* sp.
832 Moreover, also males of *A. cactorum* most likely contribute to efficient cross pollen flow
833 because while searching for females, they inspect each flower of *P. neohorstii* on the routes of
834 their patrolling flights, frequently use the stigma as landing platform and touch anthers. In the
835 absence of *A. cactorum*, other bee species promote only very low fruit set, and especially low
836 seed set, probably due to low xenogamous pollen flow. As is the case for *P. neohorstii*, other
837 self-incompatible cacti species have been found to be dependent on specialized bee pollinators,
838 such as *Astrophytum asterias* Lem., *Ferocactus cylindraceus* (Engelm.) Orcutt and *Ferocactus*
839 *wislizeni* Britton & Rose, even when numerous other floral visitors are present (McIntosh, 2005;

840 Blair & Williamson, 2008). In these cases, the focal cacti only set seed in the presence of
 841 oligolectic bee pollinators.

842 We consider our findings to indicate that the interaction between *A. cactorum* and *P.*
 843 *neohorstii*, as is likely true with interactions of other melittophilous *Parodia* cacti, corresponds
 844 to an “ecological specialization”, *sensu* Armbruster (2017). Thus, we corroborate the idea that
 845 analyzing flower morphology alone, decoupled from analyses of pollinator feeding behavior,
 846 is not sufficient to determine the degree of “realized specialization” of flower-pollinator
 847 interactions.

848

849 *Controlled pollen removal enhances male success*

850

851 Synchronized mass flowering with pollen presentation by a population is an
 852 extraordinary attractive to pollinators. Oligolectic bees time their visits to flower opening
 853 already when petals start to unfold and concentrate their pollen collection during the first hour
 854 of anthesis. *Parodia* floral traits, such as wide-open corollas and pollen-at-once presentation,
 855 promote this fast pollen removal, but otherwise decrease cross-pollen flow (Harder & Wilson,
 856 1996). In this context, we interpret pollen partitioning mediated by thigmonastic stamen
 857 movements as a mechanism to limit pollen removal by individual bees that do not have adequate
 858 flower handling ability. Stimulated flowers hide about three quarters of their total pollen, thus
 859 restricting pollen access for generalist bees to the upper layers of stamens. This floral
 860 functioning improves male function because unspecialized floral visitors remove less pollen
 861 per visit, and thus pollen availability is maintained for more time, attractiveness to pollen
 862 seeking females is extended and movements of bees among flowers is enhanced, all of which
 863 contribute to increasing cross-pollen flow. Geitonogamy is largely avoided since that the
 864 majority of individual cacti open only one flower per day.

865

866 *Specialization of Arhysosage cactorum on cacti flowers*

867

868 Besides the high frequency of visits and stigma touches, some behavioral characteristics
 869 of *A. cactorum* make it a specialized pollinator of *P. neohorstii*.

870 (i) Pollen diet — females of this species restrict the diet of their larvae to only cacti
 871 pollen. As shown by scopa pollen load analyses, females always carry monospecific pollen
 872 content of the cactus species in flower during a foraging trip (Schlindwein & Wittmann, 1995).

873 (ii) Timing foraging activity with flower opening — females of *A. cactorum* are capable
 874 of adjusting their pollen foraging to the exact moment of flower opening. This finding is
 875 surprising because whole populations of *P. neohorstii* have synchronized, but episodic,
 876 flowering during only five to six days per month. The fine-tuned synchronization of foraging
 877 on a sporadic flowering cacti species maybe related to (a) perception of specific chemical and/or
 878 visual cues that attract the specialist bees to the flowers (Burger *et al.*, 2010; Dobson *et al.*,
 879 2012; Milet-Pinheiro *et al.*, 2012; Carvalho *et al.*, 2014) ; (b) recognition of changes in weather
 880 conditions that induce flowering (changes in air humidity, rainfall regime, temperature,
 881 sunshine intensity); or (c) monitoring and inspection of cactus sites.

882 (iii) Behavioral adaptation, flower and pollen handling — females of *A. cactorum* have
 883 the most efficient pollen collection behavior among the flower visiting bee species in *P.*
 884 *neohorstii*. During the first hour of anthesis, period with high rates of bee visitation, the position
 885 of stimulated stamens makes access to pollen difficult for floral visitors, except for females of
 886 *A. cactorum*, which are the only bees with the capacity to always cross the bundle of stamens
 887 to access the hidden pollen. Thus, the formation of this pollen chamber can also be interpreted
 888 as a mechanical filter against generalized bees by restricting access to the major pollen reservoir
 889 to only bees with flower handling “know-how”. The same functional interpretation of
 890 thigmonastic stamen movements in cacti flowers was previously proposed for flowers of
 891 *Opuntia* cacti, where stimulated stamens hide about 80% of the pollen grains inside the pollen
 892 chamber and also where only oligolectic specialist bees gained access to this pollen
 893 (Schlindwein & Wittmann, 1997).

894 (iv) Mating behavior of *A. cactorum* — similar to many other oligolectic bee species,
 895 the reproductive strategy of male *A. cactorum* is associated with the pollen host plants of
 896 females (Wittmann *et al.*, 1990; Medeiros & Schlindwein, 2003; Oliveira & Schlindwein, 2010;
 897 Oliveira *et al.*, 2012; Oliveira, 2015). Males of *A. cactorum* wait and search for females in
 898 cactus flowers, while couples flying *in copula* among cacti flowers strongly enhance cross
 899 pollination (Schlindwein & Wittmann, 1995). This mating behavior is only known for a few
 900 other species of Andrenidae (Alcock *et al.*, 1978; Wittmann *et al.*, 1990).

901

902 *Pollen fate*

903

904 Pollen flow is complex with many factors influencing pollen destination (Harder &
 905 Wilson, 1996). We estimated the main pollen fates here by integrating extensive field

906 observation of pollinator behavior and flower functioning. Our analysis of pollen destination
907 for the studied population of *P. neohorstii* revealed that less than 2% of the pollen grains
908 produced in a flower reach stigmas, and about half of this comprises xenogamous pollen.
909 Although only a small portion of pollen is destined directly for cacti reproduction, the mean
910 amount of cross-pollen deposited on the stigmas is about 12-times greater than the number of
911 ovules, and thus sufficient to achieve almost optimal seed output. Thus, fruit and seed set were
912 maximal for the cacti-bee association, but not when bees of *A. cactorum* were absent. Bees of
913 the genus *Arhysosage* are closely related with cacti species in different areas of South America
914 (Schlindwein & Wittman, 1995; Moure, 1999; Engel, 2000; Ramos, 2013). These bees forage
915 for pollen only in Cactaceae and exhibit reproductive behavior associated with cactus flowers
916 (Schlindwein & Wittman, 1995; Ramos, 2013), indicating that the evolutionary history of the
917 genus *Arhysosage* might be tightly related to melittophilous cacti species. High frequency of
918 flower visitation rates of these bees in *P. neohorstii* promote high reproductive success of the
919 female function. The male function, however, gets uncertain due the huge amount of pollen
920 removed from plant reproduction. Because of the variation in the destiny of pollen, it can be
921 expected that selection pressures act more deeply in the male than in the female component of
922 fitness (Moore & Pannell, 2011). Based on our data, however, it is not possible to determine to
923 what extent this apparently exaggerated sexual performance of male function is consequence
924 of sexual selection via an intra-sexual competition mechanism to improve male reproductive
925 success, or a result of natural selection via consumption of pollen grains by specialized
926 pollinators.

927 Although we cannot quantify exactly the amount of pollen collected by females of the
928 two frequent visitor species *A. cactorum* and *A. micans*, it is clear that a larger portion by far
929 flows into the reproduction of the oligolectic *A. cactorum*. This greater flow is due the following
930 attributes: (1) visiting *A. cactorum* females are five times more frequent, and their visits three
931 times longer, than females of *A. micans*; (2) body and scopa size of *A. cactorum* are also
932 somewhat greater, which likely results in a greater capacity to carry pollen.

933 Similar large amounts of pollen (86-95%) were found to be collected by oligolectic
934 pollinators to feed their larvae in other specialized associations between oligolectic bees and
935 plants (Schlindwein *et al.*, 2005; Carvalho & Schlindwein, 2011; Pick & Schlindwein, 2011).
936 Although huge quantities of pollen flow to oligolectic bee nests, plants that maintain
937 relationships with such bees have great chances to achieve high reproductive success due to
938 high degree of conspecific pollen transfer among individuals (Schlindwein *et al.*, 2005). Thus,

939 this huge quantity of pollen that goes to oligolectic bee nests is the "fidelity cost" of attracting
 940 pollen-specialist bees as main vectors of effective pollination service.

941 Another 11.8% can be considered pollen loss due removal failure, because uncollected
 942 pollen inside of flowers does not serve pollination nor feed future pollinators.

943 Analysis of stigmatic pollen loads in *P. neohorstii* revealed almost pure compositions of
 944 conspecific pollen and, thus, an efficient flower-pollinator association. Heterospecific pollen
 945 grains were negligible, and probably were carried by females of *A. micans*, because some of
 946 them arrived to the flowers carrying pollen loads of different color (Fig. 3C). Furthermore, this
 947 species has been documented to carry mixed scopa pollen loads when visiting cacti flowers
 948 (Schlindwein & Wittmann, 1995).

949

950 *Interdependence of cacti and oligolectic bees: a relationship in danger?*

951

952 The interaction between *P. neohorstii* and *A. cactorum* is highly seasonal and occurs
 953 only during a few weeks of the year. The flight periods of oligolectic bees usually encompass
 954 the blooming period of their host plants (Wcislo & Cane, 1996). A mismatch between the
 955 period of emergence of *Arhysosage* bees from nests and flowering of *P. neohorstii* could
 956 substantially impact the reproductive success of both the plant (without effective cross-pollen
 957 transfer) and the pollinator (without rendezvous and mating sites).

958 We noted that other cacti species from the genera *Parodia*, *Gymnocalycium* and *Frailea*
 959 that occur at the study site, and which are also intensely visited by the oligolectic cacti bees
 960 (Schlindwein & Wittmann, 1995), blossomed during the breaks in flowering of *P. neohorstii*
 961 (unpubl. data). This might guarantee continuous pollen supply to *A. cactorum*. Such staggered
 962 flowering among alternate days has been reported previously for other melittophilous sympatric
 963 cacti species (Giorgis *et al.*, 2015; Eggli & Giorgetta, 2017). This pattern is an effective strategy
 964 to share pollinators and maintain reproductive isolation from similar species with overlapping
 965 populations (Giorgis *et al.*, 2015). Our data indicate that the synchronized mass flowering of *P.*
 966 *neohorstii* was triggered mainly by rainfall, which is similar to the findings of Eggli & Giorgetta
 967 (2017). It is possible that other abiotic factors, such as temperature, air humidity and direct
 968 sunlight, also influence synchronized flower opening of cacti populations and contribute to
 969 alternating flowering of sympatric cacti species.

970

971 *Arhysosage cactorum*, therefore, is the key pollinator of the community of small cacti
 in the study region. The co-occurrence of a range of cacti species that bloom, in a staggered

972 manner, during the flight season of *A. cactorum* may be necessary to maintain this bee species
973 throughout the entire flowering season, as observed for cacti oligoleges in Arizona and Texas
974 (Simpson & Neff, 1987; Blair & Williamson, 2008).

975 Our study shows that the specialized relationship between *P. neohorstii* and *A. cactorum*
976 is effective but also fragile. Both partners are threatened, red listed species (Larocca &
977 Machado, 2013; Sul, 2014). It is improbable that pollinator limitation puts the cactus in danger,
978 but there are other factors that could disrupt the interaction between *P. neohorstii* and *A.*
979 *cactorum*. Any environmental impact, such as climatic change, that causes phenological
980 variation in the periods of plant flowering and pollinator activity (Memmott *et al.*, 2007), may
981 promote a mismatch between partners and lead to reproductive failure. Changes in land use,
982 driven by agriculture and livestock farming, affect cactus habitats, availability of nesting sites
983 for pollinators, and the conservation of the threatened partner species.

984

985 **ACKNOWLEDGEMENTS**

986

987 We thank Vinícius Brito, Cristiane Martins and Eduardo Borba for suggestions on a
988 previous version of the manuscript. João Larocca, Rosana Singer, Josy Zarur, Andréia Carneiro,
989 Ricardo Aranha, Leandro dal Ri, Ari Delmo Nilsson, Rubem Avila and Luis Melo for
990 multidisciplinary cooperation-work with the threatened cacti of Rio Grande do Sul; Marlise
991 Freitas, Vânia Pessoa, and Família Peohren for logistic support. We thank Stephen Buchmann,
992 Rodrigo Medel and an anonymous reviewer for their helpful comments that improved the final
993 version of the manuscript. We acknowledge financial support by Foundation Grupo O Boticário
994 de Proteção à Natureza (project number 1095_20171), PPGBV, CAPES and CPNq in the form
995 of research grants to CS, IC, SS-O, RO and AP. IBAMA granted the license (number 57684-1)
996 to study the red list species.

997

998 REFERENCES

999

- 1000 **Alcock J, Barrows EM, Gordh G, Hubbard LJ, Kirkendall L, Pyle DW, Ponder TL,**
 1001 **Zalom FG.** 1978. The ecology and evolution of male reproductive behaviour in the bees and
 1002 wasps. *Zoological Journal of the Linnean Society* **64**: 293-326.
- 1003 **Anceschi G, Magli A.** 2018. A synopsis of the genus *Parodia* Spegazzini s.l. (Cactaceae).
 1004 *Bradleya* **1**: 70-161.
- 1005 **Armbruster WS.** 2017. The specialization continuum in pollination systems: diversity of
 1006 concepts and implications for ecology, evolution and conservation. *Functional Ecology* **31**:
 1007 88-100.
- 1008 **Blair AW, Williamson PS.** 2008. Effectiveness and Importance of Pollinators to the Star
 1009 Cactus (*Astrophytum asterias*). *The Southwestern Naturalist* **53**: 423-430.
- 1010 **Bond WJ.** 1994. Do mutualisms matter? Assessing the impact of pollinator and disperser
 1011 disruption on plant extinction. *Philosophical Transactions of the Royal Society B* **344**: 83-
 1012 90.
- 1013 **Boyle T.** 1997. The genetics of self-incompatibility in the genus *Schlumbergera* (Cactaceae).
 1014 *Journal of Heredity* **88**: 209-214.
- 1015 **Burger H, Dötterl S, Ayasse M.** 2010. Host-plant finding and recognition by visual and
 1016 olfactory floral cues in an oligolectic bee. *Functional Ecology* **24**: 1234-1240.
- 1017 **Carvalho AT, Dötterl S, Schlindwein C.** 2014. An aromatic volatile attracts oligolectic bee
 1018 pollinators in an interdependent bee-plant relationship. *Journal of Chemical Ecology* **40**:
 1019 1126-1134.
- 1020 **Carvalho AT, Schlindwein C.** 2011. Obligate association of an oligolectic bee and a seasonal
 1021 aquatic herb in semi-arid north-eastern Brazil. *Biological Journal of the Linnean Society*
 1022 **102**: 355-368.
- 1023 **Dobson HEM, Ayasse M, O'Neal KA, Jacka JA.** 2012. Is flower selection influenced by
 1024 chemical imprinting to larval food provisions in the generalist bee *Osmia bicornis*
 1025 (Megachilidae)? *Apidologie* **43**: 698-714.
- 1026 **Eggl U, Giorgetta M.** 2017. Flowering phenology and observations on the pollination biology
 1027 of South American cacti. 3. Temporally robust sequential “big bang” flowering of two
 1028 unrelated sympatric globular cacti in northern Argentina. *Haseltonia* **23**: 97-109.

- 1029 **Giorgis MA, Cingolani AM, Gurvich DE, Astegiano J.** 2015. Flowering phenology, fruit set
1030 and seed mass and number of five coexisting *Gymnocalycium* (Cactaceae) species from
1031 Córdoba mountain, Argentina. *The Journal of the Torrey Botanical Society* **142**: 220-230.
- 1032 **Goettsch B, Duran AP, Gaston KJ.** 2018. Global gap analysis of cactus species and priority
1033 sites for their conservation. *Conservation Biology* **33**: 369-376
- 1034 **Goettsch B, Hilton-Taylor C, Cruz-Pinon G, Duffy JP, Frances A, Hernandez HM, Inger
1035 R, Pollock C, Schipper J, Superina M, Taylor NP, Tognelli M, Abba AM, Arias S,
1036 Arreola-Nava HJ, Baker MA, Barcenas RT, Barrios D, Braun P, Butterworth CA,
1037 Burquez A, Caceres F, Chazaro-Basanez M, Corral-Diaz R, Del Valle Perea M, Demaio
1038 PH, Duarte de Barros WA, Duran R, Yancas LF, Felger RS, Fitz-Maurice B, Fitz-
1039 Maurice WA, Gann G, Gomez-Hinostrosa C, Gonzales-Torres LR, Patrick Griffith M,
1040 Guerrero PC, Hammel B, Heil KD, Hernandez-Oria JG, Hoffmann M, Ishihara MI,
1041 Kiesling R, Larocca J, Leon-de la Luz JL, Loaiza SC, Lowry M, Machado MC, Majure
1042 LC, Avalos JG, Martorell C, Maschinski J, Mendez E, Mittermeier RA, Nassar JM,
1043 Negrón-Ortiz V, Oakley LJ, Ortega-Baes P, Ferreira AB, Pinkava DJ, Porter JM,
1044 Puente-Martinez R, Gamarra JR, Perez PS, Martinez ES, Smith M, Manuel
1045 Sotomayor MDCJ, Stuart SN, Muñoz JL, Terrazas T, Terry M, Trevisson M, Valverde
1046 T, Van Devender TR, Veliz-Perez ME, Walter HE, Wyatt SA, Zappi D, Alejandro
1047 Zavala-Hurtado J, Gaston KJ.** 2015. High proportion of cactus species threatened with
1048 extinction. *Nature Plants* **1**: 15142.
- 1049 **Grant V, Hurd P.** 1979. Pollination of the southwestern Opuntias. *Plant Systematics and
1050 Evolution* **133**: 15-28.
- 1051 **Harder L, Wilson W.** 1996. Theoretical perspectives on pollination. *Acta Horticulturae*. **437**:
1052 83-102
- 1053 **INMET.** 2018. Banco de Dados Meteorológicos para Ensino e Pesquisa - BDMEP. Dados
1054 históricos da estação meteorológica 83980 - Bagé - RS entre 1988 e 2018.
1055 (<http://www.inmet.gov.br/portal/index.php?r=bdmep/bdmep>) Accessed 27/09/2018.
- 1056 **Inouye DW, Gill DE, Dudash MR, Fenster CB.** 1994. A model and lexicon for pollen fate.
1057 *American Journal of Botany* **81**: 1517-1530.
- 1058 **Johnson SD, Neal PR, Harder LD.** 2005. Pollen fates and the limits on male reproductive
1059 success in an orchid population. *Biological Journal of the Linnean Society* **86**: 175-190.

- 1060 **Larocca J, Machado M.** 2013. *Parodia neohorstii*. *The IUCN Red List of Threatened Species*
1061 2013 (<http://dx.doi.org/10.2305/IUCN.UK.2013-1.RLTS.T152718A669833.en>.) Accessed
1062 27/09/2018.
- 1063 **Lenzi M, Orth AI.** 2011. Visitantes florais de *Opuntia monacantha* (Cactaceae) em restingas
1064 de Florianópolis, SC, Brasil. *Acta Biológica Paranaense* **40**: 19-32.
- 1065 **Maêda J.** 1985. Manual para uso da câmara de Neubauer para contagem de pólen em espécies
1066 florestais. Universidade Federal do Rio de Janeiro, Rio de Janeiro. Departamento de
1067 Silvicultura. Seropédica.
- 1068 **Maluf JR.** 2000. Nova classificação climática do Estado do Rio Grande do Sul. *Revista*
1069 *Brasileira de Agrometeorologia* **8**: 141-150.
- 1070 **Mandujano MdC, Carrillo-Angeles I, Martínez-Peralta C, Golubov J.** 2010. Reproductive
1071 biology of Cactaceae. pp. 197-230. In: **Ramawat KG.** Eds *Desert plants: biology and*
1072 *biotechnology*. Berlin:Heidelberg, Springer-Verlag. 503pp.
- 1073 **Martinez-Peralta C, Marquez-Guzman J, Mandujano MC.** 2014. How common is self-
1074 incompatibility across species of the herkogamous genus *Ariocarpus*? *American Journal of*
1075 *Botany* **101**: 530-538.
- 1076 **Martins C, Freitas L.** 2018. Functional specialization and phenotypic generalization in the
1077 pollination system of an epiphytic cactus. *Acta Botanica Brasilica* **32**: 359-366.
- 1078 **McFarland JD, Kevan PG, Lane MA.** 1989. Pollination biology of *Opuntia imbricata*
1079 (Cactaceae) in southern Colorado. *Canadian Journal of Botany* **67**: 24-28.
- 1080 **McIntosh ME.** 2002. Flowering phenology and reproductive output in two sister species of
1081 *Ferocactus* (Cactaceae). *Plant Ecology* **159**: 1-13.
- 1082 **McIntosh ME.** 2005. Pollination of two species of *Ferocactus*: interactions between cactus-
1083 specialist bees and their host plants. *Functional Ecology* **19**: 727-734.
- 1084 **Medeiros PCR, Schlindwein C.** 2003. Territórios de machos, acasalamento, distribuição e
1085 relação com plantas em *Protomeliturga turnerae* (Ducke, 1907) (Hymenoptera,
1086 Andrenidae). *Revista Brasileira de Entomologia* **47**: 589-596.
- 1087 **Memmott J, Craze PG, Waser NM, Price MV.** 2007. Global warming and the disruption of
1088 plant-pollinator interactions. *Ecology Letters* **10**: 710-717.
- 1089 **Milet-Pinheiro P, Ayasse M, Schlindwein C, Dobson HEM, Dötterl S.** 2012. Host location
1090 by visual and olfactory floral cues in an oligolectic bee: innate and learned behavior.
1091 *Behavioral Ecology* **23**: 531-538.

- 1092 **Milet-Pinheiro P, Schlindwein C.** 2010. Mutual reproductive dependence of distylic *Cordia*
 1093 *leucocephala* (Cordiaceae) and oligolectic *Ceblurgus longipalpis* (Halictidae, Rophitinae)
 1094 in the Caatinga. *Annals of Botany* **106**: 17-27.
- 1095 **Moore JC, Pannell JR.** 2011. Sexual selection in plants. *Current Biology* **21**: 176-182.
- 1096 **Müller A, Diener S, Schnyder S, Stutz K, Sedivy C, Dorn S.** 2006. Quantitative pollen
 1097 requirements of solitary bees: implications for bee conservation and the evolution of bee-
 1098 flower relationships. *Biological Conservation* **130**: 604-615.
- 1099 **Oliveira R.** 2015. Estratégias de acasalamento em abelhas solitárias no Brasil: o que os machos
 1100 fazer para se acasalar? pp. 439-457. In: **Aguiar AJC, Gonçalves RB, Ramos KS** eds.
 1101 *Ensaio sobre as abelhas da Região Neotropical*. Curitiba: Editora UFPR. 456 pp.
- 1102 **Oliveira R, Carvalho AT, Schlindwein C.** 2012. Territorial or wandering: how males of
 1103 *Protodiscelis palpalis* (Colletidae, Paracolletinae) behave in searching for mates. *Apidologie*
 1104 **43**: 674-684.
- 1105 **Oliveira R, Schlindwein C.** 2010. Experimental demonstration of alternative mating tactics of
 1106 male *Ptilothrix fructifera* (Hymenoptera, Apidae). *Animal Behaviour* **80**: 241-247.
- 1107 **Ollerton J, Killick A, Lamborn E, Watts S, Whiston M.** 2007. Multiple meanings and
 1108 modes: on the many ways to be a generalist flower. *Taxon* **56**: 717-728.
- 1109 **Osborn MM, Kevan PG, Lane MA.** 1988. Pollination biology of *Opuntia polyacantha* and
 1110 *Opuntia phaeacantha* (Cactaceae) in southern Colorado. *Plant Systematics and Evolution*
 1111 **159**: 85-94.
- 1112 **Pick RA, Schlindwein C.** 2011. Pollen partitioning of three species of Convolvulaceae among
 1113 oligolectic bees in the Caatinga of Brazil. *Plant Systematics and Evolution* **293**: 147-159.
- 1114 **Robertson, C.** 1925. Heterotropic Bees. *Ecology* **6**: 412-436.
- 1115 **Roig-Alsina A, Schlumpberger BO.** 2008. The cactus-specialist bees of the genus
 1116 *Brachyglossula* Hedicke (Hymenoptera: Colletidae): Notes on host associations and
 1117 description of a new species. *Journal of the Kansas Entomological Society* **81**: 84-91.
- 1118 **Schlindwein C.** 1998. Frequent Oligolecty Characterizing a Diverse Bee-Plant Community in
 1119 a Xerophytic Bushland of Subtropical Brazil. *Studies on Neotropical Fauna and*
 1120 *Environment* **33**: 46-59.
- 1121 **Schlindwein C, Martins C.** 2000. Competition between the oligolectic bee *Ptilothrix plumata*
 1122 (Anthophoridae) and the flower closing beetle *Pristimerus calcaratus* (Curculionidae) for
 1123 floral resources of *Pavonia cancellata* (Malvaceae). *Plant Systematics and Evolution* **224**:
 1124 183-194.

- 1125 **Schlindwein C, Wittmann D.** 1995. Specialized solitary bees as effective pollinators of South
1126 Brazilian species of *Notocactus* and *Gymnocalycium* (Cactaceae). *Bradleya* **13**: 25-34.
- 1127 **Schlindwein C, Wittmann D.** 1997. Stamen movements in flowers of *Opuntia* (Cactaceae)
1128 favour oligolectic pollinators. *Plant Systematics and Evolution* **204**: 179-193.
- 1129 **Schlindwein C, Wittmann D, Martins CF, Hamm A, Siqueira JA, Schiffler, D, Machado**
1130 **IC.** 2005. Pollination of *Campanula rapunculoides* L. (Campanulaceae): How much pollen
1131 flows into pollination and into reproduction of oligolectic pollinators? *Plant Systematics and*
1132 *Evolution* **250**: 147-156.
- 1133 **Simpson BB, Neff JL.** 1987. Pollination Ecology in the Southwest. *Journal of Systematic and*
1134 *Evolutionary Botany* **11**: 417-440.
- 1135 **Siriani-Oliveira S, Oliveira R, Schlindwein C.** 2018. Pollination of *Blumenbachia amana*
1136 (Loasaceae): flower morphology and partitioned pollen presentation guarantee a private
1137 reward to a specialist pollinator. *Biological Journal of the Linnean Society* **124**: 479-491.
- 1138 **Strong AW, Williamson PS.** 2007. Breeding System of *Astrophytum Asterias*: An Endangered
1139 Cactus. *The Southwestern Naturalist* **52**: 341-346.
- 1140 **Sul ALRGS.** 2014. Decreto N.º 51.797, de 8 de Setembro de 2014. *Espécies da Fauna Silvestre*
1141 *Ameaçadas de Extinção no Estado do Rio Grande do Sul*. Rio Grande do Sul, Brasil.
- 1142 **Weislo WT, Cane JH.** 1996. Floral resource utilization by solitary bees (Hymenoptera:
1143 Apoidea) and exploitation of their stored foods by natural enemies. *Annual review of*
1144 *entomology* **41**: 257-286.
- 1145 **Wittmann D, Radtke R, Cure J, Schifino-Wittmann M.** 1990. Coevolved reproductive
1146 strategies in the oligolectic bee *Callonychium petuniae* (Apoidea, Andrenidae) and three
1147 purple flowered *Petunia* species (Solanaceae) in southern Brazil. *Zeitschrift für zoologische*
1148 *Systematik und Evolutionsforschung* **28**: 157-165.
- 1149

1150 **TABLES**

1151

1152 **Table 1.** Measures of fruit and seed set for pollination treatments of *Parodia neohorstii*: (1) autonomous
 1153 self-pollination; (2) hand self-pollination; (3) hand cross-pollination; (4) natural pollination; and (5)
 1154 *Arhysosage cactorum* absence — non-manipulated marked flowers visited by *Anthrenoides micans*,
 1155 *Dialictus* sp., *Cephalocolletes rugata* and *Callonychium* sp. during the 13 day period of absence of *A.*
 1156 *cactorum*.

1157

Treatments	No. of flowers	Fruit set % (N)	Seed set mean \pm SD	Seed set in relation to ovule number (%)
Autonomous self-pollination	40	10 (4)	2.0 \pm 1.4	2.7
Hand self-pollination	31	32.2 (31)	2.3 \pm 1.2	3.2
Hand cross-pollination	20	100 (20)	66.2 \pm 27.9	91.7
Natural pollination	31	100 (31)	62.8 \pm 18.4	87.1
<i>Arhysosage cactorum</i> absence	33	33.3 (33)	13.1 \pm 13.2	18.2

1158

1159

1160

1161

1162

1163

1164 **Table 2.** Flower-visiting bees of *Parodia neohorstii*. Frequency of floral visits were measured only in
 1165 2016 during 75 hours of observation (N = 2284 visits). (*) Species not recorded in 2016 and not included
 1166 in the calculation of the frequency. We only observed *Ancyloscelis romeroi* in 2017 as a sporadic visitor
 1167 (N = 4 visits).

