UNIVERSIDADE FEDERAL DE MINAS GERAIS Instituto de Ciência Biológicas Programa de Pós-graduação em Genética

Pedro Heringer Lisboa Teixeira

ORIGEM E EVOLUÇÃO DOS HELITRONS

Belo Horizonte 2022 Pedro Heringer Lisboa Teixeira

Origem e Evolução dos Helitrons

Tese apresentada ao Programa de Pós-Graduação em Genética da Universidade Federal de Minas Gerais como requisito parcial à obtenção do título de Doutor em Genética.

Orientador: Prof. Dr. Gustavo Campos e Silva Kuhn

Belo Horizonte

2022

043 Teixeira, Pedro Heringer Lisboa. Origem e evolução dos Helitrons [manuscrito] / Pedro Heringer Lisboa Teixeira. – 2022.

160 f. : il. ; 29,5 cm.

Orientador: Prof. Dr. Gustavo Campos e Silva Kuhn. Tese (doutorado) – Universidade Federal de Minas Gerais, Instituto de Ciências Biológicas. Programa de Pós-Graduação em Genética.

1. Genética. 2. Elementos de DNA transponíveis. I. Kuhn, Gustavo Campos e Silva. II. Universidade Federal de Minas Gerais. Instituto de Ciências Biológicas. III. Título.

CDU: 575

Ficha catalográfica elaborada pela bibliotecária Fabiane C M Reis – CRB 6 – 2680



UNIVERSIDADE FEDERAL DE MINAS GERAIS Instituto de Ciências Biológicas Programa de Pós-Graduação em Genética

ATA DE DEFESA DE DISSERTAÇÃO / TESE

ATA DA DEFEGA DE TEGE	153/2022
AIA DA DEFESA DE TESE	entrada
	2°/2017
Pedro Heringer Lisboa Teixeira	CPF: 083.643.596-65

As oito horas e trinta minutos do dia 24 de fevereiro de 2022, reuniu-se remotamente (rede mundial de computadores), a Comissão Examinadora de Tese, indicada pelo Colegiado do Programa, para julgar, em exame final, o trabalho intitulado: "Origem e Evolução dos Helitrons", requisito para obtenção do grau de Doutor em Genética. Abrindo a sessão, o Presidente da Comissão, Gustavo Campos e Silva Kuhn, após dar a conhecer aos presentes o teor das Normas Regulamentares do Trabalho Final, passou a palavra ao candidato, para apresentação de seu trabalho. Seguiu-se a arguição pelos Examinadores, com a respectiva defesa do candidato. Logo após, a Comissão se reuniu, sem a presença do candidato e do público, para julgamento e expedição de resultado final. Foram atribuídas as seguintes indicações:

Prof./Pesq.	Instituição	CPF	Indicação
Gustavo Campos e Silva Kuhn	UFMG	260.136.648-62	Aprovado
Elgion Lucio da Silva Loreto	UFSM	324127700-34	Aprovado
Claudia Marcia Aparecida Carareto	UNESP	785924538-87	Aprovado
Leonardo Barbosa Koerich	UFMG	033.549.409-99	Aprovado
Renato Santana de Aguiar	UFMG	000.086.336-06	Aprovado

Pelas indicações, o candidato foi considerado: APROVADO.

O resultado final foi comunicado publicamente ao candidato pelo Presidente da Comissão. Nada mais havendo a tratar, o Presidente encerrou a reunião e lavrou a presente ATA, que será assinada por todos os membros participantes da Comissão Examinadora.

Belo Horizonte, 24 de fevereiro de 2022.

Gustavo Campos e Silva Kuhn - UFMG

Elgion Lucio da Silva Loreto - UFSM

Claudia Marcia Aparecida Carareto - UNESP

Leonardo Barbosa Koerich - UFMG

Renato Santana de Aguiar - UFMG

Assinatura dos membros da banca examinadora:

SEI/UFMG - 1273169 - Ata de defesa de Dissertação/Tese

https://sei.ufmg.br/sei/controlador.php?acao=documento_imprimir_w...

seil assinatura	ocumento assinado eletronicamente por Claudia Marcia Aparecida Carareto, Usuário Externo , em 24/02/2022, às 13:35,
eletrônica	onforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u> .
seil	ocumento assinado eletronicamente por Gustavo Campos e Silva Kuhn, Professor do Magistério Superior, em
assinatura	4/02/2022, às 13:53, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de</u>
eletrônica	ovembro de 2020.
seil	ocumento assinado eletronicamente por Renato Santana de Aguiar, Professor do Magistério Superior, em 24/02/2022, às 3:15, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u> .
seil	ocumento assinado eletronicamente por Leonardo Barbosa Koerich, Professor do Magistério Superior , em 27/02/2022,
assinatura	s 20:15, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de</u>
eletrônica	020.
seil assinatura eletrónica	ocumento assinado eletronicamente por Élgion Lúcio da Silva Loreto, Usuário Externo , em 04/03/2022, às 17:33, onforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u> .
	autenticidade deste documento pode ser conferida no site <u>https://sei.ufmg.br</u> ei <u>/controlador_externo.php?acao=documento_conferir&id_orgao_acesso_externo=0</u> , informando o código verificador 73169 e o código CRC 268CA550.

Referência: Processo nº 23072.210628/2022-97

SEI nº 1273169

SEI/UFMG - 1273176 - Folha de Aprovação

https://sei.ufmg.br/sei/controlador.php?acao=documento_imprimir_w...



UNIVERSIDADE FEDERAL DE MINAS GERAIS Instituto de Ciências Biológicas Programa de Pós-Graduação em Genética

FOLHA DE APROVAÇÃO

"Origem e Evolução dos Helitrons"

Pedro Heringer Lisboa Teixeira

Tese aprovada pela banca examinadora constituída pelos Professores:

Gustavo Campos e Silva Kuhn UFMG

Elgion Lucio da Silva Loreto UFSM

Claudia Marcia Aparecida Carareto UNESP

> Leonardo Barbosa Koerich UFMG

> Renato Santana de Aguiar UFMG

Belo Horizonte, 24 de fevereiro de 2022.

1.2	- ail	
1.5	sel:	p
as	sinatura	14
el	etrônica	

Documento assinado eletronicamente por **Claudia Marcia Aparecida Carareto, Usuário Externo**, em 24/02/2022, às 13:35, conforme horário oficial de Brasília, com fundamento no art. 5º do Decreto nº 10.543, de 13 de novembro de 2020.

seľ

Documento assinado eletronicamente por **Gustavo Campos e Silva Kuhn, Professor do Magistério Superior**, em 24/02/2022, às 13:52, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u>.



Documento assinado eletronicamente por **Renato Santana de Aguiar**, **Professor do Magistério Superior**, em 24/02/2022, às 18:15, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u>.



Documento assinado eletronicamente por **Leonardo Barbosa Koerich**, **Professor do Magistério Superior**, em 27/02/2022, às 20:15, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto nº 10.543, de 13 de novembro de 2020</u>.



Documento assinado eletronicamente por Élgion Lúcio da Silva Loreto, Usuário Externo, em 04/03/2022, às 17:33, conforme horário oficial de Brasília, com fundamento no art. 5º do <u>Decreto</u> nº 10.543, de 13 de novembro de 2020.



A autenticidade deste documento pode ser conferida no site <u>https://sei.ufmg.br</u> /<u>sei/controlador_externo.php?acao=documento_conferir&id_orgao_acesso_externo=0,</u> informando o código verificador **1273176** e o código CRC **4FBADABC**.

Referência: Processo nº 23072.210628/2022-97

SEI nº 1273176

AGRADECIMENTOS

Gostaria de agradecer primeiramente ao Professor Gustavo C. S. Kuhn que contribuiu de forma essencial não só para a concretização deste trabalho, mas também para a minha formação acadêmica e profissional. Sua orientação desde a graduação, passando pelo mestrado, e agora no doutorado, foi fundamental para que eu pudesse me tornar o cientista que sou hoje. Obrigado por ser o melhor mentor possível e sempre confiar no meu potencial.

Agradeço aos membros atuais e os que já passaram pelo Laboratório de Citogenômica Evolutiva. Além de todas as discussões científicas interessantes, o companheirismo e a amizade de vocês ajudaram a tornar minha jornada acadêmica mais leve e agradável.

Muito obrigado à CAPES por financiar todo o período do meu mestrado e doutorado, contribuindo para que eu me mantivesse fazendo o que mais gosto na vida.

Também não poderia deixar de agradecer aos meus amigos que estão fora do meio acadêmico. Apesar de não entenderem muito bem o que eu faço, vocês sempre têm uma palavra de apoio e reconhecimento sobre o meu trabalho, e mesmo com a minha ausência em vários momentos, vocês se mantiveram companheiros.

Sou extremamente grato a todos da minha numerosa família. Aos avôs e avós que se foram, mas deixaram para mim o exemplo de vida e caráter a ser seguido, e minha querida avó que, em vida, continua a me ensinar o que de fato é importante, mesmo estando longe. Obrigado aos meus tios, tias e primos que me fazem sentir especial com tanto amor. Agradeço à minha irmã Laura, que considero um exemplo de sucesso e, mesmo estando distante, sempre foi uma das primeiras pessoas a torcer por mim.

Em especial, agradeço a minha amada esposa Isabela que tem sido uma fonte de suporte, alegria e amor constante nestes anos que se passaram. Mesmo sem ter participado diretamente, o fruto deste trabalho também é seu. Eu jamais teria chegado até aqui sem você.

Por fim, minha gratidão infinita ao meu pai e minha mãe, meus heróis e maiores inspirações. É impossível imaginar qualquer conquista na minha vida sem a participação deles. Não só este trabalho, mas a minha vida é dedicada a vocês dois.

"Nothing in biology makes sense except in the light of evolution"

Theodosius G. Dobzhansky

Resumo

Elementos de transposição (TEs) são sequências de DNA móveis e abundantes em genomas procarióticos e eucarióticos. Em eucariotos, TEs podem ser divididos em duas classes, denominadas classe I, que utilizam intermediários de RNA para se transporem, e classe II, que utilizam intermediários de DNA. Cada uma destas classes compreende diferentes subclasses, que por sua vez são divididas em superfamílias e famílias. Helitrons representam uma subclasse de elementos dentro da classe II que se transpõem por meio de um mecanismo único em eucariotos, sendo encontrados em todos os principais grupos taxonômicos deste domínio da vida. Estes transposons impactam genomas eucarióticos por ocuparem frações consideráveis do DNA de seus hospedeiros, além de estarem envolvidos na mobilização e duplicação de fragmentos cromossômicos adjacentes. Embora a compreensão sobre vários aspectos relacionados aos Helitrons tenha avançado consideravelmente nas duas décadas que sucederam a descoberta destes elementos, sua origem evolutiva e detalhes do seu mecanismo de transposição são temas que permaneceram amplamente inexplorados durante o mesmo período. Neste trabalho, investigamos a origem dos Helitrons através de análises evolutivas dos dois domínios principais presentes na sua transposase. Os resultados das análises de cada domínio revelam aspectos distintos, porém complementares, sobre a origem dos Helitrons. Em conjunto, nossos achados indicam que estes elementos descendem de plasmídeos procarióticos que, após invadirem genomas eucarióticos, passaram a utilizar a transposição como mecanismo de replicação em seus hospedeiros. Este cenário se opõe às principais hipóteses apresentadas até o momento para explicar a origem dos Helitrons e dos domínios da sua transposase. Além disso, com base nas evidências obtidas neste trabalho e em outros estudos, propomos que a transposase dos Helitrons desempenha funções catalíticas mais complexas do que havia sido sugerido anteriormente. Por fim, nossa investigação paralela sobre a evolução de uma família de Helitrons presente em artrópodes ilustra a capacidade notável destes transposons invadirem novos genomas hospedeiros por meio de transferências horizontais que podem ocorrer entre ordens ou mesmo classes distintas de organismos.

Palavras-chave: Helitrons. Elementos de transposição. Transposon. Transferência horizontal.

Abstract

Transposable elements (TEs) are mobile DNA sequences found in a large number of copies in prokaryotic and eukaryotic genomes. In eukaryotes, TEs can be divided into two classes, named class I, which use RNA intermediates to transpose, and class II, which use DNA intermediates. Each one of these classes include different subclasses, which in turn are divided into superfamilies and families. Helitrons represent a subclass of elements within class II that transpose by a mechanism that is unique in eukaryotes, being found in all major taxonomic groups from this domain of life. These transposons impact eukaryotic genomes by occupying considerable DNA fractions of their hosts, also being involved in the mobilization and duplication of adjacent chromosomal fragments. Although the understanding about several aspects related to Helitrons has advanced considerably in the two decades that followed their discovery, their evolutionary origin and details of their transposition mechanism are subjects that remained largely unexplored during the same period. In this work, we investigate the origin of *Helitrons* using evolutionary analyses of the two major domains present in their transposase. The results from the analyses of each domain reveal distinct, albeit complementary, aspects about the origin of *Helitrons*. Together, our findings indicate that these elements descend from procaryotic plasmids that, after invading eukaryotic genomes, started using transposition as the replication mechanism in their hosts. This scenario opposes the main hypotheses that have been advanced to explain the origin of *Helitrons* and the domains of their transposase. Furthermore, based on the evidence provided in this work and other studies, we propose that Helitron transposases execute more complex catalytic functions than it was previously suggested. Finally, our parallel investigation about the evolution of a Helitron family found in arthropods illustrate the marked capacity of these transposons to invade new host genomes through horizontal transfers that can occur between distinct orders or even classes of organisms.

Keywords: *Helitrons*. Transposable elements. Transposon. Horizontal transfer.

LISTA DE ILUSTRAÇÕES

Figura 1. Estrutura geral dos <i>Helitrons</i>	11
Figura 2. Mecanismo proposto para a transposição dos <i>Helitrons</i>	12
Figuras do Capítulo 1	
Figure 1. Modular diversity of HUH endonucleases	18
Figure 2. Phylogenetic analysis of RCRE domain sequences	20
Figure 3. NMDS of evolutionary divergence between RCRE domains	21
Figure 4. Proposed scenario for the origin of <i>Helitrons</i> and other RCR elements	22
Figuras do Capítulo 2	
Figure 1. <i>Helitron</i> structural and coding variants	30
Figure 2. Workflow with the methodology used in our study	31
Figure 3. Maximum-likelihood phylogeny of Pif1-like helicases	32
Figure 4. NMDS plot of Pif1-like helicases	34
Figure 5. Cladogram of plant groups that appear to have lost genomic Pif1 helicases	36
Figure 6. A hypothesis for the evolution of <i>Helitrons</i>	36
Figuras do Capítulo 3	12
Figure 1. Phylogeny of Hel_c35 sequences	48
Figure 2. Geographical distribution of arthropod species containing Hel_c35	50
Figure 3. Hypothesis for HTTs involving Hel_c35 sequences	52

LISTA DE ABREVIATURAS

- 3'-OH Grupo hidroxila presente na extremidade 3' do DNA
- AP apurinic–apyrimidinic ('apurínica-apirimidínica')
- bp base pairs ('pares de base')
- CvBV Cotesia vestalis bracovirus
- dsDNA Double-strand DNA ('DNA dupla-Fita')
- Hel Domínio helicase presente na RepHel
- Hel_c35 Família de Helitrons presente em artrópodes e descoberta no genoma de CvBV
- HGT Horizontal Gene Transfer ('Transferência Horizontal de Gene')
- HT Horizontal Transfer ('Transferência Horizontal')
- HTT Horizontal Transposon Transfer ('Transferência Horizontal de Transposons')
- LTR Long Terminal Repeat ('Repetição Terminal Longa')
- MGE Mobile Genetic Element ('Elemento Genético Móvel')
- MYA Million Years Ago ('Milhão de Anos Atrás')
- NCLDV Nucleocytoplasmic large DNA viruses ('Vírus nucleocitoplasmáticos de DNA grande')
- NMDS Non-metric multidimensional scaling ('Escalonamento Multidimensional Não-Métrico')
- ORF Open Reading Frame ('Fase de Leitura Aberta')
- PCNA Proliferating Cell Nuclear Antigen ('Antígeno Nuclear de Células em Proliferação')
- RC Rolling-Circle ('Círculo-Rolante')
- RCR Rolling-Circle Replication ('Replicação por Círculo-Rolante')
- RCRE RCR endonuclease domain ('Domínio endonuclease utilizado na RCR')
- RCT Rolling-Circle Transposition (Transposição por 'Círculo-Rolante')
- Rep Domínio catalítico central presente na RepHel
- RepHel Transposase dos Helitrons
- S1H Superfamily 1 helicase ('Helicase da superfamília 1')
- S3H Superfamily 3 helicase ('Helicase da superfamília 3')
- SH-aLRT Teste da razão de verossimilhança aproximada com correção Shimodaira-Hasegawa
- TE Transposable Element (Elemento Transponível)

SUMÁRIO

1. INTRODUÇÃO	13
1.1 Elementos de transposição	13
1.1.2 Classificação dos TEs	13
1.2 Helitrons	14
1.2.1 Origem evolutiva dos <i>Helitrons</i>	17
2. OBJETIVOS	19
3. CAPÍTULO 1	20
4. CAPÍTULO 2	31
5. CAPÍTULO 3	48
6. DISCUSSÃO GERAL	61
7. CONCLUSÕES	64
8 REFERÊNCIAS	66
9 ANEXOS	68
9.1 Material suplementar do Capítulo 1	68
9.2 Material suplementar do Capítulo 2	83
9.3 Material suplementar do Capítulo 3	146

1. INTRODUÇÃO

1.1 Elementos de transposição

Elementos de transposição (TEs), são sequências de DNA capazes de se mover nos genomas dos seus hospedeiros, e assim se replicar de maneira independente destes. TEs representam frações consideráveis do DNA de praticamente todos organismos eucariotos, sendo que a proporção ocupada por estes elementos apresenta uma forte correlação com o próprio tamanho genômico de seus hospedeiros. Além de impactar diretamente o tamanho genômico de eucariotos, TEs estão frequentemente associados a mutações, polimorfismos, rearranjos cromossômicos e, em alguns casos, são fonte de fatores moduladores da atividade gênica (revisado em Bourque et al. 2018, Wells & Feschotte 2020).

Apesar de estarem associados a inovações evolutivas benéficas para os seus hospedeiros em alguns casos isolados, os TEs representam entidades genéticas essencialmente 'egoístas' que geralmente evoluem nos genomas em que habitam de forma neutra ou afetando estes negativamente. Por esta razão, é esperado que, com o passar do tempo, linhagens de TEs sejam eliminadas dos seus genomas hospedeiros por seleção negativa e/ou deriva genética em algum momento de sua evolução. De fato, assim como outros parasitas, TEs podem utilizar diferentes estratégias para evadir tais processos que promovem sua eliminação. Entretanto, durante longos períodos evolutivos (dezenas ou centenas de milhões de anos) de transmissão vertical em seus genomas hospedeiros, tais estratégias seriam capazes de apenas adiar a extinção aparentemente inevitável destes elementos (revisado em Schaack et al. 2010).

Ao contrário da herança vertical, o processo conhecido como transferência horizontal (horizontal transfer, HT) ocorre através da transmissão de um segmento de DNA de um organismo para o genoma de outro (Wallau et al. 2018, Van Etten & Bhattacharya 2020), sendo assim uma alternativa para TEs escaparem sua extinção. Deste modo, a HT de TEs para novos genomas hospedeiros representa o principal mecanismo para explicar a persistência destes elementos no longo prazo (Schaack et al. 2010).

1.1.2 Classificação dos TEs

Quanto à sua classificação, TEs eucarióticos podem ser divididos em duas classes principais, definidas pelo tipo de intermediário de transposição gerado. Cada uma destas classes pode ser dividida em subclasses, definidas pelo mecanismo enzimático em que intermediários são gerados e inseridos, além de superfamílias e famílias, definidas pela relação filogenética dos seus membros (Bourque et al. 2018, Wells & Feschotte 2020). Elementos de classe I, também conhecidos como retrotransposons, utilizam intermediários de

RNA para se replicar. Estes intermediários são gerados por transcrição e posteriormente transcritos reversamente em DNA antes de serem integrados em um novo local do genoma hospedeiro, sendo que os elementos geradores dos intermediários permanecem intactos. Por esta razão, os elementos pertencentes à classe I também são referidos como sendo do tipo "copia-e-cola". Já elementos de classe II, também conhecidos como transposons de DNA, utilizam intermediários de DNA para se replicar. Como a grande maioria dos grupos de TEs nesta classe geram intermediários por meio da excisão do próprio elemento doador e reinserção em uma nova localidade do genoma hospedeiro, estes elementos também são referidos como sendo do tipo "corta-e-cola".