Bee species	Tribe / Subfamily	Sex	Frequency (%)
Andrenidae			
<i>Arhysosage cactorum</i> Moure 1999	Calliopsini	♀ ♂	81.21
<i>Callonychium</i> sp.	Calliopsini	♀	0.04
<i>Anthrenoides micans</i> Urban 1955	Protandrenini	♀ ♂	16.90
Apidae			
<i>Ancyloscelis romeroi</i> Holmberg 1903	Emphorini	♀	*
<i>Ceratina</i> sp.	Ceratinini	♀	0.17
Colletidae			
<i>Cephalocolletes rugata</i> Urban 1995	Neopasiphaeinae	♀	0.48
Halictidae			
<i>Dialictus</i> sp.	Halictini	♀	1.18

1168

1169

1170 **Table 3.** Landing site, stigma contacts and duration of flower visits of frequent bee visitors (> 1% of
 1171 visits) to flowers of *Parodia neohorstii*. Visit duration was quantified only for female bees. Landing
 1172 places: Sg = stigmatic surface, P = petals, S = stamens.

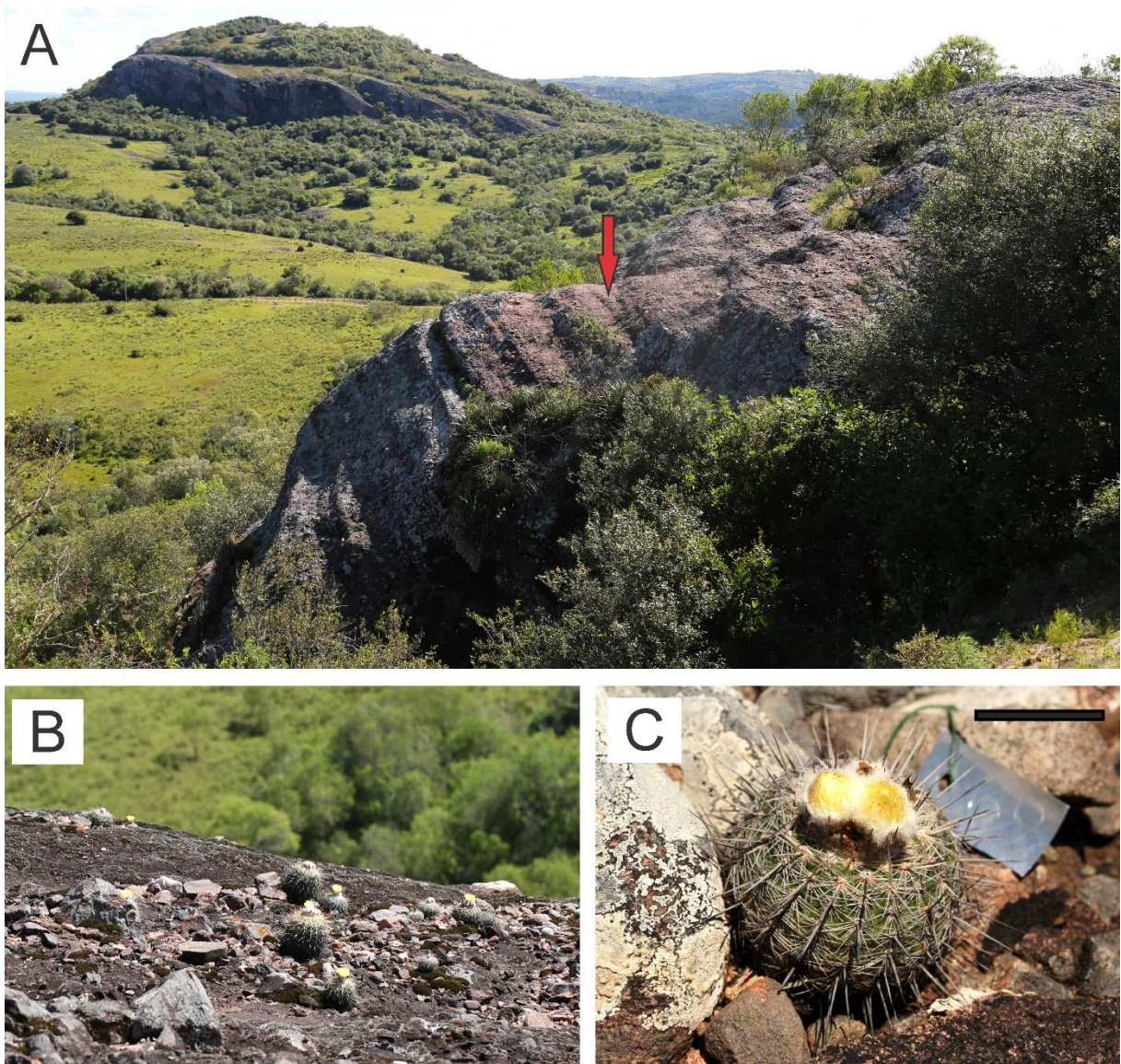
Bee species	Sex	Landing place			(N)	Stigma touches (%)	Visit duration (s)
		Sg	P	S			
<i>Arhysosage cactorum</i>	♀	62%	24%	14%	79	87% (N = 129)	18.4 ± 14.9
	♂	64%	15%	21%	140	76% (N = 222)	-
	Couple	50%	50%	0%	14	97% (N = 84)	13.8 ± 13.4
<i>Anthrenoides micans</i>	♀	21%	62%	17%	128	47% (N = 182)	6.8 ± 5.5
	♂	69%	8%	23%	49	70 % (N = 79)	-
	Couple	-	-	-	-	80 % (N = 10)	-
<i>Dialictus</i> sp.	♀	6%	82%	12%	103	22% (N = 27)	5.5 ± 6.9

1173

1174

1175 FIGURES

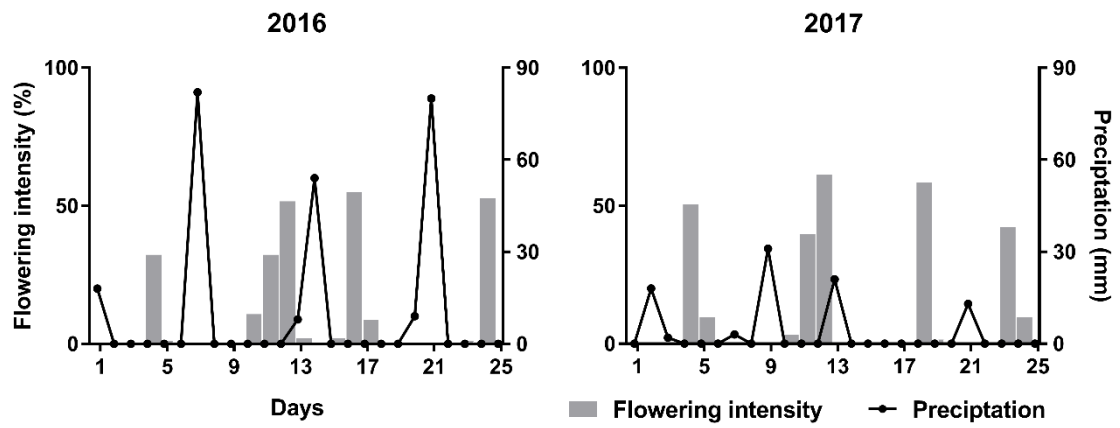
1176



1177

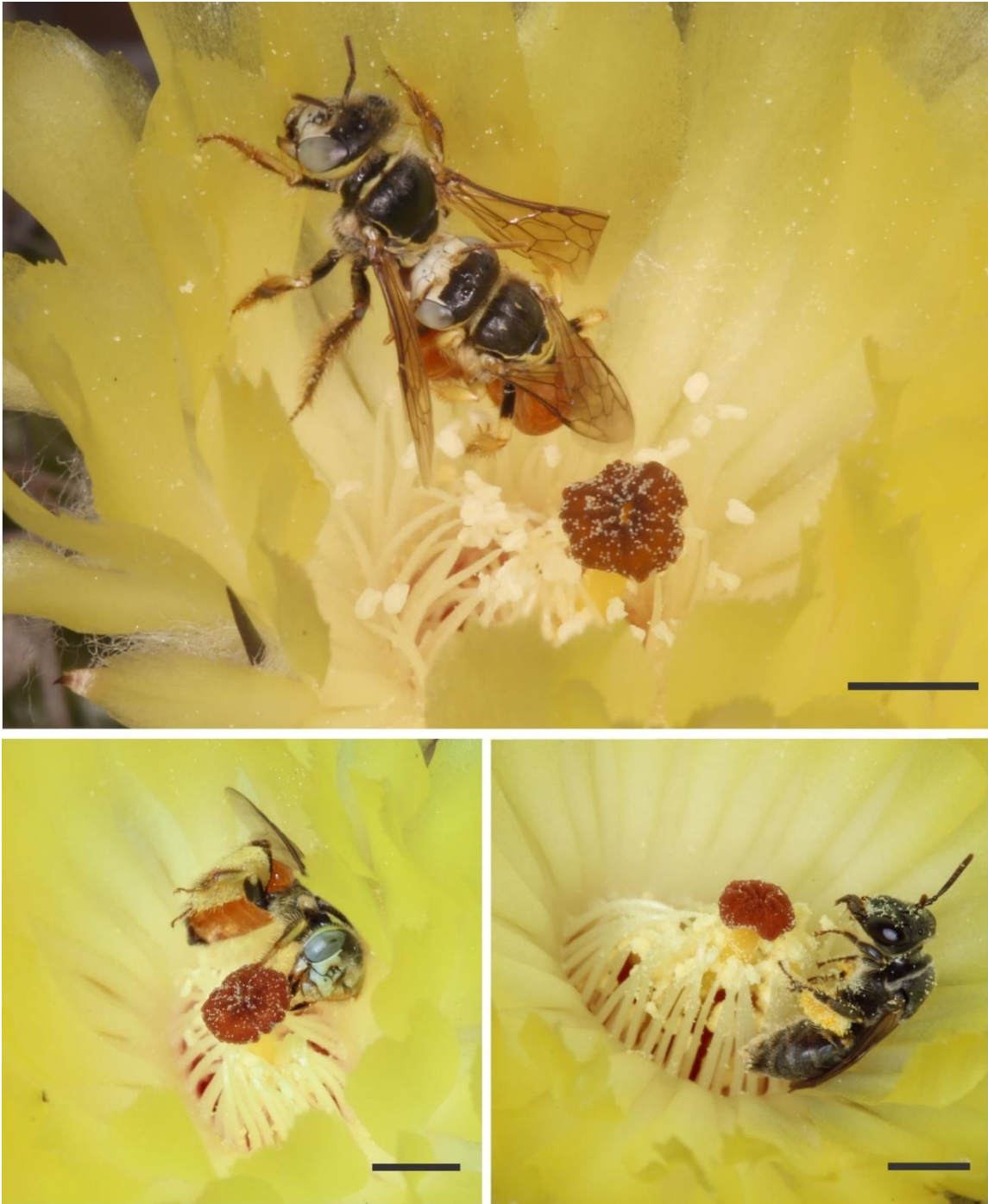
1178 **Figure 1.** Study area and habitat characteristics. (A) Sandstone and conglomerate rock outcrops in the
 1179 Guarita-region of the state of Rio Grande do Sul. The red arrow indicates the location of the studied
 1180 population. (B) Individuals of *Parodia neohorstii* growing on rock substrate almost without soil. (C)
 1181 Flowering cactus with aluminum tag. Scale bar = 35 mm.

1182



1183

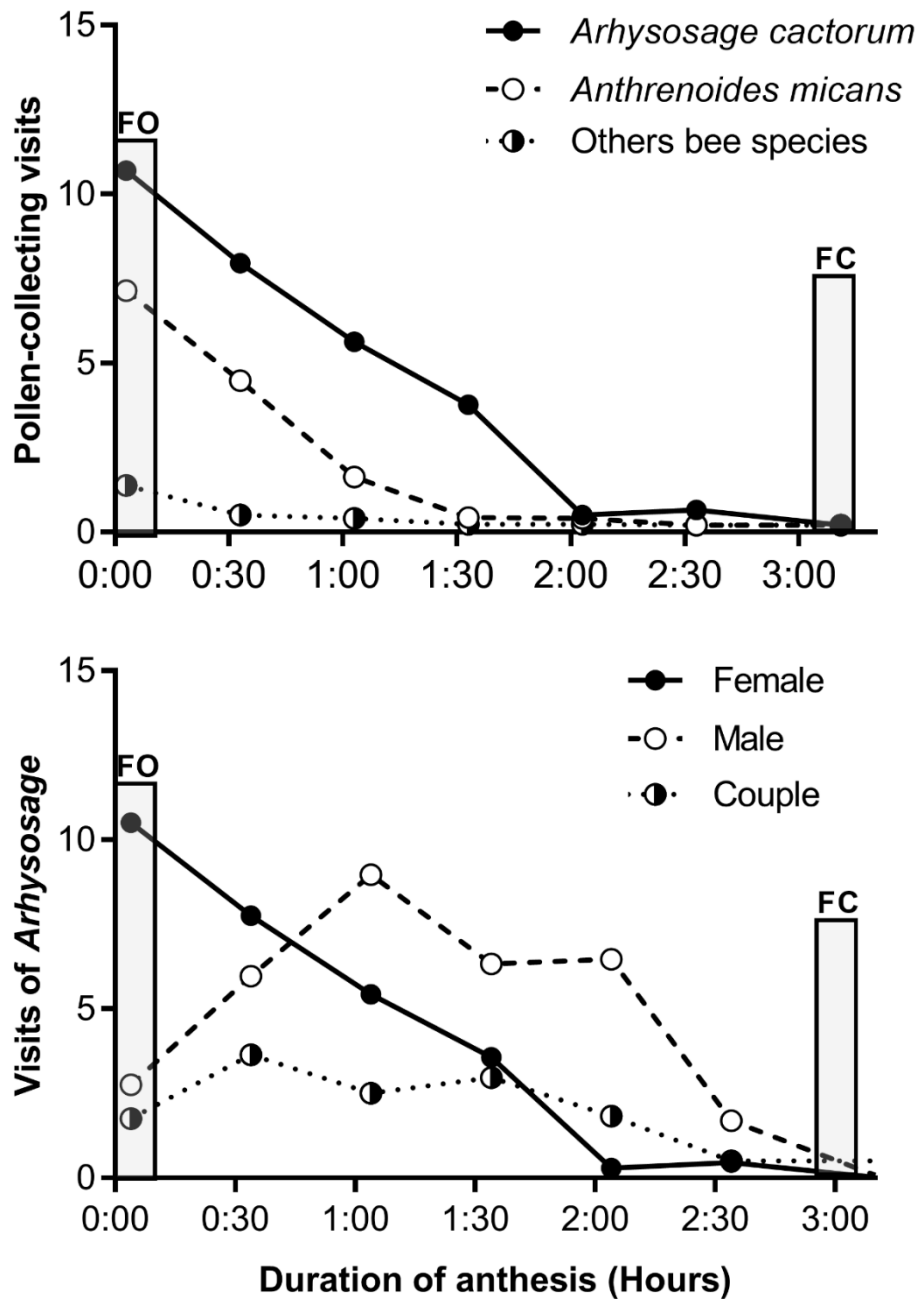
1184 **Figure 2.** Flowering intensity (number of flowering individuals/total number of reproductive
 1185 individuals) of *Parodia neohorstii* and precipitation from October 27 (day 1 of X axis) to November 20,
 1186 during 2016 and 2017.



1187

1188 **Figure 3.** Main floral visitors of *Parodia neohorstii*. (A) A pair of *A. cactorum* in copula: male is
 1189 clutching the female with its mandibles. (B) Female of *A. cactorum* collecting pollen. (C) Female of *A.*
 1190 *micans* collecting pollen. Note the different color of the pollen load in the scopa of *A. micans*. Stimulated
 1191 stamens contacting the style in (B) and (C). Scale bars = 2 mm.

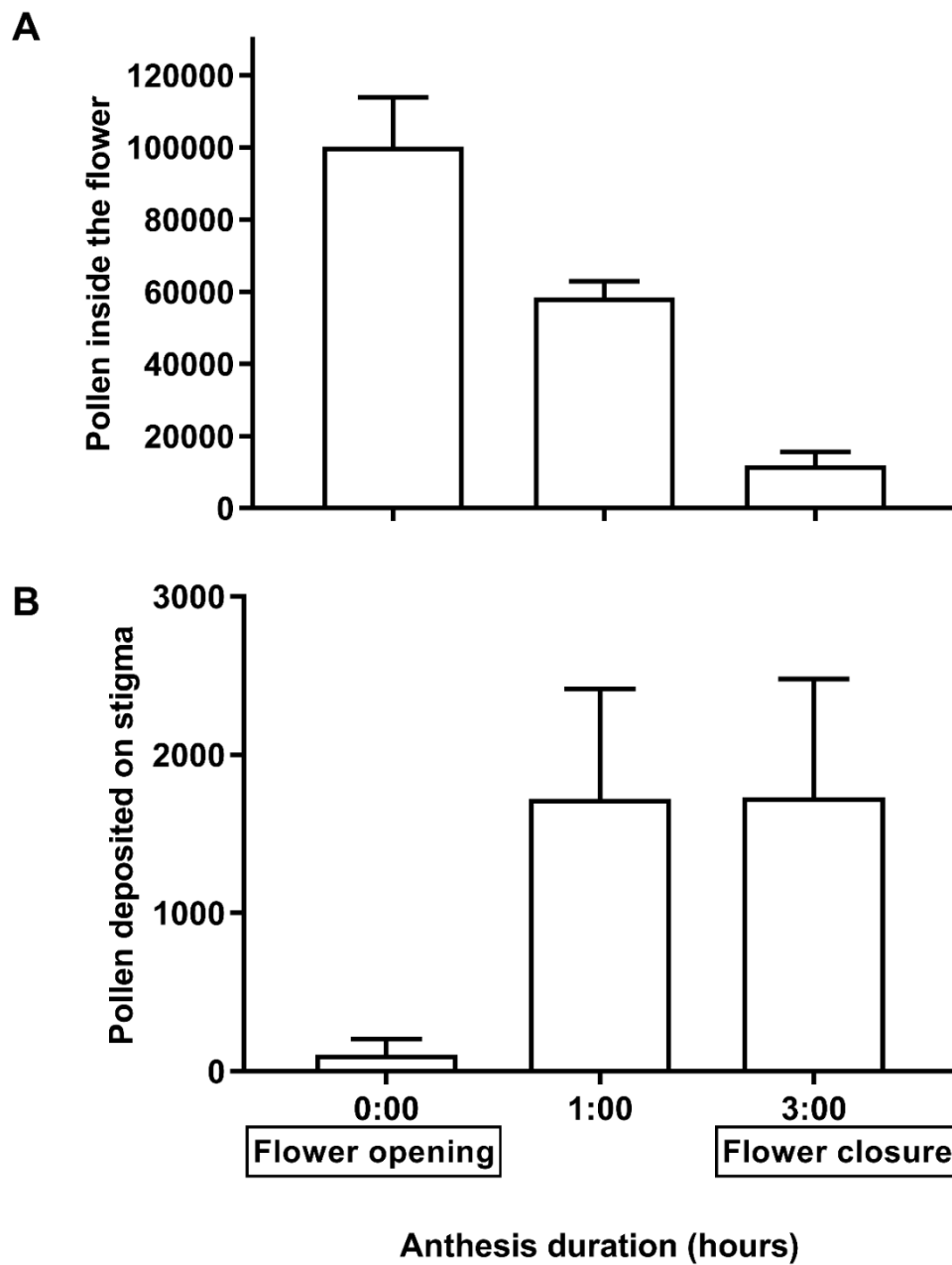
1192



1193

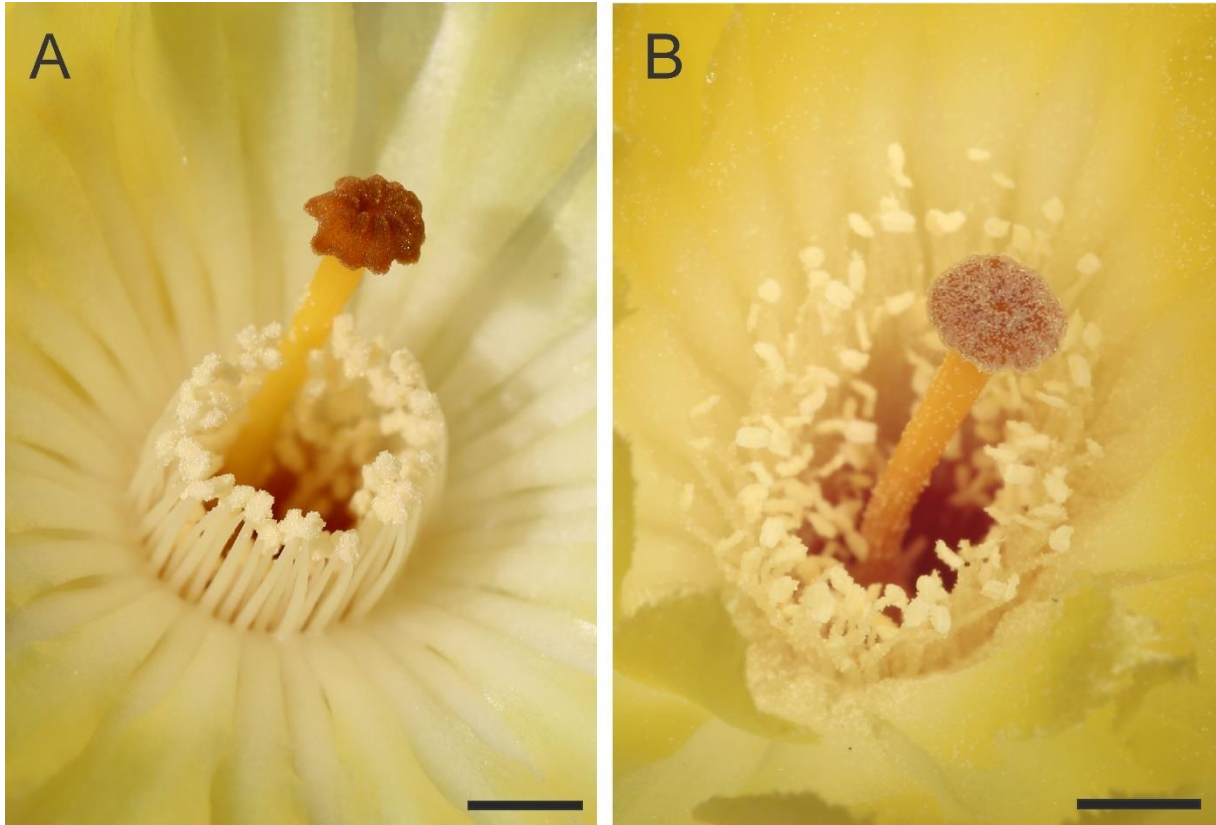
1194 **Figure 4.** Flower visits for *Parodia neohorstii* throughout anthesis. (A) Mean number of pollen1195 collection visits by female bees. (B) Mean number of visits by males, females and couples of *A.*1196 *cactorum*. FO = flower opening and FC = flower closure.

1197



1198

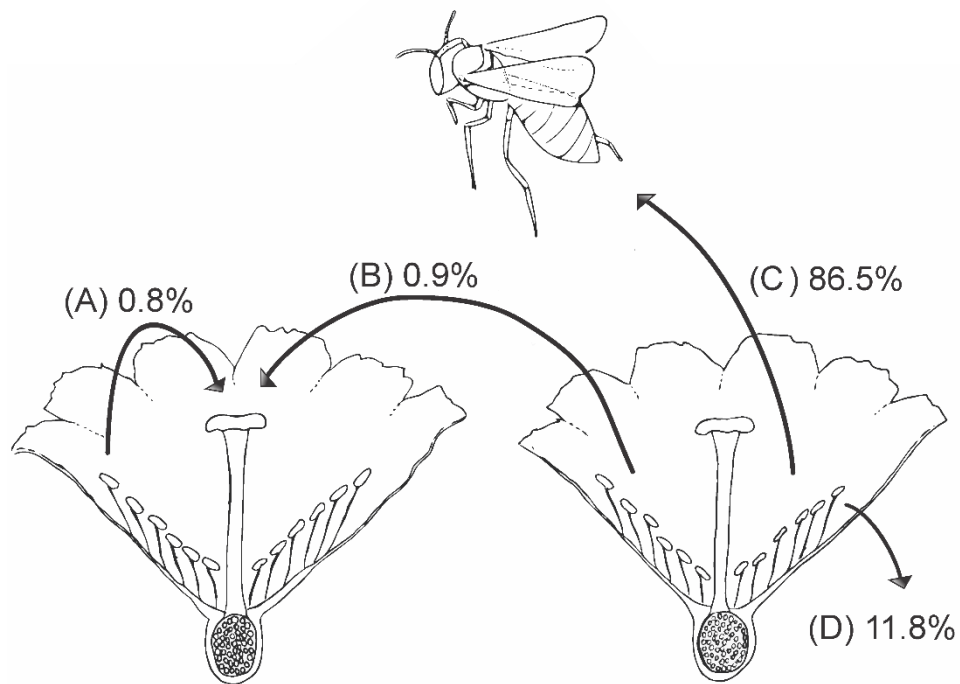
1199 **Figure 5.** Pollen removal and deposition by bees in flowers of *Parodia neohorstii*. Pollen grains inside
 1200 the flower (A) and pollen deposited on the stigma (B) after bee visits during different moments of
 1201 anthesis (N = 10).



1202

1203 **Figure 6.** Deposition of pollen grains on the stigma of *Parodia neohorstii*. (A) Freshly open flower with
1204 stamens still upright and dehiscent anthers. Pollen and anthers do not contact the stigmatic surface. (B)
1205 Flower at the end of anthesis near flower closure. Stigmatic surface saturated with pollen grains. Note:
1206 pollen grains adhered to petals and style. Scale bar = 2 mm

1207



1208

1209 **Figure 7.** Pollen fate for *Parodia neohorstii*. Percentages represent the fate of all pollen produced by a
 1210 flower at the end of anthesis. (A) Autogamous pollen deposited on a stigma after the movement of bees
 1211 inside the flower. (B) Xenogamous pollen deposited on a stigma by bees. (C) Pollen collected by bees.
 1212 (D) Pollen remaining in the flower after closure.

1213

CAPÍTULO 2

1214

1215

1216

1217

1218

1219

1220

1221

**STIMULATION OF THIGMONASTIC STAMENS BY BEES INDUCE FLORAL
SENESCENCE IN CACTUS FLOWERS ²**

1222

² O manuscrito foi submetido para a revista *Annals of Botany*. Autores: Cerceau, I & Schlindwein, C.

1250 **INTRODUCTION**

1251

1252 Floral longevity corresponds to the period from the beginning of anthesis until floral
 1253 senescence (Primack, 1985), and its plasticity may be a feature that influences the reproductive
 1254 success of pollen dispersal and ovule fertilization (Ashman & Schoen, 1994). Pollen removal,
 1255 as well as pollen deposition on the stigma, can increase when flowers are open for a longer
 1256 period (Ishii & Sakai, 2000; Blair & Wolfe, 2007; Aximoff & Freitas, 2010). The extension of
 1257 floral longevity in situations with low pollinator activity may be an alternative to increase the
 1258 chances of reproduction (Rathcke, 2003). Flower maintenance for long periods, otherwise,
 1259 comes at an energy cost and may lead to the production of low-quality fruits (i.e. reduced seed
 1260 number) (Ashman & Schoen, 1997; Castro *et al.*, 2008).

1261 In general, the period in which flowers remain open and functional varies among species
 1262 from a few hours up to months (Primack, 1985; Ashman & Schoen, 1994). Floral lifespan can
 1263 be influenced by abiotic factors such as drought and high temperature and biotic factors
 1264 (Primack, 1985; Teixido & Valladares, 2015). Moreover, there is often a relationship between
 1265 floral longevity and pollination in long-lasting flowers, since pollen deposition to the stigmas
 1266 can trigger physiological responses that induce petal wilting and abscission (van Doorn, 1997;
 1267 Rogers, 2006; Tripathi & Tuteja, 2007; Araujo *et al.*, 2020).

1268 Pollen deposition on the stigma triggers senescence in many species, e.g. *Digitalis*
 1269 *purpurea* (Plantaginaceae), *Erythranthe guttata* (Phrymaceae) and *Epilobium angustifolium*
 1270 (Onagraceae) (Stead & Moore, 1979; Arathi *et al.*, 2002; Clark & Husband, 2007). Pollen
 1271 removal may influence floral longevity like in protandrous Campanulaceae, where the duration
 1272 of the staminate phase diminishes after pollen removal by bee pollinators (Devlin &
 1273 Stephenson, 1984; Richardson & Stephenson, 1989, Evanhoe & Galloway, 2002). In flowers
 1274 of *Blumembachia* (Loasaceae), nectar uptake by specialized bee pollinators accelerates pollen
 1275 presentation and shortens the staminate phase (Siriani-Oliveira *et al.*, 2018, 2020). Moreover,
 1276 in many orchid species, the potentially long-lived flowers rapidly wilt after pollinarium
 1277 deposition on the stigma (Ashman & Schoen, 1994; Proctor & Harder, 1995; Martini *et al.*,
 1278 2003; Abdala-Roberts *et al.*, 2007).

1279 *Parodia neohorstii* (Cactaceae) a globular cactus from southern Brazil, is an example of a
 1280 species with strong plasticity in flower longevity (Cerceau *et al.*, 2019). Flowers open
 1281 synchronously at about 13 p.m. and present thigmonastic stamens that move to the center of the
 1282 flower after mechanic stimuli at the filaments. Flowers are pollinated by pollen-specialist bees.

1283 Floral longevity is ephemeral, lasting only three hours in the presence of the specialized
1284 pollinators, which promote maximum pollen deposition to stigmas in just one hour and while
1285 continuously triggering stamen movements (Cerceau *et al.*, 2019). When pollinators are absent,
1286 or when flowers are bagged, flowers open again on the following days (Cerceau *et al.*, 2019).

1287 Having discovered this variation in floral longevity in *P. neohorstii* according to the
1288 presence or absence of pollinators and, the shortening of floral longevity after pollen deposition
1289 on the stigmas in many plant species (Stead & Moore, 1979; Arathi *et al.*, 2002; Clark &
1290 Husband, 2007), we hypothesized that floral longevity in *P. neohorstii* shortens through
1291 deposition of conspecific pollen deposition on the stigma. We also considered whether flower
1292 manipulation by bees influences flower longevity. We focused on two questions: 1) Does
1293 deposition of pollen on the stigma of *P. neohorstii* flowers shortens floral duration? 2) Does
1294 triggering of thigmonastic stamen movements during pollen collection by bees influence the
1295 floral longevity?

1296 To answer these questions, we performed controlled pollination experiments with
1297 different pollen sources (autogamous and xenogamous pollen), determined which bees trigger
1298 stamen movements, and applied mechanical stimuli to the stamens, simulating bee visits to
1299 flowers, with and without pollen deposition to the stigmatic surface.

1300

1301 MATERIAL AND METHODS

1302

1303 *Study site*

1304

1305 The study was conducted in the Serra do Sudeste in the state of Rio Grande do Sul,
1306 Brazil. We studied three populations of *Parodia neohorstii* located in the region of Guaritas
1307 near the mining town of Minas do Camaquã (30°53'36.3"S, 53°25'14.3"W) between October
1308 and December of 2016, 2017 and 2018. The region is situated in the Pampa biome and is
1309 characterized by steep blocks of sandstone and conglomerate of up to 100 meters height. The
1310 local vegetation is composed of a rich herbaceous flora with shrubs and small trees (Carneiro
1311 *et al.*, 2016). The local bee fauna is diverse and contains a high proportion of oligolectic species
1312 (Schlindwein, 1998). Oligolectic bees are pollen specialists that collect this resource only in
1313 plants of the same genus or family to feed their larvae (Robertson, 1925). The climate is sub-
1314 temperate and humid (Maluf, 2000), with an average annual temperature of 18.7 °C and an
1315 average annual precipitation of 1509 mm (INMET, 2018).

1316

1317 *Plant species*

1318

1319 *Parodia neohorstii* is an endangered cactus endemic to the region of Guaritas. The small
 1320 globose cacti grow as solitary individuals on sun-exposed rock substrate. The flowering season
 1321 is between end of October and early December. The cacti produce on average five flowers per
 1322 year but generally only one flower opens per day from 13:00 to 16:00h, which is intensely
 1323 visited by solitary bees. Oligolectic *Arhysosage cactorum* (Andrenidae, Calliopsini) and
 1324 *Anthrenoides micans* (Andrenidae, Protandrenini) are the commonest flower visitors and
 1325 pollinators (Cerceau *et al.*, 2019). *Parodia neohorstii* has thigmonastic stamens that bend
 1326 inwards toward the style after mechanic stimuli of the filaments. The stamens exhibit two main
 1327 positions: (1) Original position, where stamens are sprawled erect and flower base and style are
 1328 freely accessible; (2) Stimulated position, where stamens bent inwards or bent forwards towards
 1329 to the center of the flower and lean onto the style after mechanical stimuli. After some minutes,
 1330 the stamens bend back to the original position ("unstimulated stamens").

1331 Vouchers of the studied species were deposited at BHCB herbarium - Universidade
 1332 Federal de Minas Gerais, Belo Horizonte, Brazil (BHCB 194347 and 194073).

1333

1334 *Description of stamen movements in Parodia neohorstii*

1335

1336 To describe in detail the stamen movements, we touched the filaments with a paintbrush
 1337 and measured the time stamens took to move to the center of the flower and the time interval
 1338 to return to the original position (N = 30).

1339

1340 *Floral longevity and senescence in natural conditions*

1341

1342 Under natural conditions, floral longevity of *P. neohorstii* increases in bagged flowers
 1343 and in the absence of pollinators (Cerceau *et al.*, 2019). To quantify changes in floral longevity
 1344 according to the visiting bee species, we marked flowers and observed floral visitors throughout
 1345 the anthesis during the flowering periods of *P. neohorstii* during three years. Observations were
 1346 made at the beginning of the flowering season in 2017 and 2018 (N = 21 flowers), when the
 1347 main pollinator species, *Arhysosage cactorum*, was still inactive and in the middle of the
 1348 flowering period in 2016, 2017 and 2018 (N = 187 flowers), when bees of this species were the

1349 most frequent floral visitor. Marked flowers were accompanied daily to check for flower
1350 opening, and consistence of the petals (turgid or wilted).

1351

1352 *Factors that alter floral longevity*

1353

1354 To verify which factors influence floral longevity, we performed eight treatments
1355 involving deposition of pollen grains on the stigmas and simulation of bee visits through
1356 experimental induction of thigmonastic stamen movements by mechanic stimuli. We
1357 individually marked floral buds according to the treatment they received and monitored the
1358 flowers over time until senescence. All flowers were bagged before and after treatments. We
1359 checked whether flowers opened in subsequent days, the opening hours and the consistence of
1360 the petals (turgid or wilted). We compared the results with non-manipulated flowers accessible
1361 to pollinators.

1362 To know if pollination alters floral longevity, we performed four manual pollination
1363 treatments: (1) hand cross-pollination with one pollen donor from another individual (n = 49);
1364 (2) hand cross-pollination with five pollen donors from five individuals (n = 31); (3) hand self-
1365 pollination (n = 40); (4) autonomous self-pollination (n = 60).

1366 We performed four treatments simulating bee visits to find out whether mechanic
1367 contacts of the pollinators to the filaments alter floral longevity. For this, we gently touched
1368 filaments with a paintbrush to trigger stamen movements. The stimulation of the stamens was
1369 repeated every five minutes (roughly the period stamens took to return to their original
1370 position). The flowers submitted to these treatments were bagged again after the hand
1371 stimulations to keep them inaccessible to floral visitors.

1372 In order to verify the effect of stamen movements on floral longevity without the
1373 influence of pollination, we performed paired treatments with or without pollen deposition on
1374 the stigmas: (1) five stimuli to stamens with xenogamous pollen deposition on the stigmas (n =
1375 40); (2) five stimuli to stamens without pollen deposition on the stigmas (n = 40); (3) ten stimuli
1376 to stamens with xenogamous pollen deposition on the stigmas (n = 40); (4) ten stimuli to
1377 stamens without pollen deposition on the stigmas (n = 40).

1378

1379 *Bee species that trigger stamen movements*

1380

1381 To determine which bee species triggers stamen movements during flower visits, we
 1382 observed flowers with unstimulated stamens in the field (15 h) and made video recordings of
 1383 bee visits to flowers (10 h) with digital cameras (Canon EOS T5i and 5D) and camcorder
 1384 (Panasonic HC-X929) in November of 2016, 2017 and 2018. We recorded the duration of bee
 1385 visits and verified whether stamens moved to the flower center after the visits. We classified
 1386 floral visitors by sex and noted whether they were in copula position.

1387 To correlate body size of the bees with successful stamen movements, we collected bee
 1388 individuals in the flowers and measured nine regions of the insect body (N=20 individuals per
 1389 sex) (see below) in the laboratory. To obtain a more precise measure of the body volume of
 1390 floral visitors, we used the mathematical formula of ellipsoid volume. For this, we divided the
 1391 body of the bees into the three sections: head, mesosoma and metasoma, and each of the sections
 1392 was measured for height, diameter and width. The measurements of each body section were
 1393 inserted in the formula:

1394

1395 *Volume of each body section = $4/3 * \pi * (height/2) * (diameter/2) * (width/2)$*

1396

1397 To calculate the total volume of the bees' body, we summed the volume of each section:

1398

1399 *Volume of bee body = (Volume of head) + (Volume of mesosoma) + (Volume of metasoma)*

1400

1401 For bees in copula position, we summed the body volume of females and males.

1402 Bee specimens were deposited in the Entomological Collection of UFMG (Centro de Coleções
 1403 Taxonômicas da UFMG, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil).

1404

1405 *Statistical analyses*

1406

1407 To compare floral longevity under natural conditions between flowers visited and non-
 1408 visited by the main pollinator, we performed a Mann-Whitney Rank Sum Test.

1409 To compare floral longevity in different pollination treatments, we performed the test Kruskal-
 1410 Wallis One Way Analysis of Variance on Ranks.

1411 To compare floral longevity in flowers receiving 5 or 10 manual stimuli in stamens, we
 1412 performed a Mann-Whitney Rank Sum Test. The same test was performed to compare flowers

1413 that received the same number of manual stimuli in stamens but differed in pollen deposited on
1414 the stigma or not.

1415 To compare the body volume of different bee species, we used the following tests: 1)
1416 Kruskal-Wallis One Way Analysis of Variance on Ranks to compare more than two species;
1417 and 2) Student's t-test to compare only two species.

1418

1419 **RESULTS**

1420

1421 *Stamen movements in Parodia neohorstii*

1422

1423 After mechanical touches of the filaments, stamens immediately bent inward and
1424 reached the style (Figure 1) on average after 2.1 seconds (± 0.4 , N = 30 flowers). Stamens
1425 returned to the original position on average after 280.7 seconds (± 69.4 ; N = 30). Under natural
1426 conditions, the stamens are stimulated after visits by bees (Figure 2).

1427

1428 *Floral longevity*

1429

1430 Anthesis of new flowers of *P. neohorstii* started around 1239 h (± 26 minutes) and
1431 flowers closed at 1539 h (± 32 minutes) (N=68 flowers). Bagged flowers, not visited by bees,
1432 reopened at the subsequent day at 1138 h (± 29 min) and closed at the same time as first-day
1433 flowers (1533 h ± 17 minutes) (N=23). Flowers, thus, opened for 3 hours on the first and for 4
1434 hours on the second day of anthesis.

1435

1436 *Pollination and stamen stimulation experiments*

1437

1438 All pollination treatments with bagged flowers resulted in similar floral longevity of the
1439 *Parodia* flowers (H = 0.232, d.f. = 3, P = 0.972, N = 180, Figure 3A). Self-pollinated flowers
1440 had similar floral longevity that cross-pollinated flowers. Floral longevity was on average 2.3
1441 ± 0.7 days for all pollination treatments, and 84.4% of the flowers opened 2 or 3 days. In some
1442 of these hand cross-pollinated flowers, floral longevity reached up to 5 days.

1443 Floral longevity was short only in flowers whose filaments received experimental
1444 mechanical stimuli that resulted in thigmonastic stamen movements (Figure 3B), and 94.3% of
1445 its flowers lasted only one day (N = 160).

1446 There was no difference in the floral longevity among the hand stimulated flowers that
 1447 were hand cross-pollinated from those that were not pollinated. Pollen deposition on the stigma
 1448 surface, therefore, had no influence on the floral longevity (stamens five times hand-stimulated:
 1449 $U = 774.5$, $P = 0.477$, $N = 80$ / stamens ten times hand-stimulated: $U = 800$, $P = 0.099$, $N = 80$).
 1450 Moreover, flower longevity did not differ in flowers, whose stamens were stimulated
 1451 experimentally five or ten times (flowers with pollen deposition on stigmas: $U = 740$, $P = 0.240$,
 1452 $N = 80$ / no pollen deposition on stigmas: $U = 720$, $P = 0.115$, $N = 80$).

1453

1454 *Bee species that trigger stamens movement*

1455

1456 Visiting female bees in flowers of *P. neohorstii* search only for pollen, since flowers do
 1457 not produce nectar (Cerceau *et al.*, 2019). The flowers of *P. neohorstii* were visited by four
 1458 species of bee (Table 1). Of the two panurgine species *Arhysosage cactorum* and *Anthrenoides*
 1459 *micans*, which were the most common visitors, both females and males visited the flowers. For
 1460 both species, we found bees copulating in the flowers or visiting the flowers in copula position,
 1461 which was very common for *A. cactorum* (Figure 4, Table 1).

1462 The body volume of the four species differed significantly among the species ($P =$
 1463 <0.001 , $H = 156.81$, $d.f. = 8$, $N = 155$, Figure 5). The triggering of stamen movements in a flower
 1464 visit was related to the body volume of the bees: larger bees triggered movements in most visits,
 1465 while the smaller bees (*Dialictus sp.*) never triggered stamen movements. Visits of couples,
 1466 with around double body volume, triggered more stamen movements when compared to visits
 1467 of solitary females of the same species.

1468 The behavior of the bees inside the flower also influenced the triggering of stamen
 1469 movements. Pollen collecting females stimulated more stamen movements than males of the
 1470 same species (*A. micans*: $t = -10.263$, $d.f. = 33$, $P = <0.001$, $N = 35$; *A. cactorum*: $t = -6.021$,
 1471 $d.f. = 38$, $P = <0.001$, $N = 40$). Males remained for a shorter time in a flower than females and
 1472 rarely touched the stamens. Males of *A. cactorum* made quick flower visits on their patrol flights
 1473 and sometimes hid between the stamens, remain immobile with their head among anthers
 1474 looking outward and, in general, induce movements of only a few stamens. On their patrol
 1475 flight, males of *A. micans* predominantly fly over flowers and seldom visit them. On their few
 1476 quick visits, they alight on the petals, rarely touch stamens, and therefore never triggered stamen
 1477 movements. Females of *A. cactorum* actively collect pollen, moving their legs quickly between
 1478 the bundle of stamens and stay longer inside the flowers (on average 17.9 ± 14.6 seconds),

1479 which promoted stamen movements during almost all visits. During flights in copula position,
 1480 females of *A. cactorum* carried the male actively but continued to collect pollen in a uniform
 1481 manner. In the few flower visits observed, females of *Cactocolletes rugatus* always triggered
 1482 stamen movements due to their intense movements during pollen collection. Females of the
 1483 tiny *Dialictus sp.* generally landed on the petals and collected pollen grains fallen down from
 1484 anthers to the petals or the flower tube after flower visits of larger bees and rarely removed
 1485 pollen directly from anthers. Therefore, they had little contact with the filaments and never
 1486 triggered stamen movements (Table 1). Females of *A. micans* collect pollen actively from the
 1487 anthers, nevertheless, they stimulated stamens in only one third of the visits. However, when
 1488 females of *A. micans* were in copula position within a flower, the rate of stamen movements
 1489 rose to 90.9% (Table 1). Considering that copulations lasted a long time (up to 7 min), couples
 1490 remained immobile within the flowers during most of the time.

1491

1492 *Flower longevity in different natural conditions*

1493

1494 Under natural conditions, there are two scenarios of abundance of main flower visitors.
 1495 In the flowering peak, when females and males of *A. cactorum* were the most frequent floral
 1496 visitors, floral longevity lasted only one day (1.0 ± 0.0 d, N = 187). In this period, flower visits
 1497 of other bee species were rare. In the beginning of the flowering period, when bees of *A.*
 1498 *cactorum* were absent, flower longevity lasted on average 2.2 ± 0.4 days (N=21), demonstrating
 1499 that floral longevity varied under natural conditions (T = 4158.000, U= 0.000 P = <0.0001, N
 1500 = 208, Figure 6). In this period, flowers were visited mainly by bees of *Anthrenoides micans*
 1501 and *Dialictus sp.* (Figure 7).

1502

1503 **DISCUSSION**

1504

1505 Our results reveal that only the stimulation of the thigmonastic stamens is responsible
 1506 for the shortening of the floral longevity in *Parodia neohorstii* to the three-hour lifespan.
 1507 Neither self nor cross-pollen deposition to the stigma surface cause any effect on the duration
 1508 of the cacti flowers. This is surprising, because in most species with variable flower longevity,
 1509 the deposition of pollen to the stigmatic surface is one of the most important factors that induces
 1510 floral senescence, once the female function has been fulfilled (Arathi *et al.*, 2002; Clark &
 1511 Husband, 2007; Niu *et al.*, 2011). In *P. neohorstii*, pollen dispersal to the body of effective

1512 pollinators occurs together with the induced movements of stamens, and the lack of these
 1513 movements may indicate a scarce pollination environment. Consequently, the increased floral
 1514 longevity favors male function and enhances the probability of pollen export to conspecific
 1515 flowers.

1516

1517 *Pollination effectiveness of stamen movement triggering bees*

1518

1519 Some species can alter floral longevity according to the abundance of floral visitors and
 1520 extend flower lifespan to increase the chances of pollination in scenarios of pollinator scarcity
 1521 (Harder & Johnson, 2005). It is interesting that even receiving pollen on the stigma, floral
 1522 longevity of *P. neohorstii* does not vary with the abundance of floral visitors, but with the
 1523 quality of bee species, being regulated by its effective pollinators.

1524 Whether flower visiting bees trigger stamen movements or not is related to two main
 1525 factors:

- 1526 1. Body volume and strength of bees: Small bees do not trigger stamen movements
 1527 because they do not have enough strength and body mass to induce thigmonastic stamen
 1528 movements. The insects require a minimum force and weight to surpass the stimulus
 1529 threshold to activate the movement of stamens. In *Opuntia polyacantha* flowers,
 1530 thigmonastic stamen movements are activated only by larger floral visitors (Cota-
 1531 Sánchez *et al.*, 2013), but the force required to trigger the movements has not been
 1532 measured. The strength of bees that is closely related with body mass was shown to be
 1533 the key factor to open keels in papilionid flowers (Córdoba & Cocucci *et al.*, 2011) or
 1534 activate the catapult mechanism in a species of *Cornus* for explosive pollen dispersal
 1535 (Edwards *et al.*, 2005).
- 1536 2. Flower handling: Males exhibit different flower handling, because they do not search
 1537 for floral resources in the nectarless flowers, which reduces the triggering of stamen
 1538 movements. On their search for females, males make short inspection visits in the
 1539 flowers of *P. neohorstii* and touch the stamens only slightly. Females of *A. cactorum*,
 1540 *A. micans* and *C. rugatus*, however, intensely contact filaments and anthers during
 1541 pollen collection, stay much longer inside the flowers and rashly move around the
 1542 stamen ring, but females of the small *Dialictus sp.* rarely touch the stamens during
 1543 pollen gleaning and never trigger stamen movements.

1544 Females and males of *A. cactorum* frequently fly in copula position from flower to flower.
 1545 While the male-carrying female continues pollen foraging (Schlindwein & Wittmann, 1995;
 1546 Oliveira, 2015; Cerceau *et al.*, 2019), the couples stimulated stamen movements in all visits,
 1547 given their large body volume and the intense prolonged pollen collection by the females. This
 1548 is also true for pairs of *A. micans*. However, flowers visits in copula position of this species are
 1549 rare, the couples often remain in the same flower for minutes, and the females suspends pollen
 1550 collection in this period (Dutra *et al.*, 2020).

1551 In pollen rich cactus flowers, small bees can easily fill their scopae in a single flower visit
 1552 without contact with receptive stigmas, as occurs in cacti flowers of *Parodia*, *Gymnocalycium*
 1553 (Schlindwein & Wittmann, 1995) and *Opuntia* (Barrows *et al.*, 1976, Osborn *et al.*, 1988). Due
 1554 to the restricted foraging of small bees on the same flower and neighboring flowers, the
 1555 contribution to pollination is minimal (Schlindwein & Wittmann, 1995).

1556 Larger bees, however, forage on various flowers to fill the scopae. They usually use the
 1557 stigma as a landing platform, which favors xenogamous pollen deposition (Cerceau *et al.*,
 1558 2019). Besides that, the flowers with stimulated stamens reduce the amount of pollen collected
 1559 by individual bees (Schlindwein & Wittmann, 1997, Cerceau *et al.*, 2019), which encourages
 1560 bees to move to flowers of other plant individuals, enhancing their pollination effectiveness.

1561 In *P. neohorstii*, the triggering of the stamen movements seems to indicate the presence of
 1562 effective pollinators and likely pollen export, which thus could be interpreted as the signal to
 1563 close flowers after the first day of blooming.

1564

1565 *Enhanced flower lifespan in the absence of bees that trigger stamen movements*

1566

1567 In most species with short-lived ephemeral flowers, pollination does not induce floral
 1568 senescence (Ahmad & Tahir, 2016) and flower lifespan seems to be fixed and regulated by
 1569 internal factors (Stead, 1992). Floral longevity of *P. neohorstii* is ephemeral only after
 1570 triggering of stamen movements. The indication of a scarce pollination environment through
 1571 missing or weak thigmonastic stamen movements linked to increased flower lifespan worked
 1572 well under natural conditions: when bees of oligolectic *A. cactorum* were absent at the
 1573 beginning of the flowering period in two years (2017 and 2018), flowers opened again on
 1574 subsequent days. This occurs due to the presence of almost exclusively small bees and the
 1575 absence of stamen movement triggering bees.

1576 Curiously, flower opening of second- and third day flowers is one hour earlier than that
 1577 of new flowers. The one-hour advanced flower opening of the old in comparison to first-day
 1578 flowers might favor the old flowers by anticipated flower visits and pollination. The flower
 1579 visitation rate of the effective pollinator *A. cactorum* is highest at the opening time of first-day
 1580 flowers, and a one-hour time interval was enough for these bees to deposit the maximum
 1581 amount of pollen to the stigma to fertilize all ovules (Cerceau *et al.*, 2019).

1582

1583 *The function of stamen movements*

1584

1585 Stamen movements occur in at least 15 plant families (Sibaoka, 1969; Simons, 1981;
 1586 Schlindwein & Wittmann, 1997a, 1997b; Ren, 2010; Du *et al.*, 2012; Ren & Bu, 2014). In some
 1587 cases, it was demonstrated that stamen movements enhance pollen deposition on the pollinator
 1588 body (Rathcke & Real, 1993; Angulo *et al.*, 2014) and pollen flow among conspecific plants
 1589 through pollen partitioning (Schlindwein & Wittmann, 1997a; Henning & Weigend, 2012;
 1590 Siriani-Oliveira *et al.*, 2018, 2020).

1591 In Cactaceae, thigmonastic stamen movement occurs in at least 10 genera (Jeffries,
 1592 1966; Grant & Hurd, 1979; Rauh, 1979; Kiesling, 1982; Jeffries, 1991). It is not yet known
 1593 whether the stamen movement influences the floral longevity in other species of the genus
 1594 *Parodia* and in other genera. The function of thigmonastic stamen movements is still not clearly
 1595 understood. In species of *Opuntia* it was shown that stimulated thigmonastic stamens form a
 1596 pollen chamber that leads to that effective pollinators touch the stigma to access the hidden
 1597 pollen (Schlindwein & Wittmann, 1997), promoting pollen deposition on the pollinator body
 1598 (Lenzi & Orth, 2011) and that triggering of the stamen movements is closely related to the
 1599 insect body size (Cota-Sánchez *et al.*, 2013). This is similar to *P. neohorstii*, where stamen
 1600 movements result in pollen partitioning that limit the amount of pollen removal per floral visit
 1601 and form a mechanical barrier that prevents unspecialized bees to access the hidden pollen
 1602 reservoir (Cerceau *et al.*, 2019).

1603 However, our results point to a new function for stamen movements. The induction of
 1604 floral senescence after successive stamen movements seems to signal to the flower that the
 1605 pollinators visited the flowers and pollen was exported.

1606 These results bring new hypotheses for future research to elucidate the complex
 1607 functioning of this mechanism and clarify how it evolves in cacti lineage, once stamen
 1608 movement occurs in 2 subfamilies (Jeffries, 1966; Grant & Hurd, 1979; Rauh, 1979; Kiesling,

1609 1982; Jeffries, 1991). In *P. neohorstii*, the induction of floral senescence after visits of
1610 oligolectic pollinators may be related to the highly specialization in plant-pollinator interaction,
1611 however, further studies are needed to clarify these hypotheses.

1612

1613 **ACKNOWLEDGEMENTS**

1614

1615 We thank Samuel Siriani and Ana Laura Dutra for their help on the field experiments,
1616 Reísla Oliveira for help in discussing the results of study, Ana Luísa Cordeiro for schematic
1617 drawing of stamen movements. We thank Instituto Chico Mendes de Conservação da
1618 Biodiversidade (ICMBio) for the licence to study the both red-list species (licence number
1619 57684-1).

1620

1621 **REFERENCES**

1622

1623 **Abdala-Roberts L, Parra-Tabla V, Navarro J.** 2007. Is floral longevity influenced by
 1624 reproductive costs and pollination success in *Cohniella ascendens* (Orchidaceae)? *Annals of*
 1625 *Botany* **100**: 1367-1371.

1626 **Ahmad SS, Tahir I.** 2016. How and why of flower senescence: understanding from models to
 1627 ornamentals. *Indian Journal of Plant Physiology* **21**: 446-456.

1628 **Angulo DF, Sosa V, García-Franco JG.** 2014. Floral movements: stamen motion in *Berberis*
 1629 *trifoliolata*. *Botanical Sciences* **92**: 141-144.

1630 **Arathi H, Rasch A, Cox C, Kelly K.** 2002. Autogamy and Floral Longevity in *Mimulus*
 1631 *guttatus*. *International Journal of Plant Sciences* **163**: 567-573.

1632 **Araujo FF, Oliveira R, Mota T, Stehmann JR, Schlindwein C.** 2020. Solitary bee pollinators
 1633 adjust pollen foraging to the unpredictable flower opening of a species of *Petunia*
 1634 (Solanaceae). *Biological Journal of the Linnean Society* **129**: 273-287.

1635 **Ashman TL, Schoen D.** 1994. How long should flowers live? *Nature* **371**: 788-791.

1636 **Ashman TL, Schoen DJ.** 1996. Floral longevity: fitness consequences and resource costs. pp.
 1637 112-139. In: Lloyd DG, Barrett SCH. *Floral biology*, Springer. 410 pp.

1638 **Aximoff IA, Freitas L.** 2010. Is pollen removal or seed set favoured by flower longevity in a
 1639 hummingbird-pollinated *Salvia* species? *Annals of Botany* **106**: 413-419.

1640 **Barrows EM, Chabot MR, Michener CD, Snyder TP.** 1976. Foraging and mating behavior
 1641 in *Perdita texana* (Hymenoptera: Andrenidae). *Journal of the Kansas Entomological Society*
 1642 **49**: 275-279.

1643 **Blair AC, Wolfe LM.** 2007. The association between floral longevity and pollen removal,
 1644 pollen receipt, and fruit production in flame azalea (*Rhododendron calendulaceum*).
 1645 *Canadian Journal of Botany* **85**: 414-419.

1646 **Castro S, Silveira P, Navarro L.** 2008. Effect of pollination on floral longevity and costs of
 1647 delaying fertilization in the out-crossing *Polygala vayredae* Costa (Polygalaceae). *Annals of*
 1648 *Botany* **102**: 1043-1048.

1649 **Cerceau I, Siriani-Oliveira S, Dutra AL, Oliveira R, Schlindwein C.** 2019. The cost of
 1650 fidelity: foraging oligolectic bees gather huge amounts of pollen in a highly specialized
 1651 cactus-pollinator association. *Biological Journal of the Linnean Society* **128**: 30-43.

- 1652 **Clark MJ, Husband BC. 2007.** Plasticity and timing of flower closure in response to
1653 pollination in *Chamerion angustifolium* (Onagraceae). *International Journal of Plant*
1654 *Sciences* **168**: 619-625.
- 1655 **Córdoba SA, Cocucci AA. 2011.** Flower power: its association with bee power and floral
1656 functional morphology in papilionate legumes. *Annals of botany* **108**: 919-931.
- 1657 **Cota-Sánchez JH, Almeida OJG, Falconer DJ, Choi HJ, Bevan L. 2013.** Intriguing
1658 thigmonastic (sensitive) stamens in the Plains Prickly Pear *Opuntia polyacantha*
1659 (Cactaceae). *Flora - Morphology, Distribution, Functional Ecology of Plants* **208**: 381-389.
- 1660 **Devlin B, Stephenson A. 1984.** Factors that influence the duration of the staminate and
1661 pistillate phases of *Lobelia cardinalis* flowers. *Botanical gazette* **145**: 323-328.
- 1662 **Du W, Qin KZ, Wang XF. 2012.** The mechanism of stamen movement in *Chimonanthus*
1663 *praecox* (Calycanthaceae): differential cell growth rates on the adaxial and abaxial surfaces
1664 of filaments after flower opening. *Plant Systematics and Evolution* **298**: 561-567.
- 1665 **Dutra AL, Schlindwein C, Oliveira R. 2020.** Females of a solitary bee reject males to collect
1666 food for offspring. *Behavioral Ecology* **31**: 884–891
- 1667 **Edwards J, Whitaker D, Klionsky S, Laskowski MJ. 2005.** Botany: a record-breaking pollen
1668 catapult. *Nature* **435**: 164.
- 1669 **Evanhoe L, Galloway LF. 2002.** Floral longevity in *Campanula americana* (Campanulaceae):
1670 a comparison of morphological and functional gender phases. *American Journal of Botan,*
1671 **89**: 587-591.
- 1672 **Grant V, Hurd P. 1979.** Pollination of the southwestern Opuntias. *Plant Systematics and*
1673 *Evolution* **133**: 15-28.
- 1674 **Harder LD, Johnson SD. 2005.** Adaptive plasticity of floral display size in animal-pollinated
1675 plants. *Proceedings of the Royal Society B: Biological Sciences* **272**: 2651-2657.
- 1676 **Henning T, Weigend M. 2012.** Total control - pollen presentation and floral longevity in
1677 Loasaceae (blazing star family) are modulated by light, temperature and pollinator visitation
1678 rates. *PloS One* **7**: e41121.
- 1679 **INMET. 2018.** Banco de Dados Meteorológicos para Ensino e Pesquisa - BDMEP. Dados
1680 históricos da estação meteorológica 83980 - Bagé - RS entre 1988 e 2018.
- 1681 **Ishii H, Sakai S. 2000.** Optimal timing of corolla abscission: experimental study on
1682 *Erythronium japonicum* (Liliaceae). *Functional Ecology*, **14**: 122-128.
- 1683 **Ishii H, Sakai S. 2001.** Effects of display size and position on individual floral longevity in
1684 racemes of *Nartheicum asiaticum* (Liliaceae). *Functional Ecology* **15**: 396-405.