Entretanto, dentro da classe II, há duas subclasses de elementos que utilizam mecanismos de transposição distintos do padrão geral corta-e-cola, os Polintons (ou Mavericks) e os *Helitrons*. A primeira dessas subclasses compreende os Polintons que, apesar de não terem sido estudados em detalhe quanto ao seu mecanismo de transposição, provavelmente sintetizam intermediários de DNA diretamente a partir dos elementos doadores (Wells & Feschotte 2020). Mesmo que sejam considerados como TEs, o conjunto de evidências obtidas nos últimos anos indica de forma inequívoca que Polintons teriam se derivado de integrações virais em genomas hospedeiros e, por isso, provavelmente deveriam ser classificados como virus (Krupovic et al. 2014, Krupovic & Koonin 2015, Koonin & Krupovic 2017, Bellas & Sommaruga 2021).

1.2 Helitrons

A segunda subclasse de elementos da classe II que não utilizam um mecanismo de transposição do tipo corta-e-cola compreende os transposons conhecidos como *Helitrons*. Estes elementos eucarióticos foram identificados pela primeira vez em 2001 nos genomas de *Arabidopsis thaliana*, *Oriza sativa* e *Caenorhabditis elegans*, através de análises in silico (Kapitonov & Jurka 2001). Desde então, os *Helitrons* foram encontrados nos genomas de todos os principais grupos de organismos eucariotos em diferentes proporções. Por exemplo, *Helitrons* podem representar entre 0.1%-6.6% do DNA genômico em espécies de plantas e entre 0%-10% no caso de espécies animais (Kapitonov & Jurka 2007, Thomas & Pritham 2015). Estes transposons são encontrados em diferentes tamanhos que podem variar de poucas centenas de pb até poucos kb em elementos não-autônomos (que não codificam uma transposase funcional), e de poucos kb até várias dezenas de kb em elementos autônomos, dependendo do organismo hospedeiro e da família de *Helitron* em questão (e.g., Kapitonov & Jurka 2001, Pritham & Feschotte 2007, Du et al. 2009, Thomas et al. 2014, Chellapan et al. 2016).

Helitrons codificam uma transposase denominada RepHel, composta por dois domínios principais: uma endonuclease (Rep) e uma helicase (Hel) pertencente a superfamília 1 (S1H). O domínio Rep é o centro catalítico responsável pela clivagem do DNA nas extremidades do elemento doador e do sítio de inserção no cromossomo hospedeiro. Já o domínio Hel provavelmente é responsável por auxiliar na separação do DNA dupla fita (dsDNA) do elemento doador, gerando um intermediário de DNA fita simples (ssDNA). Além destes dois domínios comuns a todas transposases RepHel, *Helitrons* também possuem uma sequência palindrômica de 16-20 pb localizada ~ 11 pb antes da sua extremidade 3', capaz de formar estruturas secundárias do tipo hairpin ou stem-loop que provavelmente auxiliam no processo de transposição (Thomas & Pritham 2015) (Fig. 1).





Desde a descoberta dos *Helitrons*, notou-se a RepHel apresenta similaridades estruturais com transposases encontradas em elementos procarióticos (e.g., família IS91). Por isso, antes que estudos experimentais fossem conduzidos, todos os modelos sugeridos para descrever a transposição dos *Helitrons* se baseavam no mecanismo utilizado por TEs da família IS91 (Feschotte & Wessler 2001, Kapitonov & Jurka 2007, Thomas & Pritham 2015, Dias et al. 2016). Este mecanismo (Fig. 2), denominado transposição por círculo rolante (rolling-circle transposition, RCT) representa uma variação do processo conhecido como replicação por círculo rolante (rolling-circle replication, RCR), utilizado por diversos grupos de vírus e plasmídeos encontrados em organismos procariotos e eucariotos (Chandler et al. 2013, Wawrzyniak et al. 2017). Mais recentemente, análises experimentais confirmaram as principais etapas sugeridas para descrever a transposição dos *Helitrons*, além de revelar detalhes como, por exemplo, o fato de elementos circulares de dsDNA serem os intermediários viáveis de transposição (Grabundzija et al. 2016, 2018).





De acordo com o que sabemos atualmente sobre os processos RCR e RCT, inclusive em *Helitrons*, a transposição destes elementos se inicia com a ligação entre a primeira tirosina catalítica do domínio Rep e a extremidade 5' do elemento, criando um intermediário 5'fosfotirosina e uma extremidade 3'-OH livre no sítio doador. A fita líder ligada covalentemente ao domínio Rep começa a se desassociar da sua fita complementar, provavelmente com o auxílio da atividade de translocação sentido 5'-3' do domínio Hel. Ao mesmo tempo que a extremidade 5' da fita líder começa a ser desassociada, uma forquilha de replicação possivelmente se forma no mesmo local, promovendo a síntese das fitas complementares tanto do intermediário em desassociação, quanto do sítio doador a partir de sua 3'-OH terminal. Desta forma, um intermediário de dsDNA é sintetizado até que a RepHel alcança o lado oposto do elemento, clivando este com sua segunda tirosina catalítica e expondo uma extremidade 3'-OH livre que ataca a primeira ligação 5'-fosfotirosina, resultando na formação de um intermediário de dsDNA circular.

Em um segundo momento, a RepHel ligada covalentemente à extremidade 5' deste intermediário circular se associa à um segundo local do genoma hospedeiro que é clivado pela segunda tirosina presente na transposase, expondo uma extremidade 3'-OH livre do sítio receptor. Esta extremidade então ataca a ligação 5'-fosfotirosina entre a RepHel e o intermediário, ligando este ao DNA receptor. Após alcançar o lado oposto do intermediário circular, a primeira tirosina catalítica cliva este, gerando uma extremidade 3'-OH que ataca a ligação 5'-fosfotirosina entre o sítio receptor e a segunda tirosina catalítica. Tal processo resulta na inserção do *Helitron* na forma de um "loop" de DNA fita simples no sítio receptor que provavelmente é resolvido durante a replicação do genoma hospedeiro (Fig. 2).

1.2.1 Origem evolutiva dos Helitrons

Desde a descoberta dos Helitrons, foram propostas diferentes hipóteses para explicar sua origem e determinar quais seriam os elementos genéticos móveis mais próximos evolutivamente destes TEs eucarióticos. Por um lado, a semelhança estrutural e aparente semelhança funcional da RepHel com transposases de elementos procarióticos (e.g., família IS91) foi interpretada como um indício de que Helitrons seriam descendentes diretos ou parentes próximos destes últimos. Além disso, na época em que Helitrons foram descobertos em espécies de plantas e animais, vírus eucarióticos do tipo RCR haviam sido identificados apenas em espécies de plantas. Tal fato foi utilizado para sugerir a hipótese de que os Helitrons não só descenderiam de TEs procarióticos, mas talvez tivessem dado origem a vírus eucarióticos do tipo RCR (Kapitonov & Jurka 2001). Por outro lado, foi sugerida a hipótese alternativa de que os *Helitrons* poderiam ter se originado a partir de integrações ancestrais de vírus eucarióticos do tipo RCR (Feschotte & Wessler 2001). Esta hipótese foi baseada no fato de que, ao contrário dos transposons procarióticos do tipo RCT, os Helitrons codificam uma helicase e, em alguns casos, proteínas que se ligam a ssDNA (single-stranded binding proteins, SSBs), similarmente a vírus eucarióticos do tipo RCR. Além disso, integrações de vírus eucarióticos do tipo RCR já haviam sido identificadas em genomas de eucariotos, o que demonstraria a plausibilidade do cenário proposto.

Apesar de serem possíveis, ambas as hipóteses apresentam inconsistências ou requerem a ocorrência de eventos secundários para serem explicadas. No caso da primeira hipótese (origem a partir de transposons procarióticos), o principal problema se dá pelo fato de os *Helitrons* possuírem um domínio Rep seguido de uma helicase, ao contrário dos transposons procarióticos que só codificam um domínio Rep. Para explicar esta diferença foi sugerido que ancestrais dos *Helitrons* teriam adquirido seu domínio Hel por meio da captura de uma helicase proveniente de um hospedeiro eucarioto. As principais evidências que dão suporte à esta sugestão são a presença de introns no domínio Hel de alguns *Helitrons* e o fato de que o domínio Hel pertence à família de helicases Pif1 (Kapitonov & Jurka 2001, 2007, Thomas & Pritham 2015). Helicases da família Pif1 são encontradas em praticamente todos os eucariotos, sendo responsáveis por várias funções genômicas importantes como replicação e reparo do DNA, manutenção telomérica e mitocondrial, maturação de fragmentos de Okazaki, ruptura de complexos proteína-DNA, resolução de estruturas secundárias em ácidos nucleicos, dentre outras (Boule & Zakian 2006, Bochman et al. 2010, Muellner & Schmidt 2020)

A presença de uma helicase Pif1 na transposase RepHel também é inconsistente com a segunda hipótese (origem a partir de vírus eucarióticos). Apesar de *Helitrons* se assemelharem a vírus eucarióticos do tipo RCR por codificarem uma proteína com um domínio helicase, no caso dos *Helitrons* este domínio representa uma S1H, ao contrário dos vírus eucarióticos do tipo RCR em que sua helicase pertence a superfamília 3 (S3H) (Krupovic 2013, Koonin & Dolja 2014). Esta característica também é inconsistente com o cenário adicional proposto para a primeira hipótese (*Helitrons* teriam se originado de transposons procarióticos e deram origem a vírus eucarióticos). Em todo caso, até hoje nenhuma das hipóteses apresentadas acima foi investigada em detalhe, de forma que a origem dos *Helitrons*, e dos domínios presentes em sua transposase permanecem desconhecidos.

Nota-se que o conhecimento sobre os *Helitrons* tem avançado consideravelmente nas últimas duas décadas desde a sua descoberta, principalmente no que diz respeito à sua prevalência e influência nos genomas eucarióticos e, mais recentemente, ao seu mecanismo de transposição. Apesar disso, vemos que durante este mesmo período pouco, ou quase nada, foi revelado sobre a sua origem evolutiva e sua relação com outros elementos genéticos móveis.