- 1685 **Jeffries L.** 1966. Stamen irritability in Cacti. *The Cactus and Succulent Journal of Great*
 1686 *Britain* **28**: 78-79.
- 1687 **Jeffries L.** 1991. *Pereskopsis* Flowers have Irritable Stamens: Is this a Feature of all
 1688 Opuntioideae? *British Cactus and Succulent Journal* **9**: 5.
- 1689 **Kiesling R.** 1982. The genus *Pterocactus*. *The Cactus and Succulent Journal of Great Britain*,
 1690 **44**: 51-56.
- 1691 **Lenzi M, Orth AI.** 2011. Visitantes florais de *Opuntia monacantha* (Cactaceae) em originais
 1692 de Florianópolis, SC, Brasil. *Acta Biológica Paranaense* **40**: 19-32.
- 1693 **Maluf JR.** 2000. Nova classificação climática do Estado do Rio Grande do Sul. *Revista*
 1694 *Brasileira de Agrometeorologia* **8**: 141-150.
- 1695 **Martini P, Schlindwein C, Montenegro A.** 2003. Pollination, flower longevity, and
 1696 reproductive biology of *Gongora quinquenervis* Ruíz and Pavón (Orchidaceae) in an
 1697 Atlantic forest fragment of Pernambuco, Brazil. *Plant Biology* **5**: 495-503.
- 1698 **Niu Y, Yang Y, Zhang ZQ, Li ZM, Sun H.** 2011. Floral closure induced by pollination in
 1699 gynodioecious *Cyananthus delavayi* (Campanulaceae): effects of pollen load and type, floral
 1700 morph and fitness consequences. *Annals of Botany* **108**: 1257-1268.
- 1701 **Oliveira R.** 2015. Estratégias de acasalamento em abelhas solitárias no Brasil: o que os machos
 1702 fazer para se acasalar? pp. 439-457. In: **Aguiar AJC, Gonçalves RB, Ramos KS** eds.
 1703 *Ensaio sobre as abelhas da Região Neotropical*. Curitiba: Editora UFPR. 456 pp.
- 1704 **Osborn MM, Kevan PG, Lane MA.** 1988. Pollination biology of *Opuntia polyacantha* and
 1705 *Opuntia phaeacantha* (Cactaceae) in southern Colorado. *Plant Systematics and Evolution*
 1706 **159**: 85-94.
- 1707 **Primack RB.** 1985. Longevity of individual flowers. *Annual review of ecology systematics*, **16**:
 1708 15-37.
- 1709 **Proctor HC, Harder LD.** 1995. Effect of pollination success on floral longevity in the orchid
 1710 *Calypso bulbosa* (Orchidaceae). *American Journal of Botany* **82**: 1131-1136.
- 1711 **Rathcke B, Real L.** 1993. Autogamy and inbreeding depression in mountain laurel, *Kalmia*
 1712 *latifolia* (Ericaceae). *American Journal of Botany* **80**: 143-146.
- 1713 **Rathcke BJ.** 2003. Floral longevity and reproductive assurance: seasonal patterns and an
 1714 experimental test with *Kalmia latifolia* (Ericaceae). *American Journal of Botany* **90**:1328-
 1715 1332.
- 1716 **Rauh W.** 1979. Kakteen an ihren Standorten: unter besonderer Berücksichtigung ihrer.
 1717 Morphologie und Systematik. Berlin, Hamburg: Parey. 230 pp.

- 1718 **Ren M.** 2010. Stamen movements in hermaphroditic flowers: diversity and adaptive
1719 significance. *Journal of Plant Ecology* **34**: 867-875.
- 1720 **Ren MX, Bu ZJ.** 2014. Is there 'anther-anther interference' within a flower? Evidences from
1721 one-by-one stamen movement in an insect-pollinated plant. *PloS One* **9**: e86581.
- 1722 **Richardson TE, Stephenson AG.** 1989. Pollen removal and pollen deposition affect the
1723 duration of the staminate and pistillate phases in *Campanula rapunculoides*. *American*
1724 *Journal of Botany* **76**: 532-538.
- 1725 **Robertson C.** 1925. Heterotropic Bees. *Ecology* **6**: 412-436.
- 1726 **Rogers HJ.** 2006. Programmed cell death in floral organs: how and why do flowers die? *Annals*
1727 *of Botany* **97**: 309-315.
- 1728 **Schindwein C, Wittmann D.** 1995. Specialized solitary bees as effective pollinators of South
1729 Brazilian species of *Notocactus* and *Gymnocalycium* (Cactaceae). *Botanica Acta*, **13**: 25-34.
- 1730 **Schindwein C, Wittmann D.** 1997a. Micro-foraging routes of *Bicolletes pampeana*
1731 (Colletidae) and bee-induced pollen presentation in *Cajophora arechavaletae* (Loasaceae).
1732 *Botanica Acta* **110**: 177-183.
- 1733 **Schindwein C, Wittmann D.** 1997b. Stamen movements in flowers of *Opuntia* (Cactaceae)
1734 favour oligolectic pollinators. *Plant Systematics and Evolution* **204**: 179-193.
- 1735 **Schindwein C.** 1998. Frequent oligolecty characterizing a diverse bee–plant community in a
1736 xerophytic bushland of subtropical Brazil. *Studies on Neotropical Fauna and Environment*
1737 **33**: 46-59.
- 1738 **Schindwein C, Westerkamp C, Carvalho AT, Milet-Pinheiro P.** 2014. Visual signalling of
1739 nectar-offering flowers and specific morphological traits favour robust bee pollinators in the
1740 mass-flowering tree *Handroanthus impetiginosus* (Bignoniaceae). *Botanical journal of the*
1741 *Linnean Society* **176**: 396-407.
- 1742 **Sibaoka T.** 1969. Physiology of rapid movements in higher plants. *Annual Review of Plant*
1743 *Physiology* **20**: 165-184.
- 1744 **Simons PJNP.** 1981. The role of electricity in plant movements. *New Phytologist* **87**: 11-37.
- 1745 **Siriani-Oliveira S, Oliveira R, Schindwein C.** 2018. Pollination of *Blumenbachia amana*
1746 (Loasaceae): flower morphology and partitioned pollen presentation guarantee a private
1747 reward to a specialist pollinator. *Biological Journal of the Linnean Society*, **124**: 479-491.
- 1748 **Siriani-Oliveira S, Cerceau I, Schindwein C.** 2020. Specialised protagonists in a plant–
1749 pollinator interaction: the pollination of *Blumenbachia insignis* (Loasaceae). *Plant Biology*
1750 **22**: 167-176

- 1751 **Stead A, Moore K.** 1979. Studies on flower longevity in *Digitalis*: The role of ethylene in
1752 corolla abscission. *Planta* **146**: 409-414.
- 1753 **Stead AJPGR.** 1992. Pollination-induced flower senescence: a review. *Plant Growth*
1754 *Regulation* **11**: 13-20.
- 1755 **Teixido AL, Valladares F.** 2015. Temperature-limited floral longevity in the large-flowered
1756 mediterranean shrub *Cistus ladanifer* (Cistaceae). *International Journal of Plant Science*,
1757 **176**: 131-140.
- 1758 **Tripathi SK, Tuteja N.** 2007. Integrated signaling in flower senescence: an overview. *Plant*
1759 *Signaling and Behavior* **2**: 437-445.
- 1760 **van Doorn WG.** 1997. Effects of pollination on floral attraction and longevity. *Journal of*
1761 *Experimental Botan* **48**: 1615-1622.
- 1762

1763 **TABLES**

1764

1765 **Table 1:** Floral visitors of *Parodia neohorstii*, their body volume, duration of flower visits and
 1766 percentage of visits that resulted in stamen movement.

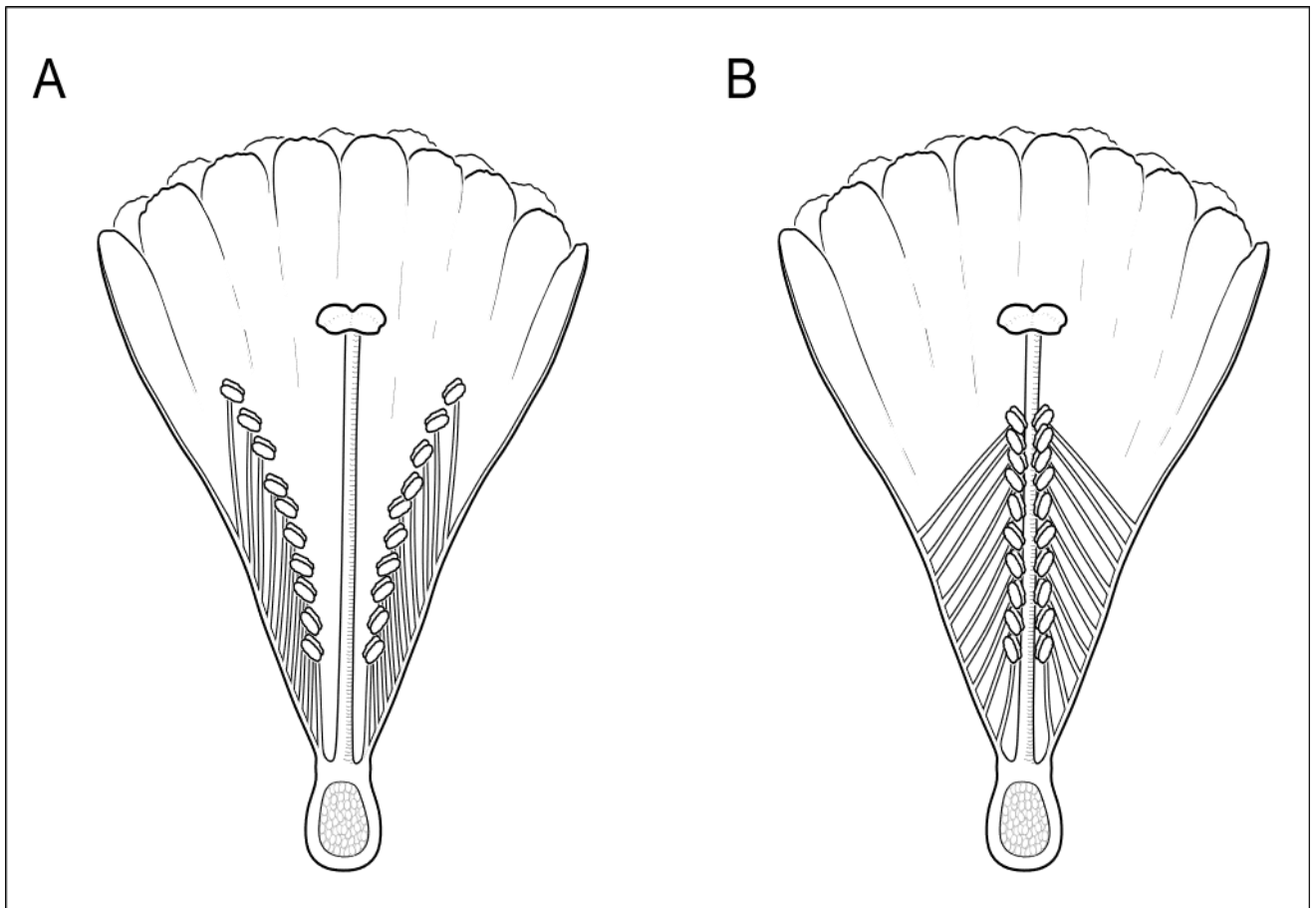
Bee specie	Sex	Body volume (mm³)	Visit duration (s) (N)	% stimuli (N)
<i>Arhysosage cactorum</i> Moure 1999	Female	27.3 ± 3.8	17.9 ± 14.6 (101)	98.0 (50)
	Male	18.8 ± 5.0	2.1 ± 1.2 (41)	23.6 (55)
	Couple	46.1 ± 7.1	13.6 ± 13.44 (103)	100.0 (60)
<i>Anthrenoides micans</i> Urban 1995	Female	15.4 ± 1.8	6.9 ± 5.4 (111)	30.1 (113)
	Male	9.2 ± 4.3	1.2 ± 0.5 (4)	0.0 (16)
	Couple	24.62 ± 2.4	132.6 ± 159.2 (5)	90.9 (11)
<i>Cactocolletes rugatus</i> (Urban)	Female	52.1 ± 6.4	19.0 ± 1.8 (4)	100 (4)
<i>Dialictus sp.</i>	Female	4.9 ± 0.9	4.1 ± 4.5 (71)	0.0 (59)

1767

1768

1769 FIGURES

1770



1771

1772 **Figure 1:** Flower of *Parodia neohorstii*, longitudinal section, schematic drawing of stamen movements.

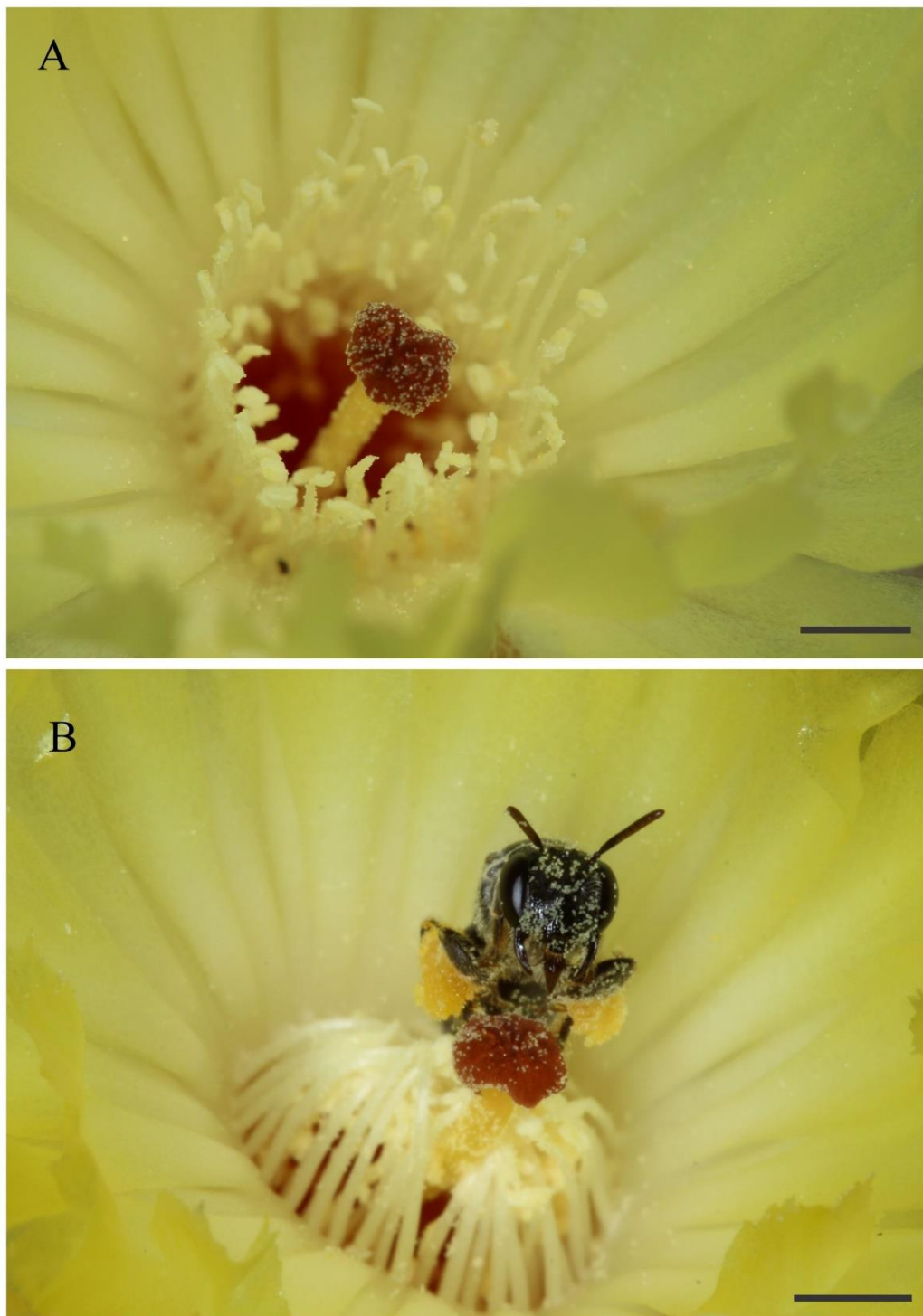
1773 (A) Unstimulated stamens in the original position, sprawled erect; anthers, flower base and style are free

1774 accessible; (B) Stamens in the stimulated position. After mechanical stimulation, stamens bend to the

1775 center of the flower and lean to the style. Only anthers of the uppermost stamens are accessible to flower

1776 visitors. After four to five minutes, the stamens bend back to the original position.

1777



1778

1779

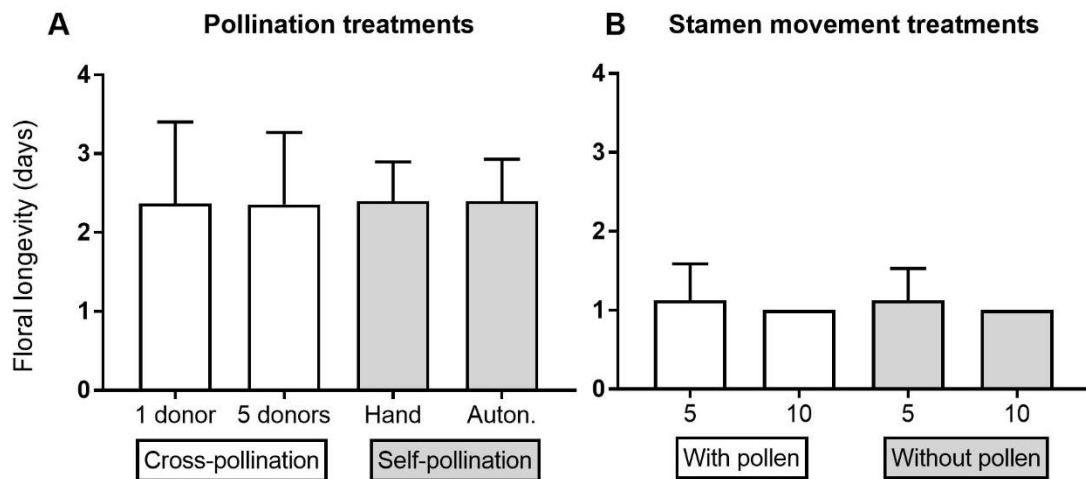
1780

1781

1782

1783

Figure 2: Flowers of *Parodia neohorstii* before and after flower visits of bees. (A) Flower with the stamens in original position before bee visits. The free style and the dark red to violet inner floral tube are visible. (B) Flower with stimulated stamens after bee visits. Stamens lean to the style and hinder the access to the anthers of lower stamens. A pollen collecting female of *Anthrenoides micans* is grooming pollen. Scale bars = 2 mm.



1784

1785 **Figure 3:** Floral longevity of *Parodia neohorstii* in different treatments. (A) Pollination treatments.
 1786 Cross-pollination treatments differed in the number of pollen donors: a single conspecific donor (1
 1787 donor) or five different conspecific donors (5 donors). In self-pollination treatments, flowers were hand
 1788 self-pollinated or maintained bagged (autonomous pollination). (B) Stamen stimulation treatments.
 1789 Stamen movements were experimentally stimulated five or ten times in intervals of 5 minutes. The
 1790 flowers were hand-cross pollinated (with pollen from conspecific donors) or not pollinated.

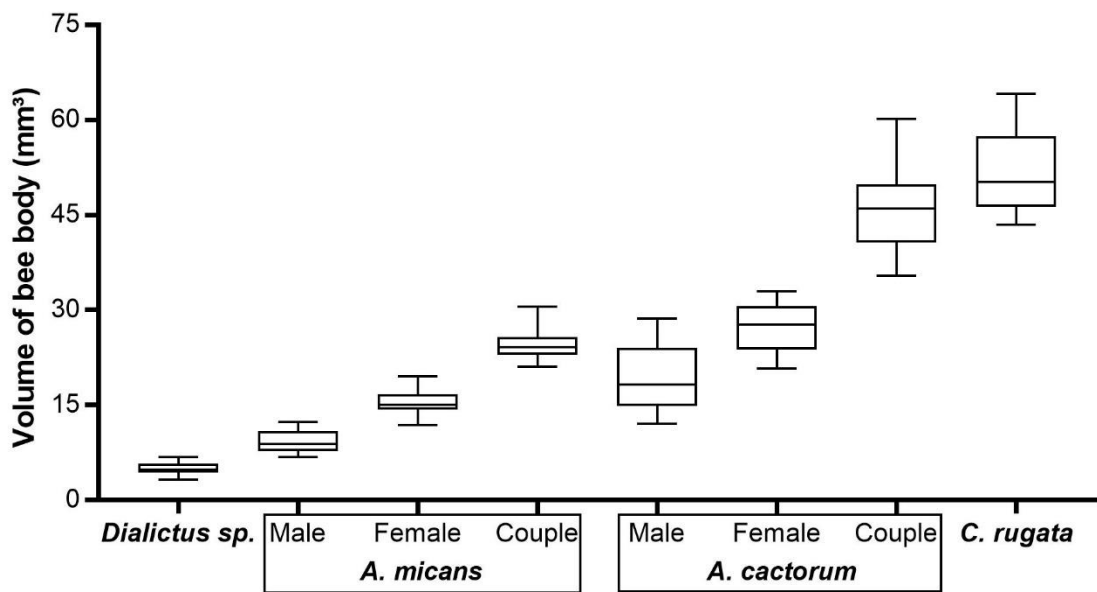
1791



1792

1793 **Figure 4:** Couple of *Arhysosage cactorum* inside a flower of *Parodia neohorstii*. The male clinches to
1794 the female with its mandibles during copulation. The female continues pollen foraging in copula
1795 position. Scale bars = 2 mm.

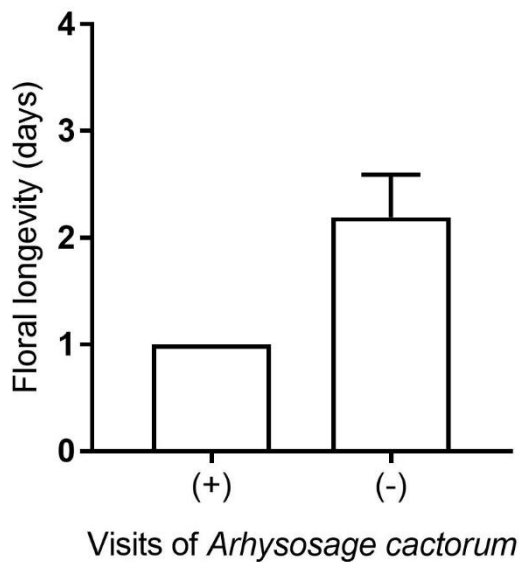
1796



1797

1798 **Figure 5:** Body volume of flower visiting bees of *Parodia neohorstii*. The number of individual bees1799 measured were: *Dialictus sp.* (N = 20 females), *Anthrenoides micans* (N = 20 females and 15 males),1800 *Arhysosage cactorum* (N = 20 females and 20 males) and *Cactocolletes rugatus* (N = 20 females).

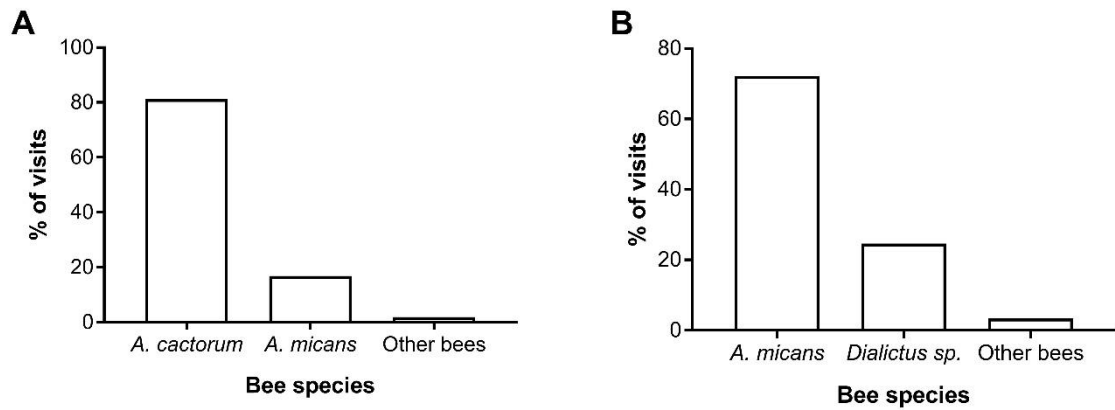
1801



1802

1803 **Figure 6:** Floral longevity of *Parodia neohorstii* under natural conditions, with the presence (+) or1804 absence (-) of the main pollinator *Arhysosage cactorum*.

1805



1806

1807 **Figure 7:** Visiting bees in *Parodia neohorstii* flowers in two different time scenarios. (A) Frequency of
1808 bee visitation during the peak of flowering, where the bee specie *A. cactorum* is the dominant floral
1809 visitor. (B) Frequency of bee visitation during the beginning of the flowering period, where *A. cactorum*
1810 was still inactive.

1811

1812

CAPÍTULO 3

1813

1814

1815

1816

1817

1818

1819

1820

1821

**AMEAÇAS À CONSERVAÇÃO DE *PARODIA NEOHORSTII* (CACTACEAE),
ESPÉCIE CRITICAMENTE AMEAÇADA DE EXTINÇÃO**

1822 **AMEAÇAS À CONSERVAÇÃO DE *PARODIA NEOHORSTII* (CACTACEAE),**
1823 **ESPÉCIE CRITICAMENTE AMEAÇADA DE EXTINÇÃO**

1824
1825 **RESUMO**

1826
1827 Dentre as 1450 espécies da família Cactaceae, aproximadamente um terço é ameaçada
1828 de extinção. A pressão antropogênica é o principal fator de ameaça às populações naturais, uma
1829 vez que áreas de ocorrência dos cactos podem ser convertidas em áreas de pastagem e
1830 agricultura. A espécie *Parodia neohorstii* é classificada como criticamente ameaçada e ocorre
1831 em afloramentos de arenito no Pampa do Rio Grande do Sul. Com o objetivo de identificar as
1832 ameaças à conservação dessa espécie, percorremos as áreas de ocorrência, quantificamos as
1833 populações encontradas e classificamos os indivíduos quanto ao tamanho, fase reprodutiva e
1834 integridade das plantas. Contabilizamos 3677 indivíduos em 9 localidades, sendo 1225 juvenis
1835 e 2452 reprodutivos. Os cactos iniciaram a fase reprodutiva ao atingirem 25.3 mm de altura e a
1836 produção de flores está positivamente correlacionada com a altura dos indivíduos. Dos cactos
1837 avaliados, 16.9% foram impactados por animais de criação. Os impactos corresponderam ao
1838 deslocamento dos indivíduos do substrato e a danos físicos causados por mordidas e por
1839 pisoteio. Cabras e gado foram os principais animais de criação observados nas áreas de
1840 ocorrência dos cactos. As cabras foram os principais agentes responsáveis pelo consumo direto
1841 dos indivíduos de *P. neohorstii* enquanto o gado foi responsável pelo pisoteio. Os cactos
1842 impactados apresentaram taxa de mortalidade de 41% e os indivíduos reprodutivos foram sete
1843 vezes mais impactados quando comparado a juvenis. Dos 9 afloramentos avaliados, 4 estão
1844 localizados dentro de áreas de pastagem. Nos afloramentos de mais fácil acesso aos animais, as
1845 populações de *P. neohorstii* apresentaram maior proporção de cactos danificados e mortos, além
1846 de menor representatividade de indivíduos jovens, quando comparadas aos locais de difícil
1847 acesso. Concluimos que medidas de contenção de animais domésticos nos locais de ocorrência
1848 das populações de *P. neohorstii* é primordial para garantir a manutenção da espécie.

1849
1850 **Palavras-chave:** Cactaceae, ameaças à conservação, impacto de animais, levantamento
1851 populacional

1852

1853

1854

1855 INTRODUÇÃO

1856

1857 A família Cactaceae é representada por cerca de 1450 espécies, distribuídas em diversos
1858 habitats nas Américas (Anderson, 2001; Hunt *et al.*, 2006, 2013). Aproximadamente um terço
1859 das espécies é classificada em algum grau de ameaça de extinção (Goettsch *et al.*, 2015). A
1860 pressão antropogênica é a maior ameaça aos representantes da família e causa destruição dos
1861 habitats, principalmente devido a expansão urbana e agropecuária (IUCN, 2020).
1862 Características intrínsecas das cactáceas, como altas taxas de mortalidade na fase juvenil,
1863 requerimentos específicos de habitat e distribuição restrita tornam as espécies vulneráveis a
1864 mudanças no habitat (Godínez-Álvarez *et al.*, 2003). Diversas espécies de cactos globosos
1865 ameaçados de extinção estão em declínio populacional, o que significa que a exposição
1866 continuada a fatores de risco agrava ainda mais o declínio das populações naturais (Mandajuno
1867 *et al.*, 2015; Martínez-Ramos *et al.*, 2016; Anceschi & Magli, 2018).