2. OBJETIVOS

O objetivo geral do presente trabalho consistiu em investigar a origem evolutiva dos *Helitrons* utilizando análises filogenéticas moleculares das sequências de aminoácidos dos domínios presentes em sua transposase (RepHel).

Os objetivos específicos foram:

- (i) Investigar as relações evolutivas entre o domínio Rep presente nos *Helitrons* e proteínas codificadas por outros elementos genéticos móveis encontrados em procariotos e eucariotos.
- (ii) Testar as duas principais hipóteses acerca da origem dos *Helitrons*, sendo a primeira a de que estes teriam se originado de transposons procarióticos, e a alternativa a de que os *Helitrons* teriam se originado de vírus eucarióticos ou seriam parentes próximos destes.
- (iii) Investigar as relações evolutivas entre helicases presente nos *Helitrons* e as encontradas em diferentes organismos e elementos genéticos móveis, de forma a testar a hipótese de que os *Helitrons* teriam adquirido seu domínio Hel de um gene Pif1 eucariótico.
- (iv) Utilizar os dados obtidos nas análises anteriores para propor um cenário abrangente sobre a origem e evolução dos *Helitrons*.
- (v) Complementarmente, decidimos reexaminar a distribuição e a história evolutiva de uma família de *Helitrons* (Hel_c35) presente em artrópodes, identificada pelo nosso grupo em um trabalho anterior, utilizando para isso análises filogenéticas moleculares das suas sequências de nucleotídeos.

3. CAPÍTULO 1

Exploring the Remote Ties between *Helitron* Transposases and Other Rolling-Circle Replication Proteins

Pedro Heringer and Gustavo C. S. Kuhn

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, CEP 31270-901, Brazil.





Article Exploring the Remote Ties between Helitron Transposases and Other Rolling-Circle Replication Proteins

Pedro Heringer¹⁰ and Gustavo C. S. Kuhn *¹⁰

Departamento de Biologia Geral, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, CEP 31270-901, Brazil; pedrohlt@ufmg.br

* Correspondence: gcskuhn@ufmg.br; Tel.: +55-(31)-3409-3062

Received: 28 August 2018; Accepted: 7 October 2018; Published: 9 October 2018



Abstract: Rolling-circle replication (RCR) elements constitute a diverse group that includes viruses, plasmids, and transposons, present in hosts from all domains of life. Eukaryotic RCR transposons, also known as Helitrons, are found in species from all eukaryotic kingdoms, sometimes representing a large portion of their genomes. Despite the impact of Helitrons on their hosts, knowledge about their relationship with other RCR elements is still elusive. Here, we compared the endonuclease domain sequence of Helitron transposases with the corresponding region from RCR proteins found in a wide variety of mobile genetic elements. To do that, we used a stepwise alignment approach followed by phylogenetic and multidimensional scaling analyses. Although it has been suggested that Helitrons might have originated from prokaryotic transposons or eukaryotic viruses, our results indicate that Helitron transposases share more similarities with proteins from prokaryotic viruses and plasmids instead. We also provide evidence for the division of RCR endonucleases into three groups (Y1, Y2, and Yx), covering the whole diversity of this protein family. Together, these results point to prokaryotic elements as the likely closest ancestors of eukaryotic RCR transposons, and further demonstrate the fluidity that characterizes the boundaries separating viruses, plasmids, and transposons.

Keywords: Helitron; rolling-circle replication; mobile genetic element; viral evolution

1. Introduction

Rolling-circle replication (RCR) proteins are essential components of many genetic elements found in all three domains of life. These proteins can be classified into three different groups according to their main function: (i) Rep proteins (vegetative replication), (ii) Mob proteins/relaxases (conjugation), and (iii) transposases (transposon mobility) [1,2]. Helitrons are the eukaryotic representatives of RCR transposable elements (TEs), found in species from all eukaryotic kingdoms in highly variable copy numbers [3,4]. Their transposition is thought to occur by a mechanism similar to the one proposed for bacterial RCR TEs, like the IS91 family of elements [4–6]. Briefly, the Helitron transposase binds to the 5'-end of the element, using one of its two catalytic tyrosines to create a 5'-phosphotyrosine intermediate and a free 3'-OH at the donor site. The leading strand covalently bound to the transposase is displaced, the lagging strand is synthesized, and the second catalytic tyrosine nicks the 3'-end, promoting the formation of a double-strand circle intermediate. The transposase then cleaves the leading strand from the circular intermediate, but this time the second tyrosine cleaves the host's genome, forming a free 3'-OH which attacks the first 5'-phosphotyrosine linkage. After the 3'-end of the circular intermediate is also joined to the recipient's free 5'-end, an integrated single-strand "loop" is formed and probably resolved during the host's genome replication. In addition, it has been recently shown that Helitron transposition shares mechanistic similarities with the replication process used by some circular viruses [7]. Despite some of the differences in their mode of propagation, the main catalytic reaction used by all RCR elements is essentially the same [1].

Helitron transposases are composed of a typical domain, the endonuclease involved in the initiation of RCR (RCRE or Rep), fused to a helicase domain (Hel) from the superfamily 1 (S1H) (Figure 1) [4,8]. This protein, also known as RepHel, belongs to the HUH (named after one of its conserved motifs with two His residues separated by a hydrophobic residue) family of endonucleases [1]. Although HUH endonucleases from eukaryotic viruses and some plasmids also have a helicase domain, they belong to the superfamily 3 (S3H), which is unrelated to the one found in Helitrons. Furthermore, prokaryote viruses only encode a RCRE domain with no helicase (Figure 1) [8,9].



Figure 1. Modular diversity of HUH endonucleases. Schematic representation of the rolling-circle replication (RCR) proteins included in the present analysis. Rolling-circle replication endonuclease (RCRE) domains have the first two motifs (I and II), in addition to the third motif represented by one or two tyrosines (Y) in the catalytic core (dots represent variable amino acid residues). Domains are not drawn to scale, and segments after helicase domains are not represented. Based on information from Chandler et al. [1], Koonin and Dolja [8], and the Conserved Domain Database (CDD) search tool [10].

Since Helitrons were discovered [11], a few preliminary suggestions about their evolutive origins have been made. These can be generally divided in two scenarios: the first suggests that Helitrons originated from a prokaryotic ancestral RCR TE [8,11], and the second adds the possibility that Helitrons descended from an ancient eukaryotic viral integration [12]. The first scenario is mainly based on the obvious similarities in the mode of propagation of eukaryotic and prokaryotic RCR TEs, while the second scenario considers the fact that, in contrast to prokaryote RCR TEs, Helitron coding sequences include a helicase domain and sometimes a ssDNA-binding protein, similarly to some RCR proteins from eukaryotic viruses. The fact that many viral copies from geminiviruses were found to be integrated in the tobacco genome [13] was also used to support this hypothesis. In fact, since this scenario was first proposed, several studies showed copies from different eukaryotic RCR viruses in host chromosomes, revealing that viral integrations of these replicons are more common than it was previously thought (reviewed in [14]). In addition, it has been shown that several geminivirus-and parvovirus-related sequences integrated in eukaryote genomes display TE features, and have apparently shifted from a viral to a transposon-like mode of replication [15].

2 of 10

Despite the above considerations, some differences between the RCR proteins of Helitrons and eukaryotic viruses argue against their evolutionary relationship. Firstly, as mentioned before, helicases from these two classes of elements belong to different superfamilies. Also, with the exception of parvoviruses [16], all RCR proteins from eukaryotic ssDNA viruses contain only one tyrosine (Y1) in their catalytic core [9,17], in contrast to the RepHel from Helitrons, which has two (Y2) [4] (Figure 1). Although the number of catalytic tyrosines has been used to tentatively classify RCR proteins between two superfamilies [17], there is currently no phylogenetic support for this distinction. In view of these observations, and considering that domain rearrangements are not uncommon during protein evolution [18], the first scenario (i.e., that Helitrons originated from a prokaryotic ancestral RCR TE) seems to be more parsimonious, as the acquisition of a S1H domain would be the only major evolutionary step in a prokaryotic to eukaryotic RCR TE transition.

The relationship between Helitrons and other RCR genetic elements was initially assessed by Poulter et al. [19]. Although their results did not indicate a relationship between these TEs with specific RCR entities, they provided evidence for an ancient monophyletic origin of Helitrons, which probably occurred early on in the evolution of eukaryotes. However, the evolutionary origin of Helitrons has not been further examined, probably as a consequence of the low sequence identity of RepHel with any other group of RCR proteins [3].

In this study, we investigated the relationship of the Helitron RepHel with other RCR proteins by analyzing the RCRE amino acid sequences from a wide variety of mobile genetic elements, including TEs, plasmids, and viruses. Our results indicate that, despite being eukaryotic TEs, Helitron transposases display more sequence similarities with prokaryotic RCR proteins from bacteriophages and plasmids. In addition, we show that the HUH family of endonucleases can be divided into three major phylogenetic groups comprised of RCR proteins from highly heterogeneous mobile genetic elements.

2. Results and Discussion

2.1. Selecting and Preparing RCRE Domain Sequences

We selected a sample of 13 Helitron RepHel amino acid sequences, representing elements from distantly-related organisms across several phyla and including the main Helitron variants (Table S1). To analyze these TEs in a broad evolutionary context, at least three sequences of each family or group of RCR genetic elements from prokaryotes and eukaryotes were selected. These included single- and double-stranded viruses, plasmids, and TEs (Table S1).

Our analysis was restricted to the RCRE (or HUH) domain of the sequences (Figure 1), which has a central role in starting RCR reactions and is the only region common to all HUH endonucleases [1] (Figure 1). Modular rearrangements often occur during protein evolution [18] which is also the case for several RCR virus lineages [20]. For those reasons, and considering that flanking domains are highly variable amongst RCR elements [1], our restriction to the RCRE domain aimed to avoid spurious evolutionary inferences. Most proteins within the HUH family have three conserved motifs (I, II, and III) in the core region of the RCRE domain, despite the high sequence divergence between groups [1,2,21]. Only amino acid sequences containing all three conserved motifs in their typical arrangement (I-II-III) were selected for our analysis; this is because some HUH endonucleases display their motifs in the reverse order (e.g., III-II-I) [1,2], and these also have highly divergent amino acid sequences, which prevent reliable sequence alignments. A total of 115 amino acid sequences, representing the overall diversity of all known HUH endonucleases, were selected for the analysis (Table S1).

To reduce spurious alignments of the RCRE sequences, we conducted a stepwise alignment approach, which consisted of aligning each group of closely-related sequences separately, excluding segments flanking the RCRE domain and trimming the portions that were exclusive of individual

3 of 10

taxa. The resulting sequences (Data S1) were aligned using PSI-Coffee, which is a method considered suitable for highly divergent protein sequences with little or no structural information available [22,23].

2.2. Major RCR Protein Phylogenetic Groups

A phylogenetic analysis was conducted and pairwise divergence values between sequences were used to generate non-metric multidimensional scaling (NMDS) ordinations. As expected for an analysis that includes highly divergent sequences, clade support values between major groups were low, although we observed an overall agreement between our results and the known topology for most of the clades (Figure 2). Our results support the monophyletic nature of all Helitron variants and the lack of any clear relationship of these TEs with other specific groups or families of mobile genetic elements, as previously suggested [19]. Nonetheless, in both the phylogenetic analysis (Figure 2) and NMDS ordinations (Figure 3) we observed an overall distinction between Y1 and Y2 RCR proteins, which we henceforth refer to as Y1 and Y2 groups. An exception is a third clade, composed of elements from both variants, which we refer to here as the Yx group because the number of tyrosines of the catalytic core of its members does not relate with the canonical Y1 and Y2 division. Although the resulting phylogeny revealed a basal segregation of Yx RCR proteins and the rest of the sequences, the Y2 group appears to be more closely related to Y1 RCR proteins, and perhaps constitutes a derivative clade of the Y1 group (Figure 2 and 3).



Figure 2. Phylogenetic analysis of RCRE domain sequences. Clade colors indicate each tyrosine group: Y1 (green), Y2 (red), and Yx (blue). Taxa colors represent the family of each element (box on the upper right). See Table S1 for taxa information. Phylogeny inferred by the Maximum Likelihood method (LG+G+I). The same phylogeny, with the numerical support values represented, is shown on Figure S1.

The topology observed within the Yx group is roughly in agreement with previous results [24], indicating that this clade represents a bona fide phylogenetic cluster composed of archaeal viruses and bacterial TEs. Recent analyses using different methods have also shown that parvoviruses belong to a separate clade from other eukaryotic RCR viruses [25]. However, we did not expect that parvoviral RCR proteins (AAV2, AAV5, and SLP) would group together with Yx elements (Figures 2 and 3). Although structural similarities indicate a distant relationship between parvoviral and other RCR proteins [26], the positioning of these viruses in the Yx group might also be the consequence of long branch attraction [27], so this result should be treated with caution.



Figure 3. Non-metric multidimensional scaling (NMDS) of evolutionary divergence between RCRE domains. (**A**) Ordinations with taxa represented by their sequence abbreviations. Colors indicate the different classes of mobile genetic elements. (**B**) Same ordinations of (**A**), with colors indicating the tyrosine group of each taxa. The scaling represents euclidean distances for two dimensions (stress: 0.26382).

As revealed by the results from both analyses, the assignment to a specific catalytic tyrosine group is not contingent on the element class (Figures 2 and 3). For instance, bacterial plasmids, and eukaryotic and archaeal viruses have members in more than one group. Likewise, the element class does not always predict its topology, even within the same tyrosine group. For example, some Y1 viral families are closer to Y1 plasmids than other Y1 viruses, and the same is true in the Y2 group. This phenomenon has been observed in different studies and emphasizes the marked fluidity at the boundaries separating different classes of mobile genetic elements (reviewed in [8,9]). Thus, our results indicate that the tyrosine group division is the only informative phylogenetic feature encompassing the whole HUH endunuclease family.

2.3. Helitron Transposase is More Similar to Prokaryotic Proteins

Even though the Helitron RepHel does not appear to be phylogenetically closer to any single family of proteins, they clustered within the Y2 group which, apart from Helitrons, is exclusively composed of prokaryotic viruses and plasmids (Figures 2 and 3). On the other hand, sequences from prokaryotic TEs clustered within the Yx group, even though some of them (including the IS91 family) have two tyrosines in their catalytic core and share a similar transposition mechanism with Helitrons [4–6,28]. It is also notable that RepHel proteins appear to be only distantly related to RCR proteins from eukaryotic viruses, which almost exclusively belong to the Y1 group. These observations indicate that the core domain from Helitron transposases is more similar to proteins from prokaryotic viruses and plasmids than to prokaryotic RCR transposases or to eukaryotic viral proteins.

As we have mentioned, in addition to the RCRE domain, RepHel proteins also have a S1 helicase domain (Figure 1); more specifically, this S1 helicase belongs to the Pif1 family [4]. Although

5 of 10

Pif1 helicases are present in essentially all eukaryote genomes, they also have been found in some prokaryotes [29,30]. Because all known prokaryotic Y2 RCR proteins lack a helicase, this domain could have been acquired from a prokaryote host by the Helitron ancestor before it colonized the first eukaryote genome. However, considering that Pif1 helicases are ubiquitous in eukaryote genomes and found less frequently in prokaryotes, it seems more plausible that Helitrons acquired their helicase domain from a eukaryotic host. Indeed, a preliminary analysis of Pif1 sequences from Helitrons, eukaryotes, and prokaryotes indicates that the helicase domain from distinct Helitron variants formed separate clusters with different fungal proteins, suggesting that Helitrons acquired their helicase domain from at least two independent events (Figure S2).

These results support the hypothesis of an ancient origin of Helitrons during the initial radiation of eukaryotes, and suggest that neither prokaryotic TEs, nor eukaryotic viruses, are among their closest relatives. Instead, we provide evidence for a closer relationship of these eukaryotic TEs with prokaryotic viruses and plasmids with Y2 RCR proteins, even though it is not possible to determine which specific family shares the most recent common ancestor with the RepHel (Figure 4). Thus, our proposition is that Helitrons descend from a prokaryotic Y2 mobile element that integrated in the genome of an early eukaryote ancestor. Like all other known prokaryotic Y2 elements, the Helitron progenitor probably coded an RCR protein devoid of a helicase domain and was dependent of its host for correct replication/transposition. Subsequently, each of the incipient Helitron variants acquired a eukaryotic helicase by the recombination of its RCRE domain with a host helicase gene. In any case, a comprehensive understanding of the Helitron origins will probably rely on the future discovery of new groups of RCR genetic elements.



Figure 4. Proposed scenario for the origin of Helitrons and other RCR elements. Arrows represent putative pathways to explain the observed relationship among RCR elements. Virion images were obtained from VIPERdb (http://viperdb.scripps.edu) [31].

6 of 10

Finally, although the RCRE phylogeny does not coincide with the taxonomic division of distinct genetic elements classes (viruses, plasmids and TEs), we suggest that the HUH family of endonucleases is composed by three major radiation groups (Y1, Y2 and Yx). Interestingly, most of the HUH endonucleases can be assigned to one of these groups simply by having a tyrosine residue at a specific position in the RCRE domain, regardless of the element's class. The extreme diversity observed in each of these groups underscore the dynamic nature of mobile genetic elements which, in the long term, do not evolve under the usual taxonomic constraints acting upon their hosts.

3. Materials and Methods

3.1. Sequences Retrieval and Selection

RepHel amino acid sequences from Helitrons were retrieved from Repbase (https://www.girinst. org/repbase/) [32] and GenBank (https://www.ncbi.nlm.nih.gov/genbank/) [33], using elements from previous studies as a reference (e.g., [11,19,34]). The structure of these proteins was verified using the Conserved Domain Database (CDD) search tool (https://www.ncbi.nlm.nih.gov/Structure/cdd/ wrpsb.cgi) [10]. RepHel sequences that could be clearly assigned to one of the three main Helitron variants [4] were selected: canonical Helitron (6 sequences), Helitron2 (1 sequence), and Helentron (6 sequences). Sequences representing each family or group of RCR proteins were retrieved on GenBank [33], based on several references (e.g., [9,21,24,35–37]). A total of 115 amino acid sequences were selected for the alignment (Table S1).

3.2. Sequence Alignment

Each family or group of sequences were aligned separately using the M-Coffee mode from T-Coffee (http://tcoffee.crg.cat/) [22] before being manually trimmed in order to exclude flanking portions of the RCRE domain and the segments that are exclusive of individual taxa. The trimmed sequences (Data S1) were aligned with PSI-Coffee (http://tcoffee.crg.cat/apps/tcoffee/do:psicoffee) [22] before manual correction. Alignment positions with less than 90% coverage were excluded.

3.3. NMDS and Phylogenetic Analysis

Pairwise evolutionary divergence between sequences was estimated using the Poisson correction model on MEGA7 [38]. The values were used to generate non-metric multidimensional scaling (NMDS) ordinations with the R package vegan [39], representing euclidean distances for two dimensions. NMDS and plotting of ordinations were conducted in RStudio v1.1.442 (Boston, MA, USA) [40]. The best-fit evolutionary model for the alignment (LG+G+I) was determined using MEGA7 [38] and the Smart model selection (SMS) in PhyML (http://www.atgc-montpellier.fr/phyml/) [41]. Maximum Likelihood phylogeny was inferred from 5000 replicates using MEGA7 [38], and the final phylogenetic tree edited using iTOL v4.2.3 (https://itol.embl.de/) [42].

Supplementary Materials: Supplementary materials can be found at http://www.mdpi.com/1422-0067/19/10/3079/s1.

Author Contributions: Conceptualization, P.H. and G.C.S.K.; Methodology, P.H.; Investigation, P.H.; Resources, G.C.S.K.; Data Curation, P.H.; Writing-Original Draft Preparation, P.H.; Writing-Review & Editing, G.C.S.K.; Visualization, P.H.; Funding Acquisition, G.C.S.K.

Funding: This research was funded by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) grant number 404620/2016-7 to G.C.S.K and doctoral fellowship from Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) to P.H.

Acknowledgments: We thank Renan P. de Souza for the insightful discussions that helped to enrich our analysis and "Pró-Reitoria de Pesquisa da Universidade Federal de Minas Gerais" for partially covering the publication costs.

Conflicts of Interest: The authors declare no conflict of interest.