1868 A criação de animais em áreas de ocorrência dos cactos impacta negativamente muitas
1869 populações devido ao pisoteio por bovinos e consumo de indivíduos por caprinos (Schlosser,
1870 1979; Machado *et al.*, 2005; McIntosh *et al.*, 2020). Além disso, os animais podem remover as
1871 plantas que agem como facilitadoras na germinação e estabelecimento de cactos (Bowers,
1872 1997). A germinação de sementes e sobrevivência de indivíduos juvenis normalmente está
1873 relacionada com microhabitats específicos ou interações de facilitação com plantas ou objetos,
1874 por exemplo rochas (Godínez-Alvarez *et al.*, 2003; Saraiva & Souza 2012), que promovem
1875 condições favoráveis para o desenvolvimento inicial das plântulas (Gurvich *et al.*, 2017).

1876 Em Cactaceae, de uma maneira geral, somente uma pequena proporção dos indivíduos
1877 jovens atinge a fase reprodutiva e contribui para o crescimento populacional através da
1878 produção de frutos e sementes (Godínez-Alvarez *et al.*, 2003). Simulações de dinâmica
1879 populacional de cactos demonstraram que pequenas modificações na estrutura populacional,
1880 como remoção de indivíduos ou o aumento da mortalidade em determinada faixa de tamanho
1881 podem gerar impactos negativos a longo prazo (Mandujano *et al.*, 2015; Martínez-Ramos *et al.*,
1882 2016).

1883 Dessa maneira, é importante identificar as principais ameaças às populações naturais de
1884 cactos para implementar ações de conservação direcionadas para a realidade de cada local.

1885 O Brasil é um dos países com maior riqueza e diversidade de cactáceas, e assim como em várias
1886 regiões das Américas, grande proporção de espécies estão ameaçadas de extinção (Goettsch *et*
1887 *al.*, 2019). Um dos centros de diversidade de cactáceas no Brasil é o estado do Rio Grande do

1888 Sul, onde estão concentrados aproximadamente 30% das espécies de cactos brasileiros
1889 (Carneiro *et al.*, 2016). No estado, cada região apresenta uma composição única das espécies e
1890 os campos do Pampa concentram a maior diversidade (Saraiva *et al.*, 2015).

1891 Para identificar as principais ameaças aos cactos em habitat natural, escolhemos
1892 *Parodia neohorstii* (S.Theun.) N.P.Taylor como modelo. Trata-se de um cacto globoso,
1893 criticamente ameaçado de extinção, com ocorrência restrita em afloramentos areníticos dos
1894 campos do Pampa brasileiro (Larocca & Machado, 2013). Identificamos as principais ameaças
1895 e analisamos os impactos causados pelos animais de criação (gado e cabras) nas populações de
1896 *P. neohorstii*. Perguntamos: (1) Quais são os principais danos causados por animais de criação
1897 nas populações naturais de *P. neohorstii*? (2) Qual a taxa de mortalidade de indivíduos em
1898 habitat natural?

1899 Para responder a essas perguntas, percorremos afloramentos rochosos em busca de *P.*
1900 *neohorstii* e avaliamos detalhadamente as populações encontradas. Em cada localidade,
1901 classificamos todos os indivíduos em faixas de tamanho, fase reprodutiva e verificamos a
1902 integridade do corpo dos cactos. Além disso, avaliamos a acessibilidade de animais de criação
1903 ao local de ocorrência de cada população.

1904

1905 **MATERIAIS E MÉTODOS**

1906

1907 *Área de estudo*

1908

1909 O estudo foi conduzido na Serra do Sudeste, entre os municípios de Santana da Boa
1910 Vista e Caçapava do Sul, no sul do Rio Grande do Sul (30°53'17.1"S 53°24'03.6"W). A região
1911 faz parte do bioma Pampa, caracterizado pela fitofisionomia de campos arbustivos, onde há
1912 abundância de afloramentos rochosos conhecidos como Guaritas, encontrados em meio a uma
1913 matriz vegetacional. A vegetação é predominantemente herbácea e arbustiva, com ocorrência
1914 de diversas espécies de cactáceas ameaçadas de extinção (Carneiro *et al.*, 2016). O clima é
1915 subtemperado (Maluf *et al.*, 2000) com média de precipitação anual de 1509 mm e média de
1916 temperatura anual de 18.7 °C. No inverno ocorrem geadas esporádicas (INMET, 2018). As
1917 expedições de campo foram realizadas entre outubro e dezembro de 2017 e 2018.

1918

1919 *Espécie estudada*

1920

1921 *Parodia neohorstii* é uma espécie de cacto globoso e solitário, endêmico da região das
1922 Guaritas. A espécie ocorre em áreas pedregosas expostas no topo de afloramentos de arenito e
1923 conglomerado (Figura 1). Antes do estudo, apenas duas populações eram conhecidas. A espécie
1924 floresce entre o final de outubro e início de dezembro. A antese dura normalmente apenas três
1925 horas na presença dos polinizadas. Os polinizadores são abelhas solitárias, que visitam
1926 exclusivamente flores de Cactaceae (Schlindwein, 1998; Cerceau *et al.*, 2019).

1927

1928 *Definição de população*

1929

1930 Neste estudo, as populações foram definidas em conformidade com o proposto por
1931 Pianka (1978): grupo de indivíduos que ocorrem em uma área geográfica e que tem maior
1932 probabilidade de reproduzirem entre si do que com indivíduos fora desta área de ocorrência.

1933 De acordo com Cerceau *et al.*, (2019), *P. neohorstii* é uma espécie autoincompatível e
1934 dependente de abelhas especialistas para se reproduzir. A floração sincronizada e a antese de
1935 apenas 3 horas promovem a intensa visitação por abelhas e polinização efetiva em curto
1936 intervalo de tempo. Individualmente, as abelhas polinizadoras são féis às flores dos cactos que
1937 visitam. Estudos recentes demonstraram que abelhas marcadas são frequentes nas mesmas
1938 flores durante toda a antese (Dutra, 2015; Dutra *et al.*, 2020). Devido ao fato de *P. neohorstii*
1939 ocorrer em afloramentos rochosos ilhados em meio à vegetação, há maior probabilidade de
1940 fluxo polínico entre indivíduos do mesmo afloramento. Dessa maneira, consideramos que cada
1941 afloramento abriga uma população.

1942

1943 *Produção de flores e tamanho reprodutivo*

1944

1945 Para avaliar a produção de flores por indivíduos de *P. neohorstii*, medimos a altura de
1946 661 indivíduos com paquímetro digital e contabilizamos o número de flores produzidas por
1947 cada indivíduo durante a estação reprodutiva. A partir desses dados, identificamos o tamanho
1948 mínimo dos indivíduos reprodutivos. Verificamos se existe correlação entre da altura do
1949 indivíduo com o número de flores produzidas durante a estação reprodutiva.

1950

1951 *Classificação dos indivíduos em faixas de tamanho*

1952

1953 A partir dos dados de correlação entre a produção de flores e altura dos indivíduos,
 1954 definimos oito faixas de tamanho, sendo elas: (1) 0-25 mm; (2) 25-50 mm; (3) 50-75 mm; (4)
 1955 75-100 mm; (5) 100-125 mm; (6) 125-150mm; (7) 150-175 mm; (8) 175-200 mm.
 1956 Confeccionamos um instrumento de medição rápida de altura, composto por um bastão de
 1957 bambu com códigos de cores. Cada cor correspondeu a uma faixa altura pré-definida (Figura
 1958 2).

1959

1960 *Levantamento populacional e as principais ameaças a Parodia neohorstii*

1961

1962 Para ampliar o conhecimento sobre a distribuição da espécie, percorremos alguns
 1963 afloramentos de arenito da região em busca de novas populações de *P. neohorstii*. Ao encontra-
 1964 las, classificamos todos os indivíduos em: 1) juvenil ou reprodutivo; 2) faixa de tamanho
 1965 (altura); 3) morto ou vivo; 4) íntegro ou impactado por animais. Os dois principais impactos
 1966 causados por animais foram: i) remoção do cacto do substrato e ii) danos físicos. A remoção
 1967 dos indivíduos do substrato ocorre após a passagem de animais, o que ocasiona no
 1968 desprendimento das raízes do substrato e deslocamento do cacto do local de estabelecimento.
 1969 Os danos físicos corresponderam a perfurações no corpo dos cactos causados por mordidas ou
 1970 pisoteio por animais. Os indivíduos com danos físicos foram encontrados amassados, com
 1971 espinhos quebrados, marcas de dentes ou grande parte do parênquima exposto.

1972 Classificamos as áreas de acordo com a acessibilidade de animais de criação: 1)
 1973 acessibilidade alta, 2) acessibilidade média, e 3) acessibilidade baixa. As áreas de alta
 1974 acessibilidade foram consideradas aquelas onde os afloramentos estão situados dentro de uma
 1975 área de pastagem e de fácil acesso aos animais de criação (gado e cabras). Áreas de média
 1976 acessibilidade corresponderam àquelas parcialmente cercadas, impedindo o acesso de gado,
 1977 porém, com acesso de cabras pelas áreas íngremes, não cercadas, dos afloramentos. Os
 1978 afloramentos de baixa acessibilidade foram aqueles muito íngremes, considerados inacessíveis
 1979 a animais de criação.

1980 Avaliamos a proporção de indivíduos jovens nas populações para inferir o índice de
 1981 regeneração, uma vez que a presença de indivíduos na primeira faixa de tamanho (0-25 mm)
 1982 indica que o recrutamento de novos indivíduos está ocorrendo através da germinação de
 1983 sementes. Para isso calculamos a proporção de cactos no primeiro estágio de tamanho (0-25
 1984 mm) em relação a quantidade total de indivíduos em cada população.

1985 Através de imagens de alta resolução espacial do Google Earth, medimos a área total de
 1986 cada afloramento, bem como a área ocupada pelas populações dentro de cada afloramento.
 1987 Através desses dados, calculamos a densidade de cactos por área de ocorrência (nº de indivíduos
 1988 / área). Além disso, registramos a altitude dos pontos de ocorrência das populações.

1989

1990 *Análises estatísticas*

1991

1992 Para verificar se existe correlação entre altura dos cactos e quantidade de flores
 1993 produzidas por estação, utilizamos o teste de Pearson.

1994

1995 **RESULTADOS**

1996

1997 *Classificação dos indivíduos em faixas de tamanho*

1998

1999 Os cactos começaram a produzir flores a partir de 25.3 mm de altura (N = 661).
 2000 Identificamos 343 cactos reprodutivos e verificamos que quantidade de flores produzidas por
 2001 indivíduo está positivamente correlacionada com a altura dos cactos ($r = 0.7475$, $P < 0.0001$, N
 2002 = 661, Figura 3). Os cactos produziram em média 4.1 ± 3.4 flores por ano (N= 343) e apenas
 2003 6% (22 indivíduos) produziram mais de 10 flores e apenas 1% (4 indivíduos) produziram 20
 2004 flores ou mais flores por ano, sendo o máximo de 23 flores.

2005

2006 *Levantamento populacional de Parodia neohorstii*

2007

2008 Encontramos 3677 indivíduos distribuídos em 9 populações, sendo 3428 indivíduos
 2009 vivos. Todas as ocorrências foram em afloramentos de arenito conglomeráticos com grânulos
 2010 e seixos pequenos a médios, em altitudes variando de 147 a 300 metros (Tabela 1).

2011 Os cactos ocorreram em áreas restritas dos afloramentos, apenas em áreas com substrato com
 2012 grânulos e seixos, uma vez que fixam as raízes entre rochas e frestas. A área ocupada pelas
 2013 populações varia entre 4.4 a 23.7% da área total do afloramento, o que corresponde a uma área
 2014 de ocupação entre 184 a 12890 m². A quantidade de indivíduos nas populações variou entre 93
 2015 até 1056 (Tabela 1). A maioria das populações é encontrada em áreas particulares, com exceção
 2016 de uma população que ocorre em um afloramento às margens de uma estrada não asfaltada.
 2017 Cinco populações tem acessibilidade alta para animais de criação, onde os afloramentos

2018 ocorreram dentro de áreas de pastagem e à beira de estrada. Três populações tem acessibilidade
 2019 média, onde eram parcialmente cercadas e impossibilitavam a passagem do gado, porém,
 2020 caprinos acessavam percorrendo áreas íngremes não cercadas do afloramento (Figura 4).
 2021 Apenas uma população é inacessível aos animais devido ao afloramento ser muito íngreme
 2022 (Tabela 1).

2023 A estrutura populacional variou entre as localidades (Figura 5). Indivíduos juvenis, com
 2024 até 25mm de altura foram os mais representativos (32.1%; 1181 indivíduos). O índice de
 2025 regeneração, que representa a proporção de indivíduos nesta faixa de tamanho nas populações,
 2026 variou de 9.6 a 82% e foi menor onde houve maior acesso de animais (Tabela 1). A população
 2027 sem acesso aos animais foi caracterizada principalmente pela abundância de indivíduos juvenis
 2028 e ausência de cactos nas maiores faixas de tamanho (Figura 5.C).

2029 Apenas 26 indivíduos foram encontrados na maior faixa de tamanho (175-200 mm)
 2030 (0.7%) e apresentaram a maior taxa de mortalidade (26.9%). Indivíduos mortos ocorreram
 2031 principalmente nas populações com maior quantidade de cactos.

2032

2033 *Estado de conservação e principais ameaças das populações de Parodia neohorstii*

2034

2035 Dentre os 3677 indivíduos avaliados em todas as populações, 83.1% estavam íntegros e
 2036 16.9% foram impactados por animais (Figura 6). Os impactos por animais se revelaram tanto
 2037 por danos físicos em 8.2% dos cactos, que apresentaram marcas de mordidas e de pisoteio,
 2038 quanto por deslocamento de 8.7% dos cactos do substrato.

2039 Adultos e filhotes de cabras removeram os cactos do substrato com as patas dianteiras
 2040 e se alimentaram tanto do sistema radicular quanto do parênquima dos cactos (Figura 7). Esses
 2041 danos foram aproximadamente sete vezes mais frequentes em indivíduos reprodutivos quando
 2042 comparado a indivíduos juvenis (Tabela 2).

2043 Populações com alta e média acessibilidade aos animais apresentaram maior proporção
 2044 de indivíduos com danos e indivíduos mortos (Tabela 2). Dentre os indivíduos avaliados, 249
 2045 estavam mortos (6.7%). A faixa de tamanho com maior proporção de indivíduos impactados e
 2046 mortos correspondeu aos indivíduos maiores, com altura entre 175-200 mm (Figura 8). A maior
 2047 causa da morte em todas as faixas de tamanho foram os danos físicos causados por mordidas e
 2048 pisoteio (Figura 8B). Dos cactos que apresentaram esse tipo de dano, 73% estavam mortos.

2049 Apenas 5 indivíduos íntegros estavam mortos e apresentaram o corpo seco sem nenhum
 2050 dano ou deslocamento, que representou apenas 0.2% dos indivíduos amostrados.

2051

2052 **DISCUSSÃO**

2053

2054 Nosso estudo revelou que a principal ameaça à espécie *P. neohorstii* é o impacto direto
2055 causado por animais de criação, principalmente cabras e gado. As áreas de fácil acesso a esses
2056 animais apresentaram maior proporção de cactos danificados e mortos, além de menor
2057 representatividade de indivíduos jovens.

2058

2059 *Principais ameaças aos cactos*

2060

2061 Os danos causados pelos animais de criação impactaram 16.9% dos indivíduos
2062 avaliados neste estudo. Das populações avaliadas, 88% foram acessíveis a animais de criação,
2063 sendo que metade delas ocorreram dentro de áreas de pastagem, o que aumenta a probabilidade
2064 de pisoteio e o consumo dos cactos. Reportamos que cabras se alimentam do parênquima de *P.*
2065 *neohorstii*. Esse tipo de comportamento também foi reportado nos Estados Unidos a espécie de
2066 carneiro selvagem (*Ovis canadensis*), que consome o conteúdo interno de *Ferocactus sp.* após
2067 quebrar e retirar o excesso de espinhos com as patas (Warrick & Krausman, 1989). A exposição
2068 do parênquima dos cactos, tanto devido a mordidas quanto ao pisoteio, leva ao ressecamento e
2069 a morte do indivíduo. Em *P. neohorstii*, 73% dos indivíduos que apresentaram perfurações no
2070 corpo estavam mortos. O ressecamento após a herbivoria foi identificado como uma das
2071 maiores causas da morte do cacto globoso *Astrophytum capricorne* (Mandujano *et al.*, 2015).
2072 Outros mamíferos herbívoros, como coelhos e esquilos, se alimentam de *Astrophytum asterias*
2073 e causam altas taxas de mortalidade (Ferguson *et al.*, 2013). Nas populações de *P. neohorstii*
2074 não observamos danos causados por animais nativos.

2075

2076 O consumo de cactos por cabras e o pisoteio por gado também foram relatados para
2077 outras espécies do gênero *Parodia* (Anceschi & Magli, 2018) e para outros gêneros como
2078 *Discocactus* (Machado *et al.*, 2005), *Frailea* (Machado, 2007), *Gymnocalycium* (Schlosser,
2079 1979) e *Echinocactus* (Jiménez-Sierra *et al.*, 2007). Além dos danos físicos, os indivíduos
2080 foram frequentemente removidos do afloramento rochoso pelo impacto causado pela passagem
2081 dos animais, sem apresentarem perfurações no corpo. Após o deslocamento do substrato, as
2082 raízes foram direcionadas para cima ou para os lados, impossibilitando o enraizamento. Vale
2083 ressaltar a resiliência de alguns indivíduos, pois o deslocamento causou a mortalidade de menos
de 8% dos cactos e a maioria dos indivíduos deslocados continuaram florescendo.

2084 As populações de *P. neohorstii* onde a acessibilidade de animais é mais fácil, reportamos
2085 maior quantidade de indivíduos impactados e menor quantidade de indivíduos juvenis. Esse
2086 fato pode sugerir que caprinos se alimentam de indivíduos jovens sem deixar vestígios, uma
2087 vez que eles possuem menor quantidade de espinhos e menor tamanho corporal, o que
2088 possibilita a ingestão completa do cacto. Porém, estudos de acompanhamento de cactos juvenis
2089 marcados devem ser realizados ao longo dos anos para avaliar o real impacto real de caprinos
2090 nas persistência de indivíduos nas menores faixas de tamanho. Na área de estudo, observamos
2091 cabras ingerindo indivíduos inteiros da espécie *Frailea gracillima*, uma espécie de cacto
2092 pequeno e com espinhos macios. Dados semelhantes foram registrados no México para cactos
2093 do gênero *Astrophytum* onde os indivíduos das menores faixas de tamanho foram mais
2094 impactados pelo consumo de animais (Martínez-Ávalos *et al.*, 2007; Ferguson *et al.*, 2013).
2095 Estudos populacionais demonstram que a mortalidade afeta negativamente o equilíbrio das
2096 populações (Mandujano *et al.*, 2015), e as flutuações na sobrevivência de indivíduos nos
2097 primeiros estágios de vida impactam a população a longo prazo (Bowers, 1997; Valverde *et al.*,
2098 2004).

2099 Outras ameaças aos cactos do Rio Grande do Sul são conversão de áreas para a plantação
2100 de *Eucalyptus*, que ameaça de pelo menos 27 espécies (Goettsch *et al.*, 2015; Carneiro *et al.*,
2101 2016; Anceschi & Magli, 2018) e a coleta ilegal (Ribeiro-Silva *et al.*, 2011).
2102 *Parodia neohorstii* não é impactada diretamente pela conversão do habitat para plantações uma
2103 vez que ocorre em afloramentos rochosos, porém, outras espécies simpátricas são impactadas.
2104 Não observamos indícios de coleta de indivíduos de *Parodia neohorstii* para finalidade
2105 ornamental, mas é uma grande ameaça a diversas espécies de *Parodia* da região (Ribeiro-Silva
2106 *et al.*, 2011).

2107

2108 *Habitat de Parodia neohorstii*

2109

2110 As populações de *P. neohorstii* ocorrem em áreas restritas dos afloramentos de arenito
2111 associados a blocos de conglomerados. A ocorrência em habitats específicos é uma
2112 característica de espécies de *Parodia*, que resulta em distribuição de populações isoladas
2113 (Anceschi & Magli, 2018). Diversos cactos utilizam agentes facilitadores como plantas ou
2114 fendas de rochas ou pedras menores para germinar e se estabelecer (Godínez-Álvarez *et al.*,
2115 2003; Peters *et al.*, 2008; Zenteno Ruíz *et al.*, 2009; Martínez-Ramos *et al.*, 2016). O
2116 crescimento à sombra de pequenas pedras pode ser auxiliado por uma redução da radiação solar

2117 e sombreamento, além da umidade prolongada no solo (Larmuth & Harvey, 1978), criando
2118 condições microclimáticas favoráveis para a germinação e desenvolvimento inicial de
2119 plântulas. A ocorrência de *P. neohorstii* parece não estar relacionada com a presença de plantas
2120 facilitadoras, porém, a presença de conglomerados no substrato pode auxiliar a fixação das
2121 raízes onde há pequeno acúmulo de solo. Os afloramentos são muito heterogêneos,
2122 apresentando porções com dominância de arbustos, bromélias e musgos, além de áreas com
2123 rocha lisa de arenito exposto, sem conglomerados. Nessas áreas sem a presença dos
2124 conglomerados que podem auxiliar na fixação das raízes, não registramos a presença de
2125 *Parodia neohorstii*.

2126

2127 *Reprodução de Parodia neohorstii*

2128

2129 O início da idade reprodutiva varia entre as espécies de cactáceas (Godínez-Álvarez *et*
2130 *al.*, 2003). Indivíduos de *Parodia neohorstii* começaram a produzir flores ao atingirem 25,3
2131 mm de altura. Com o aumento do tamanho do indivíduo, a quantidade de flores produzidas
2132 aumentou. Esses resultados foram consistentes com aqueles encontrados para outras espécies
2133 de cactos (Godínez-Álvarez *et al.*, 2003; Mandujano *et al.*, 2015; Martínez-Ramos *et al.*, 2016).

2134 A produção de sementes é um fator crítico na manutenção de diversas espécies de
2135 cactáceas, uma vez que muitas espécies não se reproduzem vegetativamente (Godínez-Álvarez
2136 *et al.*, 2003). A baixa produção de sementes viáveis pode reduzir o número de novos indivíduos
2137 na população e levar ao declínio populacional (Esparza-Olguín *et al.*, 2005). Projeções
2138 populacionais revelaram que apenas um pequeno aumento na germinação de sementes afeta
2139 positivamente a população de cactos a longo prazo (Mandujano *et al.*, 2015). *Parodia neohorstii*
2140 não se propaga vegetativamente e depende de abelhas especializadas para formar frutos
2141 (Cerceanu *et al.*, 2019). As sementes de *P. neohorstii* coletadas em campo não apresentaram
2142 sinais de dormência e a taxa de germinação variou entre 20 – 55% em condições controladas
2143 (Dal Ri não publicado). A germinação de cactos globosos em laboratório é muito variável, e
2144 foram registrados valores entre 12-95% em várias espécies (Mandujano *et al.*, 2015; Martínez-
2145 Ramos *et al.*, 2016; Gurvich *et al.*, 2017). Apesar das taxas de germinação em habitat natural
2146 serem menores do que as observadas em laboratório (Gurvich *et al.*, 2017), a presença de
2147 indivíduos jovens nas populações de *P. neohorstii* sugere que o recrutamento de novos
2148 indivíduos está ocorrendo constantemente através da germinação das sementes. De acordo com
2149 o estudo de polinização (Cerceanu *et al.*, 2019), estimamos que cada indivíduo reprodutivo de

2150 *P. neohorstii* produz em média 250 sementes e os maiores cactos podem produzir até 1250
2151 sementes por ano.

2152

2153 *Populações de Parodia neohorstii*

2154

2155 As estruturas populacionais de *P. neohorstii* sugerem que a maioria das populações está
2156 em constante regeneração. Essa constatação pode ser feita quando há maior quantidade de
2157 indivíduos jovens na população e a diminuição progressiva do número de indivíduos mais
2158 maiores e mais velhos (Godínez-Álvarez *et al.*, 2003). Observamos que a proporção de
2159 indivíduos jovens variou bastante entre as populações de *P. neohorstii*.

2160 Estudos populacionais sugerem que cactos rupícolas podem apresentar menor
2161 quantidade de indivíduos jovens devido a limitações de microhabitats favoráveis para
2162 germinação e recrutamento (Martínez *et al.*, 2010). No entanto, a variação do número de
2163 indivíduos jovens entre as populações pode ocorrer devido ao impacto por animais, uma vez
2164 que as populações mais acessíveis apresentaram menor proporção de indivíduos jovens. Isso
2165 indica que a presença de animais domésticos pode influenciar na quantidade de indivíduos
2166 jovens, tanto devido aos impactos causados em indivíduos reprodutivos e consequente redução
2167 na produção de sementes ou no consumo direto indivíduos jovens inteiros que não deixaram
2168 vestígios nos afloramentos.

2169 A presença de indivíduos maiores é rara, e observamos poucos indivíduos grandes
2170 concentrados em populações maiores. A raridade dos indivíduos pode ocorrer devido a grande
2171 mortalidade causada pelo impacto de animais.

2172 A densidade populacional de *P. neohorstii* variou de 0,01 a 1,44 indivíduos/m² nos
2173 diferentes afloramentos de arenito. Estudos populacionais sugerem que a variação na densidade
2174 de indivíduos entre populações de cactos pode estar relacionada com os distúrbios nas áreas de
2175 ocorrência, onde populações mais densas ocorrem em áreas mais preservadas (Martorell &
2176 Peters, 2005; Martínez *et al.*, 2010). Nossos resultados foram consistentes com aqueles
2177 encontrados em estudos anteriores, uma vez que a população de *P. neohorstii* com maior
2178 densidade corresponde aquela com menor acesso de animais.

2179

2180 *Ações de conservação*

2181

2182 A área de ocorrência de *P. neohorstii* é uma área rural, onde a principal atividade
2183 desenvolvida é a pecuária de pequeno porte, principalmente ovelhas, cabras e gado. Uma ação
2184 imediata para a conservação de *P. neohorstii* seria o impedimento do acesso de animais aos
2185 afloramentos de arenito, através da construção de cercas nas áreas de ocorrência da espécie. A
2186 suspensão do uso dos afloramentos como área de pastagem não causaria danos econômicos,
2187 uma vez que poucas espécies com potencial alimentício ocorrem nessas áreas. Por outro lado,
2188 a restrição do acesso aos animais de criação nessas áreas evitaria o forte dano as populações de
2189 cactos rupícolas.

2190 Uma solução mais eficiente e a longo prazo seria a criação de uma unidade de
2191 conservação. A região das Guaritas é rica em formações geológicas raras e já foi reportado a
2192 importância de criação de um parque geológico na região (Paim *et al.*, 2010). Além disso, a
2193 diversidade biológica da região é única, como alta diversidade de abelhas especialistas
2194 (Schlindwein, 1998) e de cactáceas raras, sendo considerada uma das áreas prioritárias para a
2195 conservação de cactos no estado (Saraiva *et al.*, 2015). O sul do Brasil é uma das principais
2196 regiões do mundo onde há uma concentração de ocorrências de cactos ameaçados de extinção
2197 fora de áreas protegidas (Goettsch *et al.*, 2019) e *Parodia neohorstii* não se encontra inserida
2198 em nenhuma unidade de conservação (Carneiro *et al.*, 2016).

2199 Concluímos que medidas de contenção de animais domésticos nos locais de populações
2200 de *P. neohorstii* é primordial para garantir a manutenção da espécie.

2201 A diminuição progressiva dos indivíduos nas populações naturais pode contribuir para
2202 o declínio populacional. Essa espécie foi avaliada como criticamente ameaçada de extinção por
2203 Larocca e Machado (2013), principalmente devido a ocorrência restrita, indícios de declínio
2204 populacional e pouco conhecimento sobre as populações existentes. Este estudo pode contribuir
2205 para uma reavaliação do status de conservação de *P. neohorstii*, com dados mais robustos sobre
2206 a situação das populações remanescentes e a identificação das principais ameaças em habitat
2207 natural.

2208

2209 **REFERÊNCIAS**

2210

2211 **Anceschi G, Magli A.** 2018. A synopsis of the genus *Parodia* Spegazzini s.l. (Cactaceae).
 2212 *Bradleya* **1**: 70-161.

2213 **Anderson EF.** 2001. *The cactus family*. Timber Press (OR). 776 pp.

2214 **Bowers JE.** 1997. Demographic patterns of *Ferocactus cylindraceus* in relation to substrate
 2215 age and grazing history. *Plant Ecology* **133**: 37-48.

2216 **Carneiro AM, Farias-Singer R, Ramos RA, Nilson AD.** 2016. Cactos do Rio Grande do Sul.
 2217 Fundação Zoobotânica do Rio Grande do Sul, Porto Alegre. 224 pp.

2218 **Cerceau I, Siriani-Oliveira S, Dutra AL, Oliveira R, Schlindwein C.** 2019. The cost of
 2219 fidelity: foraging oligolectic bees gather huge amounts of pollen in a highly specialized
 2220 cactus–pollinator association. *Biological Journal of the Linnean Society* **128**: 30-43.

2221 **Dutra AL.** 2015. Acasalar demais é uma perda de tempo: custos da cópula para fêmeas de duas
 2222 espécies de abelhas solitárias. Dissertação (Mestrado em Ecologia de Biomas Tropicais) -
 2223 Instituto de Ciências Exatas e Biológicas, Universidade Federal de Ouro Preto, Ouro Preto.

2224 **Dutra AL, Schlindwein C, Oliveira R.** 2020. Females of a solitary bee reject males to collect
 2225 food for offspring. *Behavioral Ecology* **31**: 884–891

2226 **Esparza-Olguín L, Valverde T, Vilchis-Anaya E.** 2002. Demographic analysis of a rare
 2227 columnar cactus (*Neobuxbaumia macrocephala*) in the Tehuacan Valley, Mexico.
 2228 *Biological Conservation* **103**: 349-359.

2229 **Ferguson AW, Strauss RE, Strong AW, Birnbaum SJ, Poole JM, Janssen GK, Williamson**
 2230 **P S.** 2013. Assessing regional differences in predation of endangered species: Implications
 2231 from Texas populations of the endangered star cactus (*Astrophytum asterias*). *Journal of*
 2232 *Arid Environments* **97**: 143-149.

2233 **Godínez-Álvarez H, Valverde T, Ortega-Baes P.** 2003. Demographic trends in the Cactaceae.
 2234 *The Botanical Review* **69**: 173-201.

2235 **Goettsch B, Hilton-Taylor C, Cruz-Pinon G, Duffy JP, Frances A, Hernandez HM, Inger**
 2236 **R, Pollock C, Schipper J, Superina M, Taylor NP, Tognelli M, Abba AM, Arias S,**
 2237 **Arreola-Nava HJ, Baker MA, Barcenas RT, Barrios D, Braun P, Butterworth CA,**
 2238 **Burquez A, Caceres F, Chazaro-Basanez M, Corral-Diaz R, Del Valle Perea M, Demaio**
 2239 **PH, Duarte de Barros WA, Duran R, Yancas LF, Felger RS, Fitz-Maurice B, Fitz-**
 2240 **Maurice WA, Gann G, Gomez-Hinostrosa C, Gonzales-Torres LR, Patrick Griffith M,**
 2241 **Guerrero PC, Hammel B, Heil KD, Hernandez-Oria JG, Hoffmann M, Ishihara MI,**

- 2242 **Kiesling R, Larocca J, Leon-de la Luz JL, Loaiza SC, Lowry M, Machado MC, Majure**
 2243 **LC, Avalos JG, Martorell C, Maschinski J, Mendez E, Mittermeier RA, Nassar JM,**
 2244 **Negron-Ortiz V, Oakley LJ, Ortega-Baes P, Ferreira AB, Pinkava DJ, Porter JM,**
 2245 **Puente-Martinez R, Gamarra JR, Perez PS, Martinez ES, Smith M, Manuel**
 2246 **Sotomayor MDCJ, Stuart SN, Munoz JL, Terrazas T, Terry M, Trevisson M, Valverde**
 2247 **T, Van Devender TR, Veliz-Perez ME, Walter HE, Wyatt SA, Zappi D, Alejandro**
 2248 **Zavala-Hurtado J, Gaston KJ.** 2015. High proportion of cactus species threatened with
 2249 extinction. *Nature Plants* **1**: 15142.
- 2250 **Goettsch B, Duran AP, Gaston KJ.** 2019. Global gap analysis of cactus species and priority
 2251 sites for their conservation. *Conservation Biology* **33**: 369-376.
- 2252 **Gurvich DE, Pérez-Sánchez R, Bauk K, Jurado E, Ferrero MC, Funes G, Flores J.** 2017.
 2253 Combined effect of water potential and temperature on seed germination and seedling
 2254 development of cacti from a mesic Argentine ecosystem. *Flora* **227**: 18-24.
- 2255 **Hunt D, Taylor NP, Charles G.** 2006. *The New Cactus Lexicon*. 2 vols. DH Books, Milborne
 2256 Port. 900 pp.
- 2257 **Hunt D, Taylor NP, Charles G.** 2013. *The New Cactus Lexicon*, Illustrations. 2nd ed. DH
 2258 Books, Milborne Port. 527 pp.
- 2259 **INMET.** 2018. Banco de Dados Meteorológicos para Ensino e Pesquisa - BDMEP. Dados
 2260 históricos da estação meteorológica 83980 - Bagé - RS entre 1988 e 2018.
- 2261 **IUCN.** 2020. The IUCN Red List of Threatened Species. Version 2020-2.
 2262 <https://www.iucnredlist.org>
- 2263 **Jiménez-Sierra C, Mandujano MC, Eguiarte LE.** 2007. Are populations of the candy barrel
 2264 cactus (*Echinocactus platyacanthus*) in the desert of Tehuacán, Mexico at risk? Population
 2265 projection matrix and life table response analysis. *Biological Conservation* **135**: 278-292.
- 2266 **Larmuth J, Harvey HJ.** 1978. Aspects of the occurrence of desert plants. *Journal of Arid*
 2267 *Environments* **1**: 129-133.
- 2268 **Larocca J, Machado M.** 2013. *Parodia neohorstii*. *The IUCN Red List of Threatened Species*
 2269 *2013* (<http://dx.doi.org/10.2305/IUCN.UK.2013-1.RLTS.T152718A669833.en.>) Accessed
 2270 27/09/2018.
- 2271 **Machado M.** 2007. Fascinating *Frailea*, Part I: General impressions. *Cactus World* **25**: 5-11.
- 2272 **Machado MC, Zappi DC, Taylor NP, Borba EL.** 2005. Taxonomy and conservation of the
 2273 *Discocactus* Pfeiff. (Cactaceae) species occurring in the state of Bahia, Brazil. *Bradleya*,
 2274 **2005**: 41-56.

- 2275 **Maluf JR.** 2000. Nova classificação climática do Estado do Rio Grande do Sul. *Revista*
2276 *Brasileira de Agrometeorologia* **8**: 141-150.
- 2277 **Mandujano MC, Bravo Y, Verhulst J, Carrillo-Angeles I, Golubov J.** 2015. The population
2278 dynamics of an endemic collectible cactus. *Acta Oecologica* **63**: 1-7.
- 2279 **Martínez AF, Medina GIM, Golubov J, Montana C, Mandujano MC.** 2010. Demography
2280 of an endangered endemic rupicolous cactus. *Plant Ecology* **210**: 53-66.
- 2281 **Martínez-Ávalos JG, Golubov J, Mandujano MC, Jurado E.** 2007. Causes of individual
2282 mortality in the endangered star cactus *Astrophytum asterias* (Cactaceae): the effect of
2283 herbivores and disease in Mexican populations. *Journal of Arid Environments* **71**: 250-258.
- 2284 **Martínez-Ramos M, Arroyo-Cosultchi G, Mandujano MC, Golubov J.** 2016. Dinámica
2285 poblacional de *Mammillaria humboldtii* una cactácea endémica de Hidalgo, México.
2286 *Botanical Sciences* **94**: 199-208.
- 2287 **Martorell C, Peters EM.** 2005. The measurement of chronic disturbance and its effects on the
2288 threatened cactus *Mammillaria pectinifera*. *Biological conservation* **124**: 199-207.
- 2289 **McIntosh ME, Boyd AE, Arnold AE, Steidl RJ, McDade LA.** 2020. Growth and
2290 demography of a declining, endangered cactus in the Sonoran Desert. *Plant Species Biology*
2291 **35**: 6-15.
- 2292 **Paim PSG, Fallgatter C, Silveira ASD.** 2010. Guaritas do Camaquã, RS-Exuberante cenário
2293 com formações geológicas de grande interesse didático e turístico. pp. 1-14. In: **Winge M,**
2294 **Schobbenhaus C, Souza CRG, Fernandes ACS, Berbert-Born M, Sallun-Filho W,**
2295 **Queiroz ET.** *Sítios Geológicos e Paleontológicos do Brasil*. Brasília. 332 pp.
- 2296 **Peters EM, Martorell C, Ezcurra E.** 2008. Nurse rocks are more important than nurse plants
2297 in determining the distribution and establishment of globose cacti (*Mammillaria*) in the
2298 Tehuacán Valley, Mexico. *Journal of Arid Environments* **72**: 593-601.
- 2299 **Pianka ER.** 1978. *Evolutionary ecology*. Harper and Row, New York, USA. 486 pp.
- 2300 **Ribeiro-Silva S, Zappi DC, Taylor N, Machado M.** 2011. Plano de ação nacional para a
2301 conservação das cactáceas. Série Espécies Ameaçadas, 24.
- 2302 **Saraiva DD, Souza AF.** 2012. Effects of environmental factors and plantation forests on
2303 endangered cactus diversity and composition in subtropical South American grasslands.
2304 *Perspectives in Plant Ecology, Evolution and Systematics* **14**: 267-274.
- 2305 **Saraiva DD, de Sousa KDS, Overbeck GE.** 2015. Multiscale partitioning of cactus species
2306 diversity in the South Brazilian grasslands: Implications for conservation. *Journal for*
2307 *Nature Conservation* **24**:117-122.

- 2308 **Schlundwein C.** 1998. Frequent Oligolecty Characterizing a Diverse Bee–Plant Community in
2309 a Xerophytic Bushland of Subtropical Brazil. *Studies on Neotropical Fauna and*
2310 *Environment* **33**: 46-59.
- 2311 **Schlosser H.** 1979. Allgemeines über Uruguay und einige seiner Kakteen. *Kakteen und andere*
2312 *Sukkulenten* 213-217.
- 2313 **Valverde T, Quijas S, López-Villavicencio M, Castell, S.** 2004. Population dynamics of
2314 *Mammillaria magnimamma* Haworth (Cactaceae) in a lava-field in central Mexico. *Plant*
2315 *Ecology* **170**:167-184.
- 2316 **Warrick GD, Krausman PR.** 1989. Barrel cacti consumption by desert bighorn sheep. *The*
2317 *Southwestern Naturalist*, **34**: 483-486.
- 2318 **Zenteno Ruíz FS, López RP, Larrea Alcázar DM.** 2009. Patrones de distribución espacial de
2319 *Parodia maassii* (Heese) A. Berger (Cactaceae) en un semidesierto de los Andes
2320 subtropicales, la prepuna. *Ecología en Bolivia* **44**: 99-108.
- 2321

2322 **TABELAS**

2323

2324 **Tabela 1:** Características das populações de *Parodia neohorstii*. A área total do afloramento e a respectiva área de ocorrência dos indivíduos foram utilizadas
 2325 para calcular a porcentagem da área ocupada dentro do afloramento de arenito. Densidade de indivíduos foi calculada de acordo com a quantidade de indivíduos
 2326 vivos presentes na área de ocorrência dos cactos. Índice de regeneração representa a proporção de indivíduos jovens vivos, pertencentes a menor faixa de
 2327 tamanho (0 – 25mm) em relação a quantidade total de indivíduos vivos na população. A porcentagem de indivíduos impactados por animais foi calculado a
 2328 partir do número total de indivíduos amostrados, mortos e vivos. Acessibilidade a animais de criação: (Alta) Afloramento dentro de área de pastagem de bovinos
 2329 e caprinos; (Média) Afloramento cercado parcialmente sem acesso de bovinos, porém, com acesso de caprinos e outros animais; (Baixa) Afloramentos
 2330 inacessíveis a animais.

Populações	Número de indivíduos vivos	Número de indivíduos mortos	Área total do afloramento (m ²)	Área de ocorrência do cacto (m ²)	Área ocupada do afloramento (%)	Altitude (m)	Densidade (indivíduos vivos /m ²)	Índice de regeneração (%)	Indivíduos impactados por animais (%)	Acessibilidade a animais de criação
Ferradura	1056	82	54412	5222	9.59	241	0.20	24.9	26.1	Alta
Bom passo	747	20	12530	1958	15.62	210	0.38	33.6	8.2	Alta
Serro do Umbu	390	1	3351	270	8.05	210	1.44	82.0	0.8	Baixa
Pedra 2	336	42	54660	7641	13.97	213	0.04	36.9	14.8	Média
Ferradura Secreta	310	42	4073	916	22.48	235	0.33	9.6	30.4	Alta
Ferradura Lateral	208	34	774	184	23.77	231	1.13	20.6	19.4	Alta
Pedra 4	171	0	179867	12890	7.16	274	0.01	15.7	1.2	Média
Serro do Renato	117	23	96318	4298	4.46	300	0.02	47.0	22.9	Média
Passo ideal	93	5	1648	284	17.23	174	0.32	20.4	14.3	Alta

2331

2332 **Tabela 2:** Integridade de indivíduos de *Parodia neohorstii* em diferentes populações. Todos os
 2333 indivíduos da população foram contabilizados, vivos ou mortos. Os indivíduos foram classificados em
 2334 íntegros quando não havia nenhum tipo de dano físico. Indivíduos apresentaram dois tipos de danos
 2335 físicos causados por animais: (1) Solto do substrato com as raízes desprendidas, ou (2) Danificado
 2336 devido a mordidas ou pisoteio de animais, que causaram exposição do parênquima dos cactos.
 2337

Populações	Juvenil – N (%)			Reprodutivo – N (%)		
	Íntegro	Solto	Danificado	Íntegro	Solto	Danificado
Ferradura	251 (22.1)	25 (2.2)	14 (1.2)	590 (51.8)	131 (11.5)	127 (11.2)
Bom passo	263 (34.3)	16 (2.1)	4 (0.5)	441 (57.5)	27 (3.5)	16 (2.1)
Serro do Umbu	305 (78.0)	3 (0.8)	0 (0.0)	83 (21.2)	0 (0.0)	0 (0.0)
Pedra 2	145 (38.4)	5 (1.3)	14 (3.7)	177 (46.8)	12 (3.2)	25 (6.6)
Ferradura Secreta	29 (8.2)	1 (0.3)	2 (0.6)	216 (61.4)	64 (18.2)	40 (11.4)
Ferradura Lateral	38 (15.7)	3 (1.2)	1 (0.4)	157 (64.9)	11 (4.5)	32 (13.2)
Pedra 4	34 (19.9)	0 (0.0)	0 (0.0)	135 (78.9)	2 (1.2)	0 (0.0)
Serro do Renato	46 (32.9)	1 (0.7)	0 (0.0)	62 (44.3)	8 (5.7)	23 (16.4)
Passo ideal	21 (21.4)	4 (4.1)	0 (0.0)	63 (64.3)	8 (8.2)	2 (2.0)
Total	1132 (30.8)	58 (1.6)	35 (1.0)	1924 (52.3)	263 (7.2)	265 (7.2)

2338

2339

2340 **FIGURAS**

2341



2342

2343 **Figura 1:** População de *Parodia neohorstii*. Indivíduos ocorrerem afloramentos de arenito e
2344 conglomerados.

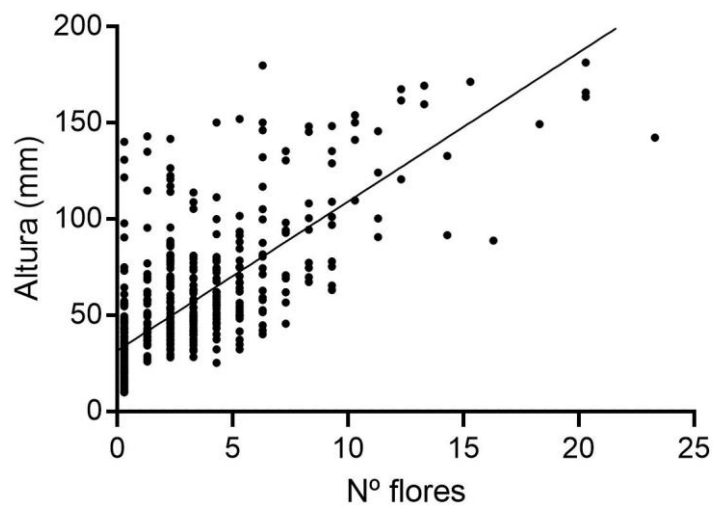
2345



2346

2347 **Figura 2:** Instrumento de medida rápida de altura de cactos. Cada cor no bastão representa uma faixa
2348 de altura: (1) Rosa: 0-25 mm; (2) Verde bandeira: 25-50 mm; (3) Verde fluorescente: 50-75 mm; (4)
2349 Vermelho: 75-100 mm; (5) Azul: 100-125 mm; (6) Amarelo: 125-150mm; (7) Preto: 150-175 mm; (8)
2350 Laranja: 175-200 mm. A figura mostra um indivíduo com altura de 75-100 (faixa vermelha).

2351



2352

2353 **Figura 3:** Correlação entre a altura dos cactos e a quantidade de flores produzidas durante a estação
2354 reprodutiva (N = 661).

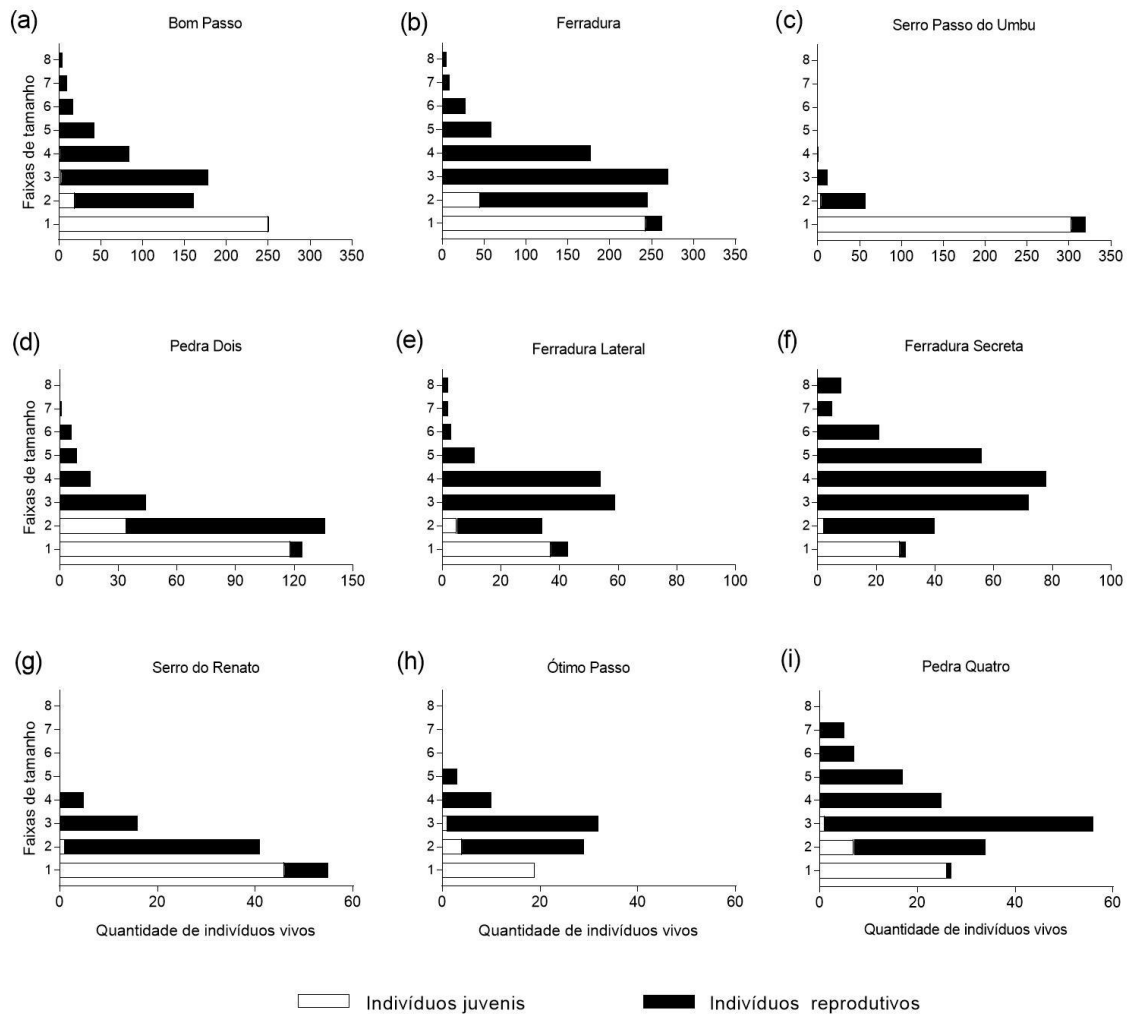
2355



2356

2357 **Figura 4:** Indivíduos de cabras na área de ocorrência de *Parodia neohorstii*. As cabras acessam o
2358 afloramento pelas partes íngremes, onde não existem cercas.

2359



2360

2361 **Figura 5:** Levantamento populacional de indivíduos vivos de *Parodia neohorstii* em nove localidades.

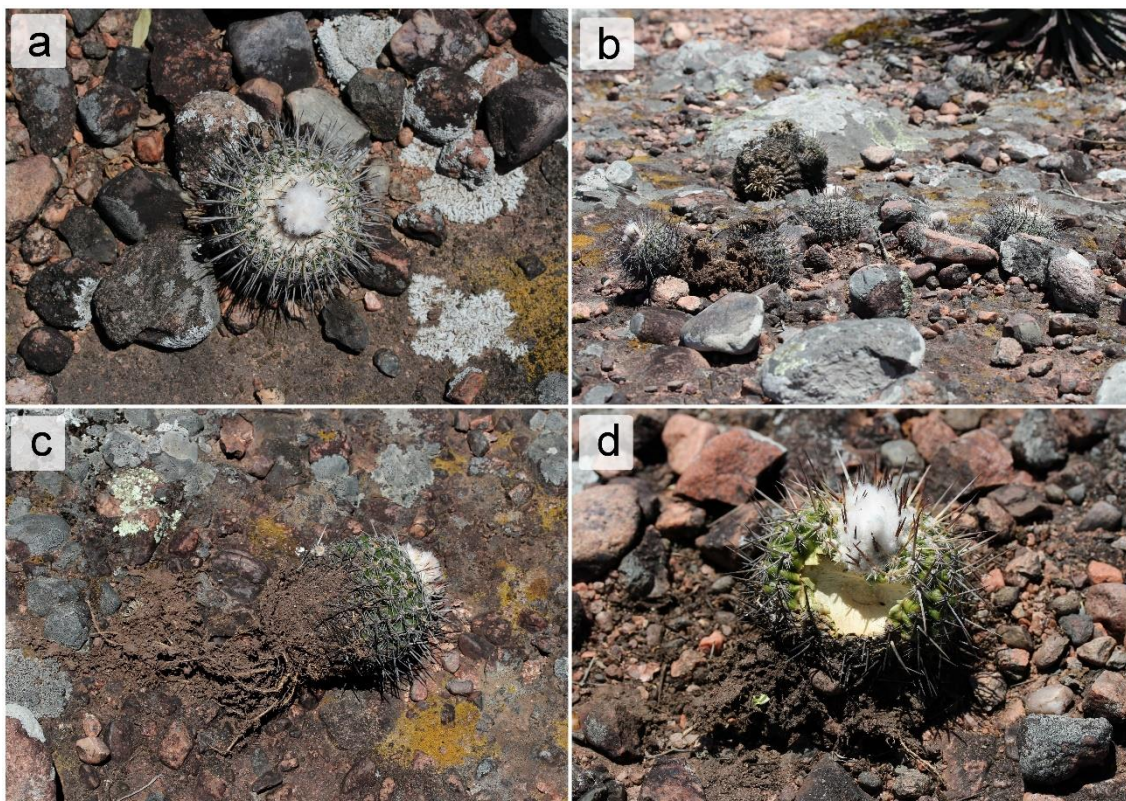
2362 Os indivíduos foram classificados de acordo com faixas de tamanho: (1) 0-25 mm; (2) 25-50 mm; (3)

2363 50-75 mm; (4) 75-100 mm; (5) 100-125 mm; (6) 125-150mm; (7) 150-175 mm; (8) 175-200 mm. Barras

2364 brancas apresentam indivíduos juvenis, que ainda não produziram flores e frutos. Barras pretas

2365 apresentam indivíduos reprodutivos.

2366



2367

2368

2369

2370

2371

2372

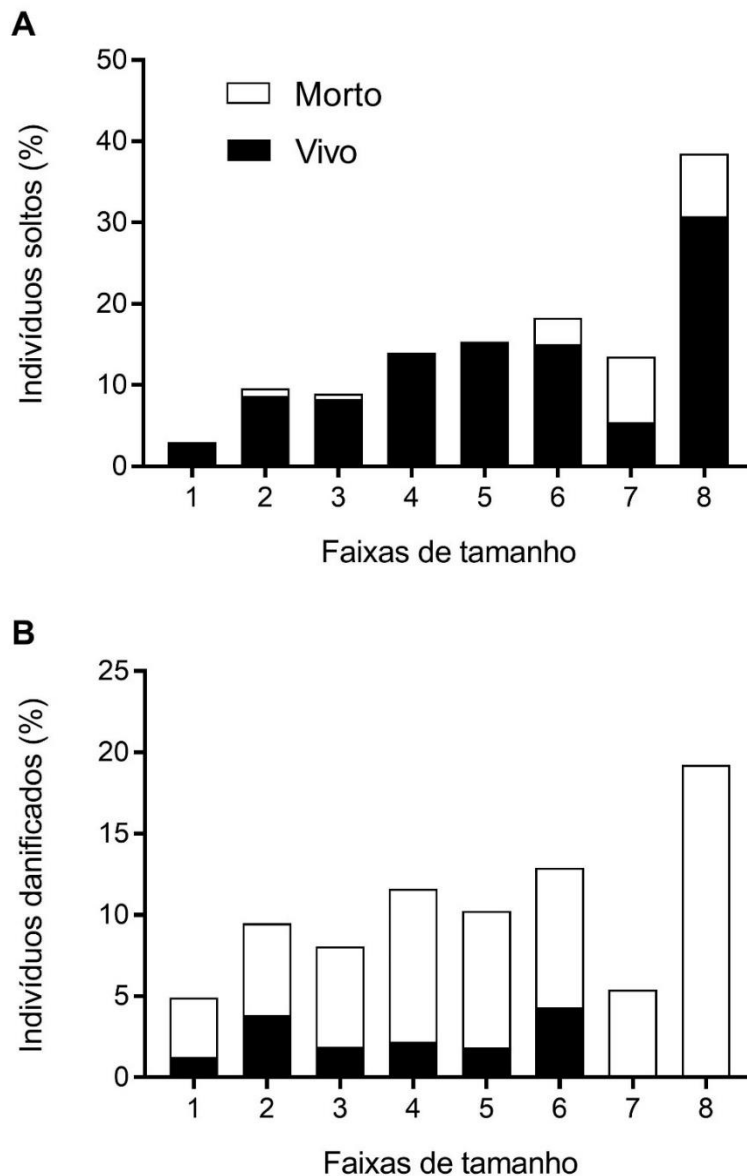
Figura 6: Indivíduos de *Parodia neohorstii*. (a) Indivíduo íntegro, com raízes fixadas entre frestas e conglomerados. (b) Grupo de indivíduos de *P. neohorstii*, indivíduos íntegros ao lado de indivíduos removidos do substrato. (c) indivíduos removidos do substrato com as raízes totalmente desprendidas. (d): Indivíduo danificado por mordida, com o parênquima exposto.



2373

2374 **Figura 7:** Filhote de cabra se alimentando de um indivíduo de *Parodia neohorstii*.

2375



2376

2377 **Figura 8:** Percentagem de indivíduos mortos e vivos de acordo com o tipo de impacto e faixa de
 2378 tamanho em todas as populações. Faixas de tamanho correspondem a altura dos cactos: (1) 0-25 mm;
 2379 (2) 25-50 mm; (3) 50-75 mm; (4) 75-100 mm; (5) 100-125 mm; (6) 125-150mm; (7) 150-175 mm; (8)
 2380 175-200 mm. (A) Percentagem de indivíduos removidos do substrato após a passagem de animais em
 2381 cada faixa de tamanho, classificados como vivos ou mortos. (B) Percentagem de indivíduos danificados
 2382 por mordidas ou pisoteio em cada faixa de tamanho, classificados como vivos ou mortos.

2383

CAPÍTULO 4

2384

2385

2386

2387

2388

2389

2390

2391

REPRODUCTION OF *FRAILEA GRACILLIMA* (CACTACEAE) IN NATURAL

2392

HABITAT: POLLINATION BY SOLITARY BEES AND MECHANISM OF

2393

DELAYED SELF-POLLINATION

2394

2395 **REPRODUCTION OF *FRAILEA GRACILLIMA* (CACTACEAE) IN NATURAL**
2396 **HABITAT: POLLINATION BY SOLITARY BEES AND MECHANISM OF**
2397 **DELAYED SELF-POLLINATION**

2398

2399 **ABSTRACT**

2400

2401 The genus *Frailea* includes small cacti that predominantly cleistogamous. However, the
2402 knowledge is based on cultivated specimens and information from individuals in the natural
2403 habitat is scarce. During three seasons, we studied pollination, characteristics of flowering and
2404 breeding system of *Frailea gracillima*, a species that occurs on sandstone outcrops in the
2405 Brazilian Pampa. No cleistogamous flowers were recorded in the field. The pollen-only flowers
2406 were visited by females of several pollen collecting solitary bee species and male bees seeking
2407 for mates. Bees deposited huge amounts of conspecific pollen on the stigma, sufficient to
2408 fertilize all ovules, but also pollen from other different plant families. Flower opening was
2409 highly synchronized in the population but flowering days were interrupted by non-blooming
2410 periods and occurred in only 11% of the days in the flowering season. Lifespan of flowers
2411 visited by bees was only ~2:30h. The flowers showed temporal herkogamy, where stamens and
2412 stigma are spatially separated towards the end of anthesis. The long stigma lobes slowly move
2413 downward during anthesis and contact anthers when petals fold thus promoting delayed
2414 autonomous self-pollination. The quantity of pollen grains deposited on stigmas autonomously,
2415 however, varied between flowers, and fruit set via autonomous pollination was 56%. Flowering
2416 characteristics in *F. gracillima* strongly favor cross-pollination but, in the absence of solitary
2417 bee pollinators, set seed through delayed self-pollination.