Int. J. Mol. Sci. 2018, 19, 3079

Abbreviations

RCR	Rolling-circle replication
TE	Transposable element
RCRE	Rolling-circle replication endonuclease domain
S1H	Superfamily 1 helicase
S3H	Superfamily 3 helicase
RepHel	Helitron transposase (Rep/Helicase)
ssDNA	Single-strand DNA
NMDS	Non-metric multidimensional scaling

References

- Chandler, M.; De La Cruz, F.; Dyda, F.; Hickman, A.B.; Moncalian, G.; Ton-Hoang, B. Breaking and joining single-stranded DNA: The HUH endonuclease superfamily. *Nat. Rev. Microbiol.* 2013, *11*, 525–538. [CrossRef] [PubMed]
- Wawrzyniak, P.; Płucienniczak, G.; Bartosik, D. The Different Faces of Rolling-Circle Replication and Its Multifunctional Initiator Proteins. *Front. Microbiol.* 2017, *8*, 2353. [CrossRef] [PubMed]
- Kapitonov, V.V.; Jurka, J. Helitrons on a roll: Eukaryotic rolling-circle transposons. *Trends Genet.* 2007, 23, 521–529. [CrossRef] [PubMed]
- 4. Thomas, J.; Pritham, E.J. *Helitrons, the Eukaryotic Rolling-Circle Transposable Elements in Mobile DNA III*, 3rd ed.; ASM Press: Washington, DC, USA, 2015; pp. 893–926.
- Dias, G.B.; Heringer, P.; Kuhn, G.C. Helitrons in *Drosophila*: Chromatin modulation and tandem insertions. *Mob. Genet. Elements* 2016, 6, e1154638. [CrossRef] [PubMed]
- Grabundzija, I.; Messing, S.A.; Thomas, J.; Cosby, R.L.; Bilic, I.; Miskey, C.; Jurka, J. A Helitron transposon reconstructed from bats reveals a novel mechanism of genome shuffling in eukaryotes. *Nat. Commun.* 2016, 7, 10716. [CrossRef] [PubMed]
- Grabundzija, I.; Hickman, A.B.; Dyda, F. Helraiser intermediates provide insight into the mechanism of eukaryotic replicative transposition. *Nat. Commun.* 2018, *9*, 1278. [CrossRef] [PubMed]
- Koonin, E.V.; Dolja, V.V. Virus world as an evolutionary network of viruses and capsidless selfish elements. Microbiol. Mol. Biol. Rev. 2014, 78, 278–303. [CrossRef] [PubMed]
- Krupovic, M. Networks of evolutionary interactions underlying the polyphyletic origin of ssDNA viruses. *Curr. Opin. Virol.* 2013, *3*, 578–586. [CrossRef] [PubMed]
- Marchler-Bauer, A.; Bo, Y.; Han, L.; He, J.; Lanczycki, C.J.; Lu, S.; Chitsaz, F.; Derbyshire, M.K.; Geer, R.C.; Gonzales, N.R.; et al. CDD/SPARCLE: Functional classification of proteins via subfamily domain architectures. *Nucleic Acids Res.* 2017, 45, D200–D203. [CrossRef] [PubMed]
- Kapitonov, V.V.; Jurka, J. Rolling-circle transposons in eukaryotes. Proc. Natl. Acad. Sci. USA 2001, 98, 8714–8719. [CrossRef] [PubMed]
- Feschotte, C.; Wessler, S.R. Treasures in the attic: Rolling circle transposons discovered in eukaryotic genomes. Proc. Natl. Acad. Sci. USA 2001, 98, 8923–8924. [CrossRef] [PubMed]
- Bejarano, E.R.; Khashoggi, A.; Witty, M.; Lichtenstein, C. Integration of multiple repeats of geminiviral DNA into the nuclear genome of tobacco during evolution. *Proc. Natl. Acad. Sci. USA* 1996, 93, 759–764. [CrossRef] [PubMed]
- 14. Krupovic, M.; Forterre, P. Single-stranded DNA viruses employ a variety of mechanisms for integration into host genomes. *Ann. N. Y. Acad. Sci.* 2015, *1341*, 41–53. [CrossRef] [PubMed]
- Liu, H.; Fu, Y.; Li, B.; Yu, X.; Xie, J.; Cheng, J.; Ghabrial, S.A.; Li, G.; Yi, X.; Jiang, D. Widespread horizontal gene transfer from circular single-stranded DNA viruses to eukaryotic genomes. *BMC Evol. Biol.* 2011, 11, 276. [CrossRef] [PubMed]
- Hickman, A.B.; Ronning, D.R.; Kotin, R.M.; Dyda, F. Structural unity among viral origin binding proteins: Crystal structure of the nuclease domain of adeno-associated virus Rep. *Mol. Cell* 2002, 10, 327–337. [CrossRef]
- 17. Rosario, K.; Duffy, S.; Breitbart, M. A field guide to eukaryotic circular single-stranded DNA viruses: Insights gained from metagenomics. *Arch. Virol.* **2012**, *157*, 1851–1871. [CrossRef] [PubMed]

8 of 10

- Björklund, Å.K.; Ekman, D.; Light, S.; Frey-Skött, J.; Elofsson, A. Domain rearrangements in protein evolution. J. Mol. Biol. 2005, 353, 911–923. [CrossRef] [PubMed]
- Poulter, R.T.; Goodwin, T.J.; Butler, M. Vertebrate helentrons and other novel Helitrons. *Gene* 2003, 313, 201–212. [CrossRef]
- Kazlauskas, D.; Varsani, A.; Krupovic, M. Pervasive Chimerism in the Replication-Associated Proteins of Uncultured Single-Stranded DNA Viruses. *Viruses* 2018, 10, 187. [CrossRef] [PubMed]
- Ilyina, T.V.; Koonin, E.V. Conserved sequence motifs in the initiator proteins for rolling circle DNA replication encoded by diverse replicons from eubacteria, eucaryotes and archaebacteria. *Nucleic Acids Res.* 1992, 20, 3279–3285. [CrossRef] [PubMed]
- 22. Di Tommaso, P.; Moretti, S.; Xenarios, I.; Orobitg, M.; Montanyola, A.; Chang, J.M.; Notredame, C. T-Coffee: A web server for the multiple sequence alignment of protein and RNA sequences using structural information and homology extension. *Nucleic Acids Res.* **2011**, *39*, W13–W17. [CrossRef] [PubMed]
- Taly, J.F.; Magis, C.; Bussotti, G.; Chang, J.M.; Di Tommaso, P.; Erb, I.; Espinosa-Carrasco, J.; Kemena, C.; Notredame, C. Using the T-Coffee package to build multiple sequence alignments of protein, RNA, DNA sequences and 3D structures. *Nat. Protoc.* 2011, *6*, 1669–1682. [CrossRef] [PubMed]
- Wang, Y.; Chen, B.; Cao, M.; Sima, L.; Prangishvili, D.; Chen, X.; Krupovic, M. Rolling-circle replication initiation protein of haloarchaeal sphaerolipovirus SNJ1 is homologous to bacterial transposases of the IS91 family insertion sequences. J. Gen. Virol. 2018, 99, 416–421. [CrossRef] [PubMed]
- 25. Aiewsakun, P.; Simmonds, P. The genomic underpinnings of eukaryotic virus taxonomy: Creating a sequence-based framework for family-level virus classification. *Microbiome* **2018**, *6*, 38. [CrossRef] [PubMed]
- Campos-Olivas, R.; Louis, J.M.; Clérot, D.; Gronenborn, B.; Gronenborn, A.M. The structure of a replication initiator unites diverse aspects of nucleic acid metabolism. *Proc. Natl. Acad. Sci. USA* 2002, 99, 10310–10315. [CrossRef] [PubMed]
- 27. Bergsten, J. A review of long-branch attraction. *Cladistics* 2005, 21, 163–193. [CrossRef]
- Garcillan-Barcia, M.P.; Bernales, I.; Mendiola, M.V.; de la Cruz, F. Single-stranded DNA intermediates in IS91 rolling-circle transposition. *Mol. Microbiol.* 2001, *39*, 494–501. [CrossRef]
- Bochman, M.L.; Sabouri, N.; Zakian, V.A. Unwinding the functions of the Pif1 family helicases. DNA Repair 2010, 9, 237–249. [CrossRef] [PubMed]
- Bochman, M.L.; Judge, C.P.; Zakian, V.A. The Pif1 family in prokaryotes: What are our helicases doing in your bacteria? *Mol. Biol. Cell* 2011, 22, 1955–1959. [CrossRef] [PubMed]
- Carrillo-Tripp, M.; Shepherd, C.M.; Borelli, I.A.; Venkataraman, S.; Lander, G.; Natarajan, P.; Johnson, J.E.; Brooks, C.L.; Reddy, V.S. VIPERdb2: An enhanced and web API enabled relational database for structural virology. *Nucleic Acids Res.* 2009, 37, D436–D442. [CrossRef] [PubMed]
- Bao, W.; Kojima, K.K.; Kohany, O. Repbase Update, a database of repetitive elements in eukaryotic genomes. Mob. DNA 2015, 6, 11. [CrossRef] [PubMed]
- Benson, D.A.; Cavanaugh, M.; Clark, K.; Karsch-Mizrachi, I.; Lipman, D.J.; Ostell, J.; Sayers, E.W. GenBank. Nucleic Acids Res. 2017, 45, D37–D42. [CrossRef] [PubMed]
- Pritham, E.J.; Feschotte, C. Massive amplification of rolling-circle transposons in the lineage of the bat Myotis lucifugus. *Proc. Natl. Acad. Sci. USA* 2007, 104, 1895–1900. [CrossRef] [PubMed]
- Zawar-Reza, P.; Argüello-Astorga, G.R.; Kraberger, S.; Julian, L.; Stainton, D.; Broady, P.A.; Varsani, A. Diverse small circular single-stranded DNA viruses identified in a freshwater pond on the McMurdo Ice Shelf (Antarctica). *Infect. Genet. Evol.* 2014, 26, 132–138. [CrossRef] [PubMed]
- Kazlauskas, D.; Dayaram, A.; Kraberger, S.; Goldstien, S.; Varsani, A.; Krupovic, M. Evolutionary history of ssDNA bacilladnaviruses features horizontal acquisition of the capsid gene from ssRNA nodaviruses. *Virology* 2017, 504, 114–121. [CrossRef] [PubMed]
- Wang, Y.; Sima, L.; Lv, J.; Huang, S.; Liu, Y.; Wang, J.; Krupovic, M.; Chen, X. Identification, characterization, and application of the replicon region of the halophilic temperate sphaerolipovirus SNJ1. *J. Bacteriol.* 2016, 198, 1952–1964. [CrossRef] [PubMed]
- Kumar, S.; Stecher, G.; Tamura, K. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol. Biol. Evol.* 2016, 33, 1870–1874. [CrossRef] [PubMed]
- 39. Dixon, P. VEGAN, a package of R functions for community ecology. J. Veg. Sci. 2003, 14, 927–930. [CrossRef]
- RStudio Team. RStudio: Integrated Development for R; RStudio, Inc.: Boston, MA, USA, 2016; Available online: http://www.rstudio.com/ (accessed on 8 October 2018).