2418 **Key words:** *Frailea gracillima*, solitary bees, delayed self-pollination, Cactaceae, floral
2419 movements, Brazil, Pampa cacti.

2420

2421

2422 **INTRODUCTION**

2423

2424 Among plant reproductive strategies, most species invest in floral traits to attract
 2425 pollinators and in mechanisms that favor cross-pollination (Sprengel, 1793; Darwin, 1862;
 2426 Faegri & van der Pijl, 1979). Autonomous self-pollination, however, can guarantee sexual
 2427 reproduction in low pollinator environments (Lloyd & Schoen, 1992; Sun *et al.*, 2005). Some
 2428 species reproduce through cleistogamy and promote self-pollination in closed flower buds,
 2429 producing fertile seeds in the absence of pollen vectors (Uphof, 1938; Lord, 1981; Schoen &
 2430 Lloyd, 1984; Richards, 1997). Cleistogamy occurs in at least 56 families in different habitats
 2431 of the world (Lord, 1981), but is common at higher altitudes, regions with extreme weather
 2432 conditions and poor pollination environments (Uphof, 1938; Faegri & van der Pijl, 1979).
 2433 Exclusively cleistogamous species are rare (Lord, 1981) and often the same individual can
 2434 produce cleistogamous and chasmogamous flowers (open flowers) according to environmental
 2435 conditions such as temperature, humidity, photoperiod and soil quality (Uphof, 1938; Brown,
 2436 1952; Faegri & van der Pijl, 1979; Langer & Wilson, 1965; Schoen & Lloyd, 1984; Richards,
 2437 1997).

2438 While floral traits in Cactaceae in general are clearly related to specific pollinator groups
 2439 such as bats, bees, hawkmoths and hummingbirds (Porsch, 1938; Marshall, 1941; Grant &
 2440 Grant, 1979; Rowley, 1980; Schlindwein & Wittmann, 1995; Anderson, 2001; Cerceau *et al.*,
 2441 2019) and xenogamy seems to be common (Gibson & Nobel, 1986; Mandujano, 2010), a few
 2442 taxa are cleistogamous. This is reported for some species of *Rhipsalis* Gaertn. (Barthlott &
 2443 Hunt, 1993), *Melocactus* Link & Otto (Taylor, 1991) and *Frailea* Britton & Rose (Putnam,
 2444 1968). Among cacti, *Frailea* is considered the genus with most cleistogamous species (Rowley,
 2445 1980). Nevertheless, both cleistogamous and chasmogamous flower production were
 2446 mentioned for the genus (Britton & Rose, 1922; Köhler, 1968; Anderson, 2001; Metzinger &
 2447 Kiesling, 2006). However, this information is based on cacti collections found outside the
 2448 natural occurrence of *Frailea* species, often where plants were grown in greenhouses. Though,
 2449 in a bee-plant community survey, flowers of *Frailea phaeodisca* (Speg.) Backeb. & F.M.Knuth
 2450 and *Frailea pygmaea* (Speg.) Britton & Rose were cited to receive visits of bees (Schlindwein,
 2451 1998). Up to the present moment, there was no

2452 no need for unfortunately, no detailed information on pollination and reproductive biology of
 2453 species of *Frailea* in natural habitats.

2454 *Frailea's* cleistogamy seems to be strongly related to abiotic conditions. In cultivation,
 2455 where climatic conditions are very different from those observed in natural habitat, cleistogamy
 2456 seems to be predominant (Köhler, 1968; Metzging & Kiesling, 2006).

2457 We studied *Frailea gracillima* (Lem.) Britton & Rose a species with a tiny solitary body
 2458 that occurs in the south of Rio Grande do Sul, in sandstone rock outcrops. In the literature, the
 2459 species is reported as reproducing via cleistogamy (Putnam, 1968) and chasmogamy (Britton
 2460 & Rose, 1922).

2461 In order to elucidate the breeding system and pollination of *Frailea gracillima* we
 2462 asked: 1) What are the characteristics of flowering and anthesis of this species in natural
 2463 habitat? 2) What is its breeding system? 3) Who are their floral visitors and effective
 2464 pollinators? 4) Is there pollen deposition on the stigmas promoted by bees and autonomously?
 2465 To answer these questions, we studied a natural population of *F. gracillima* during three
 2466 flowering seasons in the field, described the flower opening schedule and rhythm of flowering
 2467 in three seasons, determined the breeding system, as well as the spectrum of floral visitors and
 2468 quantified pollen grains on the stigma.

2469

2470 MATERIAL AND METHODS

2471

2472 *Study area and species*

2473

2474 The study was conducted in the Serra do Sudeste, in the municipalities Santana da Boa
 2475 Vista and Caçapava do Sul (30°53'16.1"S 53°24'04.1"W), Rio Grande do Sul, Brazil. The
 2476 region is part of the Pampa grasslands, and the study areas are characterized by rock outcrops
 2477 and shallow stony and sandy soils in the region of the *Guaritas*, which are steep sandstone and
 2478 conglomerate formations (Carneiro *et al.*, 2016). The local vegetation is dominated by
 2479 herbaceous and shrubby species, with some small trees (Schlindwein, 1998). The region is a
 2480 diversity centre for cactus species, many of them are threatened (Schlindwein, 1995; Carneiro
 2481 *et al.*, 2016). The average annual precipitation in the region is 1509 mm and the average annual
 2482 temperature is 18.7 ° C (INMET, 2018). In the flowering season of most cacti species, (October
 2483 to December), the maximum monthly average temperature is 27.1 ° C (INMET, 2018).

2484 With eleven among the twelve recognized species of *Frailea*, Rio Grande do Sul is the
 2485 center of diversity of this genus (Ribeiro-Silva *et al.*, 2011; Metzging 2015a, 2015b). *Frailea*
 2486 *gracillima* is a small cactus species, restricted to the Pampa, with globose to cylindrical body

2487 up to 10 cm height and 2.5 cm diameter. Individuals occur solitary, scattered or in small groups
2488 (Britton & Rose, 1922; Carneiro *et al.*, 2016) on rock soils, especially in rock cracks on the
2489 sandstone outcrops (Figure 1).

2490

2491 *Anthesis and flower longevity*

2492

2493 In the flowering season 2016, 2017 and 2018 of *F. gracilima*, we daily recorded the
2494 precipitation with a pluviometer to check whether flowering is related to the rainfall regime.
2495 We marked flower buds in pre-anthesis with color codes and daily checked whether flowers
2496 opened and noted the day of flowering (N = 131). We recorded the opening and closing times
2497 of the flowers.

2498 We determined and compared the floral longevity of: 1) flowers visited by bees (N =
2499 25); 2) bagged non-visited flowers (N = 7); 3) hand cross-pollinated flowers (N = 6); 4) hand
2500 self-pollinated flowers (N = 6). Hand cross-pollinated and hand self-pollinated flowers were
2501 bagged in the bud stage, pollinated, and then bagged again.

2502

2503 *Breeding system*

2504

2505 To determine the breeding system of *F. gracillima* we compared the amount of fruits
2506 and seeds produced in different pollination treatments: 1) natural pollination (N = 11); 2)
2507 autonomous self-pollination (N = 9); 3) hand self-pollination (N = 6); 4) hand cross-pollination
2508 (N = 2).

2509 To determine the number of ovules per flower, we fixed 17 flowers in 70% alcohol and
2510 counted the number of ovules under a stereomicroscope (Leica, WILD – M3Z).

2511

2512 *Movement of stamens and stigmas*

2513

2514 To check whether there is spatial separation of anthers and stigmas, and their movement,
2515 we followed 10 flowers throughout the anthesis and recorded: 1) position of the stigmatic lobes
2516 and stamens along the anthesis; 2) contact between stigmatic lobes and anthers along the
2517 anthesis.

2518 We checked if stamens moved in response to touches. We also recorded the number of
2519 stigmatic lobes per flower.

2520

2521 *Floral visitors*

2522

2523 The frequency of flower visitors was determined throughout anthesis of 15 flowers, in
2524 30 minutes-intervals, totaling 30 hours of observation. We record the following information
2525 during the flower visits: 1) bee species; 2) sex of the bee; 3) stigma contact. We also quantified
2526 where the bees alighted on the flowers and when they touched the stigma (beginning of visit,
2527 during, or end of the floral visit) and where they collect pollen (from anthers or adhering to
2528 petals). To analyze the behavior of the floral visitors in detail, we made video recordings (200
2529 minutes, Canon EOS T5i).

2530

2531 *Pollen deposition on the stigma*

2532

2533 In flowers accessible to flower visitors (N = 10) and in bagged flowers (N = 3) we
2534 removed the stigmas at the end of anthesis to count and identify the adhering pollen grains. The
2535 stigmas were transferred to microscope slides containing fuchsine stained glycerine gelatin, the
2536 slides were heated to melt the gelatin, and the preparations were covered with a coverslip and
2537 sealed with paraffin wax. Under a microscope we counted and identified all the pollen grains
2538 that adhered to the stigmas.

2539

2540 *Statistical analyses*

2541

2542 To check if there was a difference in the number of bee visits received by the flowers
2543 throughout the anthesis, we used the non-parametric Kruskal-Wallis one way analysis of
2544 variance on ranks. Statistical tests were performed with SigmaStat 3.5 software for Windows.

2545

2546 **RESULTS**

2547

2548 *Anthesis and floral longevity*

2549

2550 *Frailea gracillima* bloomed between October and November. In 108 days of
2551 observation during three years (30 consecutive days in 2016, 44 in 2017 and 34 in 2018), we

2552 recorded only 12 days of flowering (3 days of flowering in 2016, 4 in 2017 and 5 in 2018). We
 2553 did not find any open flowers in the field in 89% of the study period.

2554 All previously marked flower buds opened their flowers (N = 131). Most individuals
 2555 produced one flower per day (95.2%, N = 125 individuals), 6 individuals produced 2 flowers
 2556 simultaneously (4.8%, N = 125) and only one individual produced 4 flowers. Flowering
 2557 occurred only on sunny days. Individuals bloomed 3.0 ± 2.1 days after rain (N = 12 flowering
 2558 days) (Figure 2).

2559 Individuals of the population opened flowers synchronously. Anthesis in all marked
 2560 flowers started $12:24h \pm 10$ min and the flowers remained open for 150 ± 13 min (N = 26). At
 2561 the beginning of the anthesis the anthers were already dehisced and all pollen grains available
 2562 to flower visitors. Flowers did not produced nectar.

2563 Flowers visited by bees did not open on the next day (N = 25), but bagged flowers that
 2564 received different pollination treatments without access to pollinator opened for up to 2 days.
 2565 All hand self-pollinated flowers for 2 days (N = 6), 83% of hand-cross pollinated flowers
 2566 opened for 2 days (N = 6), and 85% of permanently bagged flowers opened for 2 days (N = 7).
 2567 We did not observe cleistogamous flowers during the study period.

2568

2569 *Breeding system*

2570

2571 Flowers of *F. gracillima* produced on average of 291 ± 73 ovules (N = 17). The
 2572 individuals of the studied population were self-compatible and all hand-pollinated (self and
 2573 cross) and open-pollinated flowers set 100% of fruits. Fruit set in unmanipulated bagged
 2574 flowers was 56%, with only 5 of the 9 flowers setting fruit, with 128 ± 70 seeds. Flowers
 2575 accessible to bees produced fruits with 301 ± 74 seeds (N = 11), hand-cross pollinated flowers
 2576 produced 144 ± 12 seeds (N = 2) and hand-self-pollinated flowers produced 118 ± 66 seeds (N
 2577 = 6).

2578

2579 *Movement of stamens and stigma*

2580

2581 *Frailea gracillima* produce hermaphroditic flowers with spatial separation of
 2582 reproductive organs at the beginning of anthesis. Experimental mechanical stimulation of the
 2583 stamens did not cause movement toward the style and revealed that they were not thigmonastic.

2584 In newly opened flowers, the stamens are arranged in two positions: The outer stamens
2585 are positioned near to the petals and the inner stamens are close to the style (Figure 3A, Figure
2586 4). Due to this arrangement, an empty space between both groups of stamens is formed along
2587 the floral tube. The stamens have different lengths, because of this, they form dense layers of
2588 anthers arranged around the style and in contact with the petals. During anthesis, from
2589 approximately 10 minutes after flower opening, the outer stamens slowly moved towards the
2590 center and the inner stamens also moved outwards (Figure 4). After around 60-80 min stamens
2591 were spread across the floral tube, partially detaching from contact with the petals, and filled
2592 the empty space in the floral tube between inner and outer stamens.

2593 The stigmas had on average 10 ± 1 lobes ($N = 35$), and were positioned vertically in
2594 relation to the style in the flower bud stage. From the first minutes after flower opening, the
2595 stigma lobes slowly moved downwards from its upward towards the stamens. The extremity of
2596 stigmatic lobes curls and increases the proximity to the anthers (Figure 3B). During flower
2597 closure, the stigma lobes can be pushed downwards by the petals and get into contact with
2598 anthers (Figure 5A). Eventually, in some flowers there is contact of the extremity of stigmatic
2599 lobes with pollen grains (Figure 5B). However, not all flowers set fruits autonomously,
2600 indicating that pollen deposition by this mechanism does not occur in all flowers.

2601

2602 *Floral visitors*

2603

2604 We collected 495 floral visitors, belonging 13 bee species of four families visiting
2605 flowers of *F. gracillima* (Table 1). The main floral visitor was *Anthrenoides micans* Urban,
2606 1995, which corresponded to 70.7% of the visits observed (Figure 6A), followed by *Dialictus*
2607 *sp.* with 22.6% of the visits (Figure 6B). *Arhysosage cactorum* Moure, 1999 and *Chalepogenus*
2608 *goeldianus* Friese, 1899 had low visitation frequencies (3.3 and 2.2% respectively). Other nine
2609 bee species were sporadic floral visitors.

2610 For three bee species *Anthrenoides micans*, *Arhysosage cactorum* and *Callonychium*
2611 *sp.*, we recorded both male and female individuals, although females were always more
2612 frequent.

2613 *Anthrenoides micans* showed reproductive behavior attached to cactus flowers. Males
2614 patrolled flowers in search of females and every minute they inspected a single flower five
2615 times ($N = 341$ inspections in 61 minutes). During the study period, 15 copulas were observed,
2616 which represented 3.0% of the visits received by *F. gracillima*. In this situation, couples touched

2617 the stigma in 100% of the visits. We observed only one copula of *Arhysosage cactorum* and
 2618 none of *Callonychium* sp.

2619 Female bees visited flowers throughout anthesis and there was no difference in the number of
 2620 visits over the time ($P = 0.690$, $H = 3.064$, $N = 141$) (Figure 7). Female bees of *Anthrenoides*
 2621 *micans* mainly used the stigma as landing platform (87% of visits, $N = 30$), while females of
 2622 *Dialictus* sp. alighted mainly on the petals (87% of visits, $N = 15$).

2623 Bees were positioned in two ways to collect pollen: directly on the petals or inside the
 2624 floral tube, positioned upside down between the inner and outer stamen bundle. Females of *A.*
 2625 *micans* landed upside down on the floral tube, perching on the style and collected pollen
 2626 preferentially from stamens of the inner bundle, where anthers contact the style (76.6%, $N =$
 2627 30, Figure 8). Due to the position of the bee inside the flower, there was pollen deposition on
 2628 the body of the bee in 86.6% of the visits. *Dialictus* sp. landed on the petals to collect pollen
 2629 directly from the outer bundle of stamens or pollen grains adhering to the petals (80.0%, $N =$
 2630 15). Pollen deposition on the body of *Dialictus* sp. occurred in 20% of visits. When leaving the
 2631 flower, *A. micans* mainly used the petals (66.6%, $N = 30$) while *Dialictus* sp. climbed over the
 2632 style and left from the stigma (53.3%, $N = 15$).

2633 In general, bees have often touched flower stigmas. Sporadic visitors such as
 2634 *Arhysosage cactorum*, *Chalepogenus goeldianus* and *Cactocolletes rugatus* (Urban, 1995)
 2635 Engel touched the stigma in all visits. Females of *Anthrenoides micans* females touched the
 2636 stigma in 95.8% of the visits ($N = 48$), *Dialictus* sp. touched only in 21.3% of visits ($N = 75$).

2637

2638 *Pollen deposition on the stigma*

2639

2640 In flowers visited by bees, the average number of conspecific pollen grains deposited in
 2641 stigma surface was 1847 ± 750 ($N = 10$), being six times higher than the number of ovules.
 2642 Heterospecific pollen varied from 1 - 17% of the total pollen load, mainly from Asteraceae,
 2643 Malvaceae, Poaceae and Euphorbiaceae (*Croton* sp.), Cactaceae (*Opuntia* sp.).

2644 In bagged flowers, autonomous pollen deposition on stigma was on average 2318 ± 1740 pollen
 2645 grains ($N = 3$).

2646

2647 **DISCUSSION**

2648

2649 In the natural habitat, *Frailea gracillima* exclusively produces chasmogamous flowers
 2650 and floral traits strongly favor cross-pollination. Flowering is concentrated in just a few days
 2651 during the reproductive season with periods of flowering breaks. The ephemeral and highly
 2652 synchronized flowering among the individuals of the population attracts pollinators that deposit
 2653 a huge amount of pollen grains on the stigma, leading to high seed set. The main pollinators
 2654 are the same solitary bee species that visits sympatric globose cacti of the genera *Parodia* and
 2655 *Gymnocalycium* (Schlindwein & Wittmann, 1995; Cerceau *et al.*, 2019). However, when
 2656 pollinators are absent or scarce, the chasmogamous flowers reproduces through delayed
 2657 autonomous self-pollination due movements of both stigma lobes and stamens during anthesis.

2658

2659 *Flowering pattern and floral traits of Frailea gracillima favor cross pollination*

2660

2661 Different to literature (Rowley, 1980; Metzging & Kiesling, 2006), our results reveal that
 2662 flowering pattern and floral traits of *F. gracillima* strongly favour cross pollination, which
 2663 might be also true for other species of the genus.

2664 Most individuals open only one flower per day, which can drastically reduce
 2665 geitonogamy and favours cross-pollination (de Jong *et al.*, 1993). Consequently, almost all floral
 2666 visits of the foraging trips of the bees lead to cross-pollen deposition on the stigma.

2667 The entire population of *F. gracillima* blossomed in the same few days during reproductive
 2668 season and the start of the anthesis was synchronized, therefore huge quantities of flowers were
 2669 available for a short period of time in rock outcrops. Flowering synchronization within
 2670 population may increase pollinator attractiveness, favour cross-pollination (Opler *et al.*, 1976;
 2671 Augspurger, 1981) and it is an important trait in melittophilous cacti, because bees can
 2672 concentrate visits at the beginning of anthesis (Cerceau *et al.*, 2019). Anthesis synchronization
 2673 was previously recorded in *Frailea* species both in natural habitat (Machado, 2007) and in
 2674 cultivation (Metzging & Kiesling, 2006). Both fresh pollen and receptive stigmas are exposed to
 2675 pollinators in short-time, optimizing reproduction by diminishing pollen loss in the population.
 2676 Anthesis synchronization may be related to abiotic factors, such as rainfall, as has been reported
 2677 in other species of cacti (Eggli & Giorgetta, 2017; Cerceau *et al.*, 2019). However, precipitation
 2678 does not seem to be strongly related to the flowering of *F. gracillima*, since the gap between
 2679 rainy days and open-flower days was very variable.

2680 *Frailea gracillima* share floral traits with other sympatric cacti species that receive visits
 2681 of the same specialized effective bee pollinators, such as flower colour, shape, apical position

2682 of flowers in cactus body and erect position of flowers near the soil surface. Sympatric *Parodia*
 2683 and *Gymnocalycium* also synchronized flower opening and closure within the population at a
 2684 similar time schedule to *F. gracilima*, between 11:00 and 17:30 (Schlindwein & Wittmann,
 2685 1995; Cerceau *et al.*, 2019), however, surprisingly there were no flowering days overlap in rock
 2686 outcrops (our unpubl. data).

2687 Another characteristic that favours cross-pollination in *F. gracillima* is hercogamy,
 2688 which prevent contact between stigma and anthers (Faegri & van der Pijl, 1979; Webb & Lloyd,
 2689 1986).

2690 The prominent stigma composed by numerous and long lobes, positioned at flower
 2691 entrance diminish the free space to flower visitors reach pollen resources below, which favour
 2692 contact between bees and stigma. The stigma of *F. gracillima* is bigger than that of other
 2693 sympatric melittophilous cacti, such as *Parodia* and *Gymnocalycium*, which have a tiny central
 2694 almost circular stigmatic surface. However, specialist bees use the stigmatic surface of these
 2695 cacti species as a landing platform (Schlindwein & Wittmann, 1995; Cerceau *et al.*, 2019),
 2696 similar to *F. gracilima*.

2697 *Frailea* species in Rio Grande do Sul have high habitat specificity and occur in rock
 2698 outcrops of different lithologies (Pontes *et al.*, 2018). Delimitation of populations on rock
 2699 outcrops can favors cross-pollen flow among individuals, since the outcrops are isolated in the
 2700 landscape of the shrubby fields.

2701

2702 *Solitary bee pollinators*

2703

2704 The floral visitors that contributed to the pollination of *F. gracillima* were solitary bees
 2705 that touched the stigma during flower visits. The position of the two bundles of stamen within
 2706 the flower favored pollen deposition on the body of bees, which is transferred to conspecific
 2707 flowers on the next visits, favoring cross-pollination. Several cacti species have floral traits that
 2708 promote pollen deposition to insect body, such stamen movement (Lenzi & Orth, 2011; Cota-
 2709 Sánchez *et al.*, 2013; Schlindwein & Wittmann, 1997). Thigmonastic stamen movements were
 2710 not observed in *Frailea*, but it is frequent trait in *Parodia* and *Opuntia* flowers (Schlindwein &
 2711 Wittmann 1995, 1997; Cerceau *et al.*, 2019).

2712 The main floral visitor of *F. gracillima* is *A. micans*, registered as a frequent floral visitor
 2713 of sympatric globose yellow-flowered cacti (Schlindwein & Wittmann, 1995; Schlindwein,
 2714 1998; Cerceau *et al.*, 2019). *Anthrenoides micans* exhibits reproductive behavior attached to

2715 cactus flowers, which considerably increases its presence inside the flowers. Males intensely
 2716 patrol flowers in the search of pollen-collecting females and also copulate inside them (Dutra
 2717 *et al.* 2020). We observed that copulating bees in flowers of *F. gracillima* frequently touch the
 2718 stigmas, different to copulas in *Parodia* flowers that have smaller stigma (Schlindwein &
 2719 Wittmann, 1995; Cerceau *et al.*, 2019). The reproductive behavior of specialized bees attached
 2720 to host plant greatly increases flower visits and constancy, as well as effectiveness of pollination
 2721 (Schlindwein & Wittmann, 1995; Oliveira & Schlindwein, 2010; Oliveira *et al.*, 2012, 2013).

2722 Besides *A. micans*, the oligolectic bee species *A. cactorum* and *C. rugatus* also collected
 2723 pollen in flowers of *F. gracillima*, however with a low visitation frequency. During the short
 2724 floral lifespan of *F. gracillima*, these bees were observed flying among conspecific flowers,
 2725 which provided a stigma pollen load with huge amounts of conspecific grains.

2726 The tiny *Dialictus* bees rarely touched the stigma and little contributed to pollination similar as
 2727 observed for other cacti species (Schlindwein & Wittmann, 1995, 1997; Cerceau *et al.*, 2019).

2728

2729 *Temporal herkogamy and delayed autonomous self-pollination*

2730

2731 The flowers of *F. gracillima* show herkogamy, which corresponds to spatial separation
 2732 of reproductive organs that favors cross-pollination and reduces interference between the male
 2733 and female function (Faegri & van der Pijl, 1979; Webb & Lloyd, 1986).

2734 Hercogamy in the flowers of *F. gracillima* occurs only at the beginning of anthesis. The
 2735 spatial separation of stigmas and anthers slowly diminishes during anthesis with the stigmas
 2736 moving downwards, therefore contacting the pollen available in the anthers. The temporal
 2737 herkogamy prevents autonomous self-pollination during the first hours of anthesis. The curving
 2738 down of stigma lobes is well known in representatives of Campanulaceae and Asteraceae that
 2739 have secondary pollen presentation, where stigmas can touch pollen grains that eventually still
 2740 adhere to the style at the end of anthesis (Faegri & van der Pijl, 1979; Shetler, 1979). In
 2741 Malvaceae also occurs curving down of un-pollinated stigma lobes during anthesis and
 2742 consequently promote delayed autonomous self-pollination (Buttrose *et al.*, 1977; Klips &
 2743 Snow, 1997; Ruan *et al.*, 2005; Ruan & Mopper, 2008).

2744 In closing flowers of *F. gracillima*, where cross-pollination is no longer possible,
 2745 autonomous pollen deposition result in fruit set. Delayed self-pollination occurs when floral
 2746 movements at the end of anthesis promote contact between anthers and stigma (Lloyd &
 2747 Schoen, 1992). This mechanism ensures reproduction when flowers were not pollinated (Faegri

2748 & van der Pijl, 1979) and can be considered a mechanism of reproductive assurance in *F.*
2749 *gracillima* flowers.

2750 Curving down of stigma lobes seems to be frequent among species of *Frailea* and often
2751 appears in photographic records (Metzing, 2015a, 2015b; Carneiro *et al.*, 2016). It would be
2752 interesting to confirm delayed self-pollination after floral movements also in other species of
2753 *Frailea* to elucidate whether this reproductive strategy is a common trait of the genus.

2754

2755 *Cleistogamy in Frailea*

2756

2757 Our study revealed that the individuals of *F. gracillima* do not exhibit cleistogamous
2758 flowers in the natural habitat. This contradicts information common in literature that most
2759 *Frailea* species reproduce via cleistogamy and the opening of chasmogamous flowers is rare
2760 (Britton & Rose, 1922; Rowley, 1980; Metzing & Kiesling, 2006)

2761 In several species that produce both cleistogamous and chasmogamous flowers,
2762 cleistogamy is mainly related to abiotic conditions, such as photoperiod, temperature, soil
2763 moisture, shade and nutritional quality (Uphof, 1938; Brown, 1952; Langer & Wilson, 1964;
2764 Faegri & van der Pijl, 1979). In *Frailea* species, flowering is highly seasonal and seems to be
2765 related to temperature, humidity and day length (Köhler, 1968; Metzing & Kiesling, 2006;
2766 Machado, 2007; Gomes *et al.*, 2019). In nature, the flowering of *F. gracillima* occurs during
2767 months with longer daytime (13-14 sunlight hours), high sunlight intensity, very hot
2768 temperatures at noon and in the early afternoon when the plants bloom (12:30h -15:00h)
2769 (INMET, 2018). Moreover, days with rain are common in the flowering period. The cacti
2770 collections in greenhouses or in higher latitudes, where most *Frailea* individuals are cultivated,
2771 experience rather different environmental conditions. This might strongly influence petal
2772 unfolding in the species of *Frailea*. Thus, it has been suspected that *Frailea* species cultivated
2773 in warmer countries result in higher frequency of chasmogamous flower production. Moreover,
2774 when grown in shaded greenhouses, flowers seem to remain closed and set fruit (Köhler, 1968).
2775 Laboratory tests under controlled conditions may identify which abiotic factors influences the
2776 production of chasmogamous and cleistogamous flowers in *Frailea* species.

2777

2778 *Conclusions*

2779

2780 Although *Frailea* is considered a typically cleistogamous genus, *F. gracillima* presents floral
2781 traits that favor cross-pollination. Chasmogamous flower production was favored in natural
2782 habitat during three years of study. However, movements of stigma and stamens during anthesis
2783 promoted delayed autonomous self-pollination when there was no more chance of cross
2784 pollination due flower closure, which probably ensuring sexual reproduction in adverse
2785 conditions and low pollinator environments.

2786 REFERENCES

2787

2788 **Anderson EF.** 2001. *The cactus family*. Timber Press (OR). 776 pp.2789 **Augspurger CK.** 1981. Reproductive synchrony of a tropical shrub: experimental studies on
2790 effects of pollinators and seed predators in *Hybanthus prunifolius* (Violaceae). *Ecology* **62**:
2791 775-788.2792 **Barthlott W, Hunt DR.** 1993. Cactaceae. pp 161 – 197. In: **Kubitzki K, Rohwer JG, Bittrich**
2793 **V.** (eds) *Flowering Plants Dicotyledons. The Families and Genera of Vascular Plants*, v.2
2794 Springer, Berlin, Heidelberg. 653 pp.2795 **Britton NL, Rose JN.** 1922. *The Cactaceae: descriptions and illustrations of plants of the*
2796 *cactus family*. Vol. 3. 208-211. The Carnegie Institution of Washington. 320 pp.2797 **Brown WV.** 1952. The relation of soil moisture to cleistogamy in *Stipa leucotricha*. *Botanical*
2798 *Gazette* **113**: 438-444.2799 **Buttrose MS, Grant WJR, Lott JNA.** 1977. Reversible curvature of style branches of
2800 *Hibiscus trionum* L., a pollination mechanism. *Australian Journal of Botany* **25**: 567-570.2801 **Carneiro AM, Farias-Singer R, Ramos RA, Nilson AD.** 2016. Cactos do Rio Grande do Sul.
2802 Fundação Zoobotânica do Rio Grande do Sul, Porto Alegre. 224 pp.2803 **Cerceau I, Siriani-Oliveira S, Dutra AL, Oliveira R, Schlindwein C.** 2019. The cost of
2804 fidelity: foraging oligolectic bees gather huge amounts of pollen in a highly specialized
2805 cactus–pollinator association. *Biological Journal of the Linnean Society* **128**: 30-43.2806 **Cota-Sánchez JH, Almeida OJG, Falconer DJ, Choi HJ, Bevan L.** 2013. Intriguing
2807 thigmonastic (sensitive) stamens in the Plains Prickly Pear *Opuntia polyacantha*
2808 (Cactaceae). *Flora - Morphology, Distribution, Functional Ecology of Plants* **208**: 381-389.2809 **Darwin C.** 1862. On the various contrivances by which British and foreign orchids are
2810 fertilized. *Murray, London*, 365 pp.2811 **de Jong TJ, Waser NM, Klinkhamer PG.** 1993. Geitonogamy: the neglected side of selfing.
2812 *Trends in Ecology & Evolution* **8**: 321-325.2813 **Dutra AL, Schlindwein C, Oliveira R.** 2020. Females of a solitary bee reject males to collect
2814 food for offspring. *Behavioral Ecology* **31**: 884–891.2815 **Eggli U, Giorgetta M.** 2017. Flowering phenology and observations on the pollination biology
2816 of South American cacti. 3. Temporally robust sequential “big bang” flowering of two
2817 unrelated sympatric globular cacti in northern Argentina. *Haseltonia* **23**: 97-109.

- 2818 **Faegri K, Van der Pijl L.** 1979. *The Principles of Pollination Ecology*. 3rd edn. Pergamon.
2819 244 pp.
- 2820 **Glavich T.** 2016. Beginner's Guide to *Frailea*. *Cactus and Succulent Journal* **88**: 144-147.
- 2821 **Gomes VGN, Valiente-Banuet A, Araujo AC.** 2019. Reproductive phenology of cacti species
2822 in the Brazilian Chaco. *Journal of Arid Environments* **161**: 85-93.
- 2823 **Grant V, Hurd P.** 1979. Pollination of the southwestern Opuntias. *Plant Systematics and*
2824 *Evolution* **133**: 15-28.
- 2825 **INMET.** 2018. Banco de Dados Meteorológicos para Ensino e Pesquisa - BDMEP. Dados
2826 históricos da estação meteorológica 83980 - Bagé - RS entre 1988 e 2018.
- 2827 **Klips RA, Snow AA.** 1997. Delayed autonomous self-pollination in *Hibiscus laevis*
2828 (Malvaceae). *American Journal of Botany* **84**: 48-53.
- 2829 **Köhler U.** 1968. Fraileen – und wie lange sie blühen. *Kakteen und Andere Sukkulente* **19**:
2830 166–167.
- 2831 **Langer RHM, Wilson D.** 1965. Environmental control of cleistogamy in prairie grass (*Bromus*
2832 *unioloides* HBK). *New Phytologist* **64**: 80-85.
- 2833 **Lenzi M, Orth AI.** 2011. Visitantes florais de *Opuntia monacantha* (Cactaceae) em originais
2834 de Florianópolis, SC, Brasil. *Acta Biológica Paranaense* **40**: 19-32.
- 2835 **Lloyd DG, Schoen DJ.** 1992. Self-and cross-fertilization in plants. I. Functional dimensions.
2836 *International Journal of Plant Sciences* **153**:358-369.
- 2837 **Lord EM.** 1981. Cleistogamy: a tool for the study of floral morphogenesis, function and
2838 evolution. *The Botanical Review* **47**: 421-449.
- 2839 **Machado M.** 2007. Fascinating *Frailea*, Part I: General impressions. *Cactus World* **25**: 5-11.
- 2840 **Mandujano MdC, Carrillo-Angeles I, Martínez-Peralta C, Golubov J.** 2010. Reproductive
2841 biology of Cactaceae. pp. 197-230. In: **Ramawat KG.** Eds *Desert plants: biology and*
2842 *biotechnology*. Berlin: Heidelberg, Springer-Verlag. 503pp.
- 2843 **Marshall WT, Bock TM.** 1941. Cactaceae: With Illustrated Keys of All Tribes, Sub-tribes and
2844 Genera. Abbey Garden Press. 227 pp.
- 2845 **Metzing D, Kiesling R.** 2006. Notes on the diversity, biology, and taxonomy of *Frailea*
2846 (Cactaceae). *Bradleya* **24**: 115-128.
- 2847 **Metzing D.** 2015a. Merveilleuses petites plantes, le genre *Frailea*. *Terra Seca* **3**: 4–17.
- 2848 **Metzing D.** 2015b. Merveilleuses petites plantes, le genre *Frailea*. *Terra Seca* **4**: 42–52.
- 2849 **Oliveira R, Schlindwein C.** 2010. Experimental demonstration of alternative mating tactics of
2850 male *Ptilothrix fructifera* (Hymenoptera, Apidae). *Animal Behaviour* **80**: 241-247.

- 2851 **Oliveira R, Carvalho AT, Schlindwein C.** 2012. Territorial or wandering: how males of
 2852 *Protodiscelis palpalis* (Colletidae, Paracolletinae) behave in searching for mates. *Apidologie*
 2853 **43**: 674-684.
- 2854 **Oliveira R, Carvalho AT, Schlindwein C.** 2013. Plasticity in male territoriality of a solitary
 2855 bee under different environmental conditions. *Journal of insect behavior* **26**: 690-694.
- 2856 **Opler PA, Frankie GW, Baker HG.** 1976. Rainfall as a factor in the release, timing, and
 2857 synchronization of anthesis by tropical trees and shrubs. *Journal of Biogeography* **3**: 231-
 2858 236.
- 2859 **Pontes RC, Deble ASDO, Deble, LP.** 2018. *Frailea erythracantha* (Cactaceae): a new species
 2860 from Pampa Grasslands, Rio Grande do Sul State, Brazil. *Phytotaxa* **369**: 211-218.
- 2861 **Porsch O.** 1938. Deutsche Kakteen-Gesellschaft. Das Bestäubungsleben der Kakteenblüte.
- 2862 **Putnam EW.** 1968. The Genus *Frailea*. *The National Cactus and Succulent Journal* **23**: 54-
 2863 56.
- 2864 **Ribeiro-Silva S, Zappi DC, Taylor N, Machado M.** 2011. Plano de ação nacional para a
 2865 conservação das cactáceas. Série Espécies Ameaçadas, 24.
- 2866 **Richards AJ.** 1997. *Plant breeding systems*. Garland Science. 529 pp.
- 2867 **Rowley G.** 1980. Pollination syndromes and cactus taxonomy. *The Cactus and Succulent*
 2868 *Journal of Great Britain*, 42: 95-98.
- 2869 **Ruan C, Qin P, Han R.** 2005. Strategies of delayed self-pollination in *Kosteletzkya virginica*.
 2870 *Chinese Science Bulletin* **50**: 94-96.
- 2871 **Ruan C, Li H, Mopper S.** 2008. The impact of pollen tube growth on stigma lobe curvature in
 2872 *Kosteletzkya virginica*: the best of both worlds. *South African Journal of Botany* **74**: 65-70.
- 2873 **Schlindwein C, Wittmann D.** 1995. Specialized solitary bees as effective pollinators of South
 2874 Brazilian species of *Notocactus* and *Gymnocalycium* (Cactaceae). *Bradleya* **13**: 25-34.
- 2875 **Schlindwein C, Wittmann D.** 1997. Stamen movements in flowers of *Opuntia* (Cactaceae)
 2876 favour oligolectic pollinators. *Plant Systematics and Evolution* **204**: 179-193.
- 2877 **Schlindwein C.** 1998. Frequent Oligolecty Characterizing a Diverse Bee–Plant Community in
 2878 a Xerophytic Bushland of Subtropical Brazil. *Studies on Neotropical Fauna and*
 2879 *Environment* **33**: 46-59.
- 2880 **Schoen DJ, Lloyd DG.** 1984. The selection of cleistogamy and heteromorphic
 2881 diaspores. *Biological Journal of the Linnean Society* **23**: 303-322.
- 2882 **Shetler SG.** 1979. Pollen-collecting hairs of *Campanula* (Campanulaceae), I: Historical
 2883 review. *Taxon* **28**: 205-215.

- 2884 **Sprengel CK.** 1793. Das entdeckte Geheimnis der Natur. *Wilhelm Engelmann, Leipzig.*
- 2885 **Sun SG, Guo YH, Gituru RW, Huang SQ.** 2005. Corolla wilting facilitates delayed
2886 autonomous self-pollination in *Pedicularis dunniana* (Orobanchaceae). *Plant Systematics*
2887 *and Evolution* **251**: 229-237.
- 2888 **Taylor NP.** 1991. The genus *Melocactus* (Cactaceae) in Central and South America. *Bradleya*
2889 **9**: 1-80.
- 2890 **Uphof JT.** 1938. Cleistogamic flowers. *The Botanical Review* **4**: 21-49.
- 2891 **Putnam EW.** 1968. The Genus *Frailea*. *The National Cactus and Succulent Journal* **23**: 54-
2892 56.
- 2893 **Webb CJ, Lloyd DG.** 1986. The avoidance of interference between the presentation of pollen
2894 and stigmas in angiosperms II. Herkogamy. *New Zealand journal of botany* **24**: 163-178.
2895

2896 TABLES

2897 **Table 1:** Flower-visiting bees of *Frailea gracilima*. Frequency of floral visits were measured during 30
 2898 hours of observation (N = 495 visits). (-) Species not recorded during 30 hours of flower observation,
 2899 but collected during fieldwork.

2900

Bee species	Tribe / Subfamily	Sex	Frequency (%)
Andrenidae			
<i>Arhysosage cactorum</i> Moure 1999	Calliopsini	♀ ♂	3.3%
<i>Callonychium</i> sp.	Calliopsini	♀ ♂	1%
<i>Anthrenoides micans</i> Urban 1955	Protandrenini	♀ ♂	70.7%
<i>Psaenythia</i> sp.	Protandrenini	♀	-
Apidae			
<i>Ancyloscelis romeroi</i> Holmberg 1903	Emphorini	♀	-
<i>Ceratina</i> sp. 1	Ceratinini	♀	-
<i>Ceratina</i> sp. 2	Ceratinini	♀	-
<i>Chalepogenus goeldianus</i> Friese, 1899	Tapinotaspidini	♀	2.2%
<i>Plebeia</i> sp.	Meliponini	♀	-
<i>Tapinotaspis</i> sp.	Tapinotaspidini	♀	-
Colletidae			
<i>Cactocolletes rugatus</i> Urban 1995	Neopasiphaeinae	♀	0.2%
Halictidae			
<i>Augochloropsis</i> sp.	Halictini	♀	-
<i>Dialictus</i> sp.	Halictini	♀	22.6%

2901 **FIGURES**

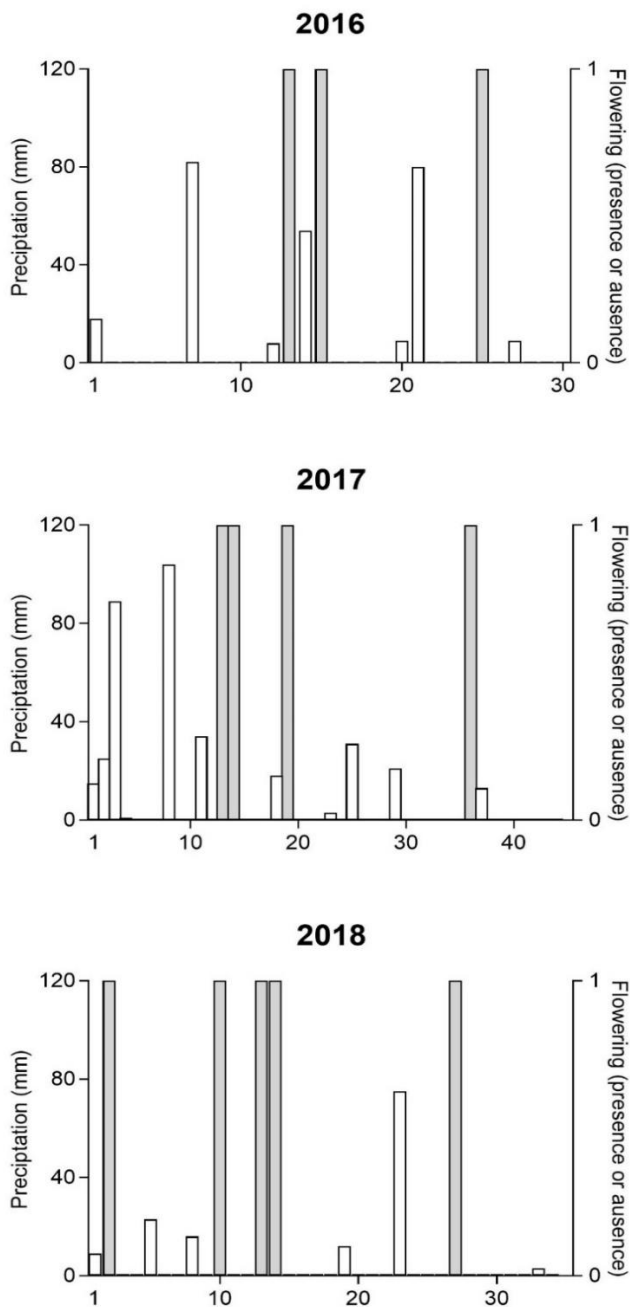
2902



2903

2904 **Figure 1:** *Frailea gracillima* habitat and synchronized flower opening in the field. Cacti grow between
2905 rock cracks in sandstone outcrops in Santana da Boa Vista, Rio Grande do Sul.

2906



2907

2908 **Figure 2:** Daily variation in precipitation and flowering events of *Frailea gracillima* during three years.

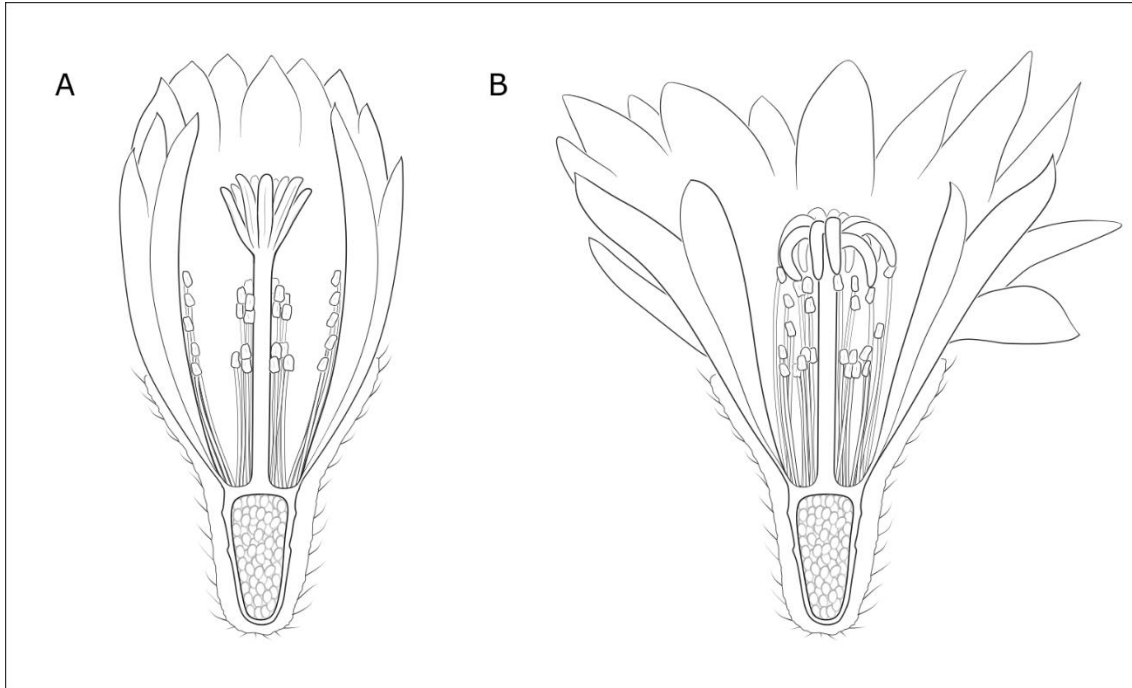
2909 The study period in each year was: 2016 (October 27 to November 25), 2017 (October 11 to November

2910 23) and 2018 (October 26 to November 28).

2911 Daily rainfall (mm) represented by white bars, varied between 0 and 104 mm in three years. The events

2912 of presence or absence of flowering are represented by the gray bars. Flowering occurred from 1 to 7

2913 days after precipitation.



2914

2915

2916

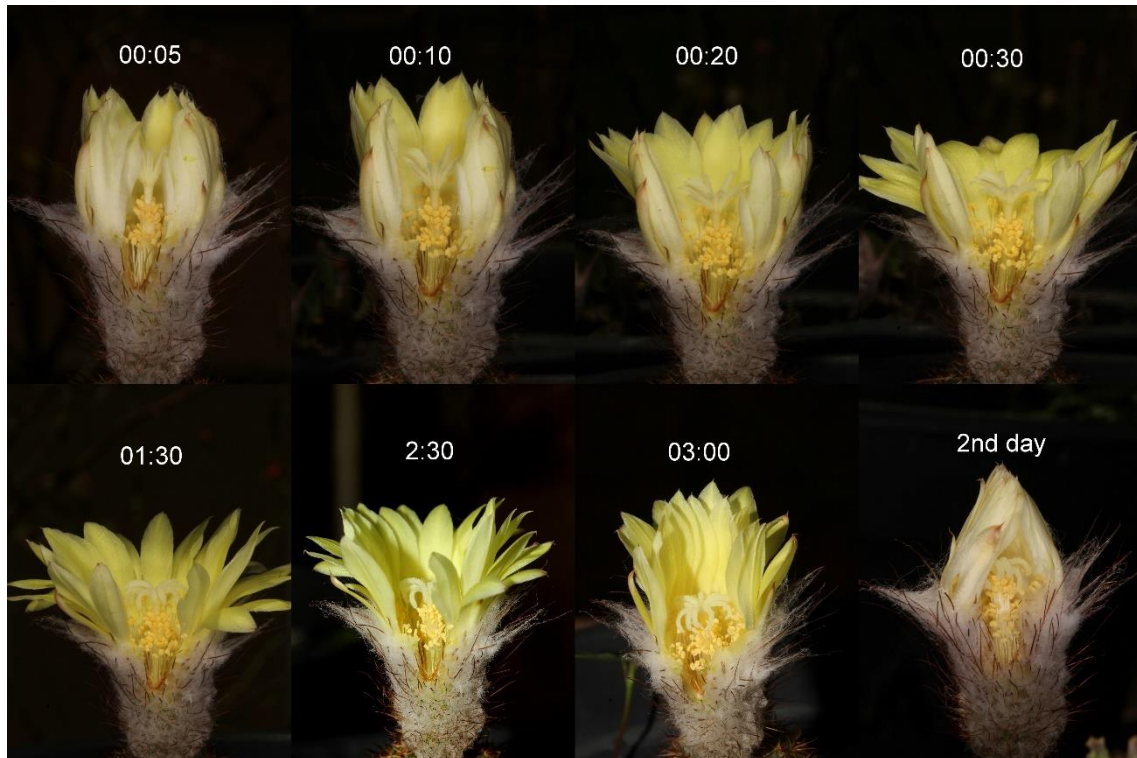
2917

2918

2919

2920

Figure 3: Movement of reproductive organs during the anthesis. (A) In the first minutes of anthesis, the stigmatic lobes are positioned vertically and the stamens are positioned in two main positions: in contact with the petals or in contact with the style. (B) At the end of anthesis, approximately 2:30 hours after opening the flowers, the stigmatic lobes move towards the stamens and come into contact with the anthers. Stamens move gradually along the anthesis and are distributed along the floral tube.



2921

2922 **Figure 4:** Movement of stigmatic lobes and stamens during anthesis of *Frailea gracilima*. The schedules
 2923 in the photo correspond to the time elapsed since the beginning of the flower opening. During the
 2924 beginning of anthesis, there is no contact between stamens and stigma. Throughout the anthesis, the
 2925 structures move: stigmatic lobes curl up and the stamens spread to the center of the flowers. Flower at
 2926 the time of 3:00 had the petals partially open with hands to demonstrate the position of the stamens and
 2927 stigma inside the flower. On the second day of anthesis, the reproductive structures are in dense contact.
 2928



2929

2930 **Figure 5:** Contact of stigmatic lobes with anthers in *Frailea gracillima* flowers. (A) Flowers in the end
 2931 of anthesis shows intense contact of the stigmatic lobes with anthers. (B) Stigmatic lobes with pollen
 2932 grains in flower not visited by bees.

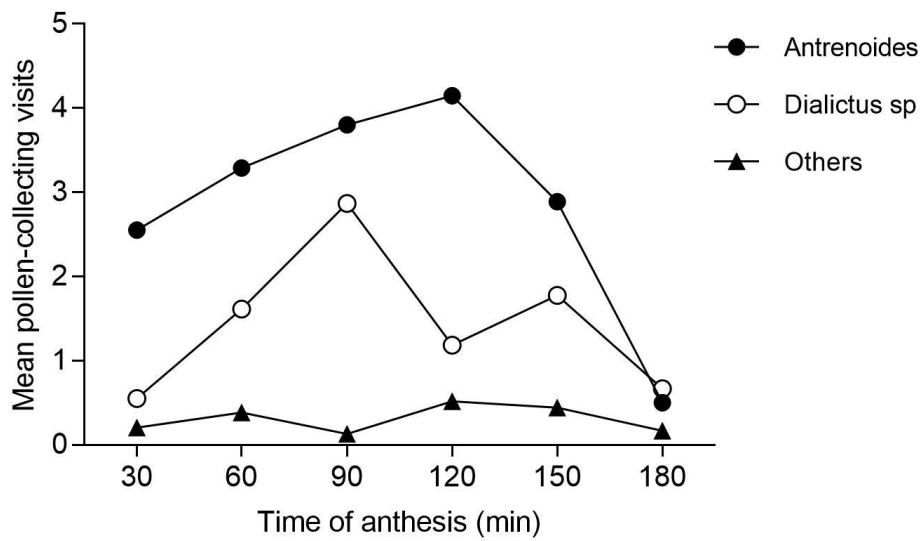
2933



2934

2935 **Figure 6:** Main floral visitors of *Frailea gracillima*. (A) *Anthrenoides micans*. (B) *Dialictus sp.*

2936



2937

2938 **Figure 7:** Frequency of visits by females to *Frailea gracillima* flowers throughout anthesis. Time of
 2939 anthesis was represented on the Y axis through the minutes that elapsed after the opening of the flower
 2940 (time 0) and flower closure after 150 ± 13 min. We represent an anthesis up to 180 minutes to
 2941 characterize all floral visitors until the complete petal closure. The X axis represents the average number
 2942 of visits by female bees received on a flower. Total number of visits recorded was 365 in 15 flowers.
 2943 Male visits were excluded.

2944



2945

2946 **Figure 8:** Bee positioned between the two bundles of stamens to collect pollen. *Anthrenoides micans*
2947 preferably collects pollen from the stamens attached to the style.

2948

2949 CONSIDERAÇÕES FINAIS

2950

2951 A observação detalhada das espécies em habitat natural é essencial para a construção do
2952 conhecimento sobre as interações entre plantas e polinizadores. A imersão em campo durante
2953 o desenvolvimento desta tese revelou aspectos únicos das interações, como a sincronia temporal
2954 da atividade das espécies envolvidas, funcionamento floral e comportamento de forrageio das
2955 abelhas nas flores complexas dos cactos. Devido ao fato das flores de cactos serem efêmeras,
2956 muitas vezes com períodos curtos de floração, estudos minuciosos sobre interações em habitat
2957 natural são escassos.

2958 Estudamos detalhadamente duas espécies de cactáceas melitófilas pertencentes a dois
2959 gêneros. *Parodia*, um gênero que já era conhecido por manter relações com abelhas
2960 especialistas e *Frailea*, um gênero considerado majoritariamente cleistógamo que nunca foi
2961 estudado na perspectiva reprodutiva em habitat natural.

2962 Em relação a *Parodia neohorstii*, que floresce em apenas alguns dias no ano,
2963 observamos que as abelhas sincronizaram a atividade de voo com abertura das flores e as
2964 polinizaram na primeira hora da antese. A eficiência na polinização é essencial para a
2965 reprodução de *P. neohorstii*, uma vez que a espécie é auto incompatível. O desencontro
2966 fenológico registrado entre o início da floração de *P. neohorstii* e a atividade de seus
2967 polinizadores efetivos foi uma oportunidade de quantificar qual é o real impacto da ausência
2968 das abelhas oligoléticas na reprodução da espécie. A drástica redução na produção de frutos e
2969 sementes na ausência de *Arhysosage cactorum* indica que *P. neohorstii* é dependente de
2970 polinizadores específicos para se reproduzir e caso o desencontro temporal entre as espécies se
2971 torne mais frequente, podem ocorrer prejuízos na reprodução dessa cactácea em habitat natural.

2972 Um fator chave do funcionamento floral de *P. neohorstii* é o movimento tigonástico
2973 dos estames. Este movimento age como um mecanismo de particionamento de pólen,
2974 restringindo a quantidade de recurso coletado pelos visitantes florais, uma vez que as flores
2975 oferecem todo o pólen de uma única vez. Estes movimentos, que são induzidos pelos
2976 polinizadores efetivos durante a coleta de pólen, atua como gatilho para a senescência floral.
2977 Após consecutivos movimentos de estames durante a antese, induzidos de forma experimental,
2978 as flores senesceram mesmo na ausência de polinização. Isso indica que o movimento dos
2979 estames pode agir como um sinalizador da presença dos polinizadores efetivos, e,
2980 consequentemente, da dispersão polínica. A ausência de movimentos pode indicar um ambiente
2981 com baixa disponibilidade de polinizadores, fator que induz a extensão da longevidade floral e

2982 colabora para o aumento das chances de exportação de polínica. Esses movimentos já foram
2983 reportados em várias espécies de cactáceas, mas até o momento não foram descritas funções
2984 relacionadas com a indução da senescência floral.

2985 *Parodia neohorstii* é uma espécie criticamente ameaçada de extinção e em declínio
2986 populacional. O diagnóstico reprodutivo em habitat natural revelou que a espécie está
2987 produzindo frutos e sementes com muita eficiência, o que pode contribuir para a manutenção
2988 das populações a longo prazo. Porém, a espécie está sendo impactada negativamente pela
2989 criação de animais na área de ocorrência. Os danos físicos causados pelo consumo dos cactos
2990 por caprinos e pelo pisoteio por bovinos, geraram altas taxas de mortalidade nas populações
2991 estudadas. Esse fator pode impactar fortemente na viabilidade das populações em habitat
2992 natural a médio e longo prazo e contribuir para o declínio populacional. O impacto gerado pelos
2993 animais de criação nas populações de *P. neohorstii* pode ser semelhante ou até pior em
2994 populações de cactos simpátricos, como espécies dos gêneros *Parodia*, *Frailea*,
2995 *Gymnocalycium* e *Echinopsis*, que muitas vezes possuem espinhos mais macios e menos
2996 numerosos, o que aumenta a vulnerabilidade das espécies aos herbívoros. Uma ação para a
2997 conservação das cactáceas da região seria a restrição do acesso de animais nas áreas de
2998 ocorrência, principalmente em afloramentos rochosos, onde a oferta de alimentos para animais
2999 de criação é baixa e a restrição não prejudicaria na qualidade da pastagem disponível para os
3000 animais.

3001 Em relação a *Frailea gracillima*, observamos que a cleistogamia não ocorreu em habitat
3002 natural durante o período de estudo e as flores apresentam características que favorecem a
3003 polinização cruzada. A grande quantidade de pólen co-específico depositado no estigma por
3004 abelhas contribuiu para a formação de frutos e sementes. Observamos que *F. gracillima* pode
3005 promover autopolinização autônoma tardia, possibilitada após o movimento dos estames e
3006 estigma ao longo da antese. Esse fator é importante, pois contribui para a produção de frutos e
3007 sementes mesmo em cenários de baixa densidade de polinizadores, o que não ocorre na espécie
3008 simpátrica *P. neohorstii*.

3009 As espécies de cactáceas estudadas não produzem néctar. Por isso, a manutenção dos
3010 polinizadores no ambiente depende de outras espécies vegetais para fornecimento desse
3011 recurso. Dessa maneira, a conservação da comunidade vegetal do entorno é essencial para a
3012 qualidade reprodutiva dos cactos da região, uma vez que as mesmas espécies de abelha são
3013 encontradas em flores de diversas espécies de cactos melitófilos simpátricos.

3014 Em síntese, este trabalho gerou conhecimento sobre aspectos da história natural de duas
3015 espécies de cactáceas. Contribuímos com a descrição de novos mecanismos de funcionamento
3016 floral em ambas as espécies estudadas, além de diagnosticar que a formação de frutos e
3017 sementes está ocorrendo com sucesso em habitat natural. Em relação a *Parodia neohorstii*,
3018 identificamos duas principais ameaças à espécie: 1) mudanças fenológicas podem alterar a
3019 interação com polinizadores e causar impactos negativos na reprodução em habitat natural; 2)
3020 a mortalidade de cactos é provocada principalmente pelos impactos gerados por animais de
3021 criação.

3022 Estes resultados podem contribuir com ações direcionadas para a conservação, tanto das
3023 espécies foco deste estudo, quanto para outras cactáceas da região das Guaritas.