Int. J. Mol. Sci. 2018, 19, 3079

- 41. Lefort, V.; Longueville, J.E.; Gascuel, O. SMS: Smart model selection in PhyML. *Mol. Biol. Evol.* **2017**, *34*, 2422–2424. [CrossRef] [PubMed]
- 42. Letunic, I.; Bork, P. Interactive tree of life (iTOL) v3: An online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Res.* **2016**, *44*, W242–W245. [CrossRef] [PubMed]



@ 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).

10 of 10

4. CAPÍTULO 2

Pif1 Helicases and the Evidence for a Prokaryotic Origin of Helitrons

Pedro Heringer and Gustavo C. S. Kuhn

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, CEP 31270-901, Brazil.

MINE Cest on 27 January 2022

Pif1 Helicases and the Evidence for a Prokaryotic Origin of *Helitrons*

Pedro Heringer and Gustavo C.S. Kuhn*

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil

*Corresponding author: E-mail: gcskuhn@ufmg.br. Associate editor: Irina Arkhipova

Abstract

Helitrons are the only group of rolling-circle transposons that encode a transposase with a helicase domain (Hel), which belongs to the Pif1 family. Because Pif1 helicases are important components of eukaryotic genomes, it has been suggested that Hel domains probably originated after a host eukaryotic Pif1 gene was captured by a *Helitron* ancestor. However, the few analyses exploring the evolution of *Helitron* transposases (RepHel) have focused on its Rep domain, which is also present in other mobile genetic elements. Here, we used phylogenetic and nonmetric multidimensional scaling analyses to investigate the relationship between Hel domains and Pif1-like helicases from a variety of organisms. Our results reveal that Hel domains are only distantly related to genomic helicases from eukaryotes and prokaryotes, and thus are unlikely to have originated from a captured Pif1 gene. Based on this evidence, and on recent studies indicating that Rep domains are more closely related to rolling-circle plasmids and phages, we suggest that *Helitrons* are descendants of a RepHelencoding prokaryotic plasmid element that invaded eukaryotic genomes before the radiation of its major groups. We discuss how a Pif1-like helicase domain might have favored the transposition of *Helitrons* in eukaryotes beyond simply unwinding DNA intermediates. Finally, we demonstrate that some examples in the literature describing genomic helicases from eukaryotes actually consist of Hel domains from *Helitrons*, a finding that underscores how transposons can hamper the analysis of eukaryotic genes. This investigation also revealed that two groups of land plants appear to have lost genomic Pif1 helicases independently.

Key words: Helitrons, transposon, Pif1, helicase.

Introduction

Helitrons are DNA transposable elements (TEs) found in a wide variety of species from all eukaryotic kingdoms but make up variable genomic proportions across different taxa. For instance, they constitute between 0.1% and 6.6% of the genomic DNA in plants and between 0% and 10% in animals (reviewed in Kapitonov and Jurka [2007] and Thomas and Pritham [2015]). These TEs have been shown to mobilize within a genome by a process known as rolling-circle (RC) transposition (RCT) (Grabundzija et al. 2016, 2018) which could be viewed as a variation of the RC replication (RCR) process employed by several groups of plasmids and viruses from prokaryotes and eukaryotes (reviewed in Chandler et al. [2013] and Wawrzyniak et al. [2017]). In Helitrons, the RCT is executed by the Rep/Helicase (RepHel) transposase, which is composed by two major domains: an endonuclease (Rep) domain and a superfamily 1 helicase (Hel) domain (Thomas and Pritham 2015) (fig. 1).

Helitrons can be classified into four structural and coding variants, namely Helitron, Helentron, Helitron2, and Proto-Helentron (Thomas and Pritham 2015). In contrast to the first three variants, which have been shown to represent distinct

phylogenetic groups (Poulter et al. 2003; Thomas et al. 2014; Heringer and Kuhn 2018), Proto-Helentron elements seem to constitute a subtype of Helentrons with derived Helitron-like structural features (Thomas et al. 2014). Although all Helitrons have RepHel proteins with two major domains, distinct variants, or specific variant lineages, can encode additional domains in their transposase or/and additional genes. Likewise, specific sets of structural features, like inverted repeats, can be used to identify major lineages or variants (fig. 1).

The Hel domain present in *Helitron* transposases is a superfamily 1 helicase, more specifically from the Pif1 family (Kapitonov and Jurka 2001; Thomas and Pritham 2015). Pif1 helicases have been found in essentially all eukaryotes studied to date (Bochman et al. 2010) and are involved in several processes, like DNA replication and repair, telomere maintenance, Okazaki fragment maturation, disruption of protein–DNA complexes, resolution of nucleic acid secondary structures, mitochondrial DNA maintenance, among others (reviewed in Boule and Zakian [2006]; Bochman et al. [2010]; and Muellner and Schmidt [2020]). Although typically known as eukaryotic proteins, Pif1-like helicases can

© The Author(s) 2021. Published by Oxford University Press on behalf of the Society for Molecular Biology and Evolution. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https:// creativecommons.org/licenses/by-nc/40/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com



Mol. Biol. Evol. 39(1):msab334 doi:10.1093/molbev/msab334 Advance Access publication November 25, 2021

also be found in some prokaryotic species, bacteriophages, and eukaryotic viruses (Bochman et al. 2011). We henceforth refer to eukaryotic and prokaryotic proteins that perform genomic-related tasks as genomic Pif1 helicases, in order to distinguish them from Pif1-like viral helicases or Hel domains found in *Helitron* transposases.

The structural and mechanistic similarities between eukaryotic and prokaryotic RC transposons initially prompted the hypothesis that Helitrons could be descendants of bacterial elements (e.g., IS91 family). Furthermore, it was suggested that Helitron ancestors could have given rise to eukaryotic RCR viruses, as these viruses were only found in plant species at that time (Kapitonov and Jurka 2001). Conversely, because geminiviruses had been found integrated into plant chromosomes, it was also proposed that Helitrons could likewise be derived from an ancient genomic integration of a eukaryotic RCR virus (Feschotte and Wessler 2001). However, as revealed by recent findings, Rep domains from Helitrons are distantly related to proteins from prokaryotic TEs and eukaryotic viruses, and share more similarities with RCR plasmids and viruses from bacteria (Heringer and Kuhn 2018; Kazlauskas et al. 2019). In spite of these similarities, the prokaryotic plasmid and viral elements which are more closely related to Helitrons do not encode a helicase domain (Heringer and Kuhn 2018), what makes the origin of Hel domains a still unsolved issue. The absence of helicases on the coding sequences of prokaryotic RC TEs, together with the presence of introns in some Hel domains from plants and Caenorhabditis elegans Helitrons, have been considered as tentative evidences that a Helitron ancestor acquired its Hel domain by capturing a helicase gene from its eukaryotic host (Kapitonov and Jurka 2001, 2007; Thomas and Pritham 2015). However, we still lack information about the evolutionary origins of Helitron Hel domains and their relationship with other helicases, as these issues have never been investigated in detail.

The fact that Pif1 family helicases are present in virtually all eukaryotes but absent in RC mobile genetic elements (MGEs), except Helitrons, renders the investigation about the origin of Hel domains more difficult. Moreover, to our knowledge there are no automated methods to clearly distinguish genomic Pif1 helicases from Helitron Pif1-like helicases. Regarding the later issue, both genomic and Helitron Pif1-like sequences can be found in eukaryotic genomes and sometimes is not possible to discriminate them without a more detailed analysis. For instance, Blastp searches on eukaryotic genomes using Pif1 proteins as queries often result in multiple significant hits, even though most eukaryotic species apparently have only one or two genomic Pif1 helicases (Bochman et al. 2010). Therefore, although up to few hits are expected to represent genomic Pif1 helicases in eukaryotic species, most of them often constitute Helitron Pif1-like protein sequences. In addition, some eukaryotes apparently have multiple genomic Pif1 paralogs (Bochman et al. 2010, 2011; Harman and Manna 2016), which makes their distinction from Helitron Pif1-like helicases even more complex.

In the present study, we retrieved prokaryotic, eukaryotic and viral Pif1-like proteins in silico using a stepwise searching method to avoid classifying *Helitron* coding sequences as genomic helicases. After doing so, we were able to investigate the relationship between Hel domains and Pif1-like genes from a wide variety of organisms and MGEs. Our results reveal further valuable information about the evolution of RepHel transposases, indicating that Hel domains are only distantly related to genomic Pif1 helicases and were likely present in *Helitrons* before they invaded eukaryotic hosts. We discuss the general implications of our findings considering the known mechanistic features of RepHel transposases and Pif1 helicases, also demonstrating how the similarities between these proteins can interfere with their classification and analysis.

Results

Finding Genomic Helicases

Before conducting searches to retrieve genomic Pif1-like helicases, we first expanded our sample of Helitrons from different variants (Helitrons, Helentrons, and Helitron2) selected previously (Heringer and Kuhn 2018). Consensus sequences from the helicase domains (Hel) found in those Helitrons were used as queries to obtain Pif1-like helicases from a wide diversity of organisms (see Materials and Methods). Because Helitrons are found throughout a large portion of eukaryotic genomes, the distinction between genomic Pif1 and Helitron Pif1-like helicases (Hel domains) across individual species is highly prone to identification errors (supplementary fig. S1, Supplementary Material online). For that reason, we initially selected only organisms lacking Helitron Rep sequences in their genomes, so that genomic Pif1 helicases could be correctly identified before our analyses. Helitron Rep sequences can be used as unique identifiers for the presence of Helitrons as they are exclusive of these RC elements and do not have genomic counterparts in eukaryotes.

The larger or smaller representation of specific taxonomic groups in the Pif1 helicases selected initially, depended on the number of available genomes and on the presence or absence of *Helitrons* in each taxon. For instance, although our searches on Embryophyta (land plants) revealed the presence of Pif1like proteins in most species, only the common liverwort *Marchantia polymorpha* was devoid of Rep sequences from *Helitrons*, thus being the single representative of land plants selected in the first round of searches.

Although almost all retrieved sequences from prokaryotes and eukaryotes were annotated as genomic Pif1 helicases, one of the hits from the searches on archaea was a TraA relaxase annotated as belonging to a species from the *Methanothrix* genus (*Methanothrix* sp., accession number: TFH49976.1). This hit displays a relatively low sequence coverage (62%) and identity (24%) to the query (*Helentron* Hel consensus) (supplementary data S1, Supplementary Material online). Nevertheless, as TraA relaxases constitute a group of proteins involved in conjugation of bacterial plasmids and are also known to have a helicase domain (Alt-Mörbe et al. 1996; Pérez-Mendoza et al. 2006), we decided to include additional TraA relaxase representatives in our analysis. To do that, the *Methanothrix* TraA relaxase (TFH49976.1) was used as query 33

MBE



Fig. 1. Helitron structural and coding variants. Each variant can be identified by a set of structural (symbols) and coding sequences (colored boxes). Helitrons, Helitron2, and Helentrons are major phylogenetic variants, with Proto-Helentrons representing an internal group of Helentrons that have intermediate features found in Helitrons and Helentrons. Adapted from Thomas and Pritham (2015).

in Blastp searches on the nonredundant protein sequences (nr) database from GenBank (Sayers et al. 2019). Interestingly, the best hits from this search consisted of TraA sequences from the phylum Proteobacteria (supplementary table S1, Supplementary Material online), with no hits from archaeal species, indicating that TFH49976.1 could either represent a horizontally transferred gene (from a bacterium to an archaeon) or a misannotated sequence from a bacterium species (discussed in the next topic).

Using our stepwise search and selection method (schematic workflow depicted in fig. 2), we retrieved an initial sample of 76 putative genomic Pif1 helicases from a wide variety of eukaryotes, prokaryotes, and plasmids, all lacking Helitron sequences in their genomes. After retrieving this sample of genomic (and plasmid) helicases, we further expanded the number of proteins in our data set by selecting Pif1-like helicases in all major groups of eukaryotes, prokaryotes and viruses, without filtering taxa by the presence of Helitron sequences. In addition to Hel domain consensus sequences, this time we also used the Saccharomyces cerevisiae Pif1 (NP_013650.1) as a query in Blastp searches. The proteins identified and selected previously with the Rep-filtering procedure were used to aid in the classification of this new set of Pif1-like proteins as genomic helicases or Hel domains from Helitrons by their relationship revealed in the phylogenetic analysis. We also included eukaryotic and prokaryotic viruses in this step of Blastp searches. All taxa selected for further analyses are shown in supplementary table S1. Supplementary Material online.

Phylogenetic Analysis

We used our final sample of 310 aligned protein sequences from *Helitrons*, eukaryotic and prokaryotic organisms, plasmids and viruses, to infer their phylogenetic relationship using the Maximum Likelihood method. Our resulting phylogeny revealed seven well supported major clades (or groups), named as follows: 1) TraA, 2) Myoviridae, 3) nucleocytoplasmic large DNA viruses (NCLDV)/Baculoviridae, 4) Helentron/ Helitron2, 5) Helitron, 6) Prokaryotic, and 7) Eukaryotic clade (fig. 3). The TraA clade included exclusively TraA relaxases and constitute a sister group of the Myoviridae clade, which is composed by helicases from a subset myoviruses. The NCLDV/Baculoviridae group included helicases from a subset of NCLDV and all retrieved baculoviruses. Together with the Helentron/Helitron2 and Helitron clades, they represent a basal group relative to the Prokaryotic and Eukaryotic major clades, as shown in the rooted tree (supplementary fig. S2, Supplementary Material online). The Prokaryotic clade includes most bacterial, archaeal and bacteriophage sequences. In contrast, the Eukaryotic major clade, which formed a sister group with the Prokaryotic clade, included all eukaryotic sequences, plus some bacterial, archaeal, eukaryotic viruses, and bacteriophage sequences, being the most diverse group in the phylogeny.

Regarding the distribution of *Helitron* variants, we observed two distinct and well supported clades, one with *Helitron* and the other containing *Helentron* plus *Helitron2* sequences (fig. 3). However, the connection between these two clades, and between each one of them and other groups of helicases, have low branch support values, and thus are presented collapsed in the phylogeny (fig. 3; supplementary fig. S2, Supplementary Material online). Considering previous analyses involving the Rep domain (Poulter et al. 2003; Heringer and Kuhn 2018) and the fact that a monophyletic origin of all *Helitrons* seems more parsimonious, the observed paraphyletic distribution of two major *Helitron* groups in our phylogeny could represent a methodological artifact (see Discussion). Nevertheless, the


FIG. 2. Workflow with the methodology used in our study. See Materials and Methods for a more comprehensive description.

fact that Helitrons in general did not group closer to any other major clade, indicates that Hel domains are only distantly related to genomic Pif1 helicases and belong to completely independent lineages. An interesting aspect of the Helentron/Helitron2 major clade is the presence of a Hel domain from the dinoflagellate Symbiodinium microadriaticum (CAE7237458.1) branching externally to the divergence of Helitron2 and Helentron sequences (fig. 3; supplementary fig. S2, Supplementary Material online). This RepHel lacks the apurinic-apyrimidinic (AP) endonuclease domain typical of Helentrons, and the element corresponding to this transposase (CAJNJV010003184.1) is structurally more similar to a Helitron2 variant (fig. 1). Hence, this Helitron2-like element appears to represent an intermediate variant that should be more closely related to the common ancestor of Helentron and Helitron2 elements. To our knowledge, this is the first identification of a putative evolutionary intermediate between two Helitron variants. In this specific case, the putative intermediate variant was not identified before most likely because the S. microadriaticum sequence (CAE7237458.1) was submitted only recently (February 2021).

One of the prokaryotic sequences in the Eukaryotic major clade is a Pif1-like helicase from a Rickettsiales bacterium (MBO87943.1), positioned before the radiation including most eukaryotic Pif1 sequences (fig. 3). Most phylogenetic analyses conducted to date place the order Rickettsiales as the closest relative of mitochondria (reviewed in Roger et al. [2017]). Although this hypothesis has been challenged by some studies (Roger et al. 2017; Martijn et al. 2018), a recent analysis that used more robust methods confirmed the close relationship between Rickettsiales and the mitochondrion ancestor (Fan et al. 2020). Hence, the topology observed in our phylogeny seems to reflect the known evolutionary link between eukaryotic Pif1 proteins and their prokaryotic ancestor, which probably belonged to the symbiont that later gave rise to mitochondria (Bochman et al. 2011).

Another marked feature observed in our phylogeny is the presence of Pif1-like sequences from three eukaryotic species (*Perkinsela* sp., *Phytomonas* sp., and *Strigomonas* culicis) preceding the prokaryotic radiation within the Eukaryotic major clade (fig. 3; supplementary fig S2, Supplementary Material online). These sequences belong to kinetoplastids from the phylum Euglenozoa which, accordingly, is considered the group that diverged earliest during eukaryotic evolution (Cavalier-Smith et al. 2014). Although other kinetoplastid species are grouped separately from these three basal taxa (fig. 3), this distribution could be explained by the presence of



Fig. 3. Maximum-likelihood phylogeny of Pif1-like helicases. The resulting phylogeny includes Pif1-like helicases from *Helitron* variants, viruses, plasmids, and organisms, with seven major clades indicated around the tree. Specific taxa mentioned in the text are shown in branch tips. Kinetoplastids are marked with red stars and amoebae are marked with asterisks. Branches with <0.7 SH-aLRT statistical support were collapsed. The rooted tree with all taxa names and branch support values is shown in supplementary figure S2, Supplementary Material online.

multiple Pif1 paralogs in species from this class, which have been shown to encode up to eight Pif1-like genes (Liu et al. 2009; Bochman et al. 2010). If these three basal sequences represent some of the Pif1 paralogs adapted for kinetoplastidspecific functions (Bochman et al. 2010), a process of positive evolution following subfunctionalization, might have caused them to be artificially positioned externally in relation to other eukaryotic Pif1 helicases. In addition to kinetoplastids, other taxa also displayed a somewhat scattered distribution on the Eukaryotic major clade, instead of forming monophyletic clusters. For instance, amoebal Pif1 helicases were grouped in five separate clades (fig. 3). Interestingly, a scattered distribution of amoebal Pif1-like proteins was also observed in a previous study and it was explained as the result of horizontal gene transfer (HGT) and duplication events (Harman and Manna 2016). Also in the Eukaryotic major clade, eukaryotic viruses, mostly NCLDVs, were found dispersed in different clades, sometimes closer to eukaryotic and prokaryotic organisms than to other groups of viruses (fig. 3; supplementary fig. S2, Supplementary Material online). Although noteworthy, this result agrees with the growing evidence for multiple HGT events between these large viruses and a variety of organisms (reviewed in Barreat and Katzourakis [2021]).

Overall, the scattered topology observed for several taxa from the Eukaryotic major clade might have been the consequence of two main factors. First, as a result of our searching and selection method designed to retrieve Pif1-like helicases with the highest similarity to specific queries. Because we only selected the best results from each taxonomic group, and eukaryotes may have multiple Pif1 genes adapted for distinct functions, it is likely that our sampled sequences represent a mixture of paralogs and orthologs. Second, as a consequence of several HGT events between eukaryotes, prokaryotes, and

viruses. Eukaryotes have been involved in HGT exchanges not only with viruses, as mentioned above, but also with multiple prokaryotic groups and sometimes with distinct eukaryotic taxa (reviewed in Husnik and McCutcheon [2018] and Van Etten and Bhattacharya [2020]). Thus, it is possible that Pif1 genes have been horizontally transferred several times during the evolution of eukaryotes.

In the Prokaryotic major clade, cases of interspersed branches from bacteria, archaea, and phages were also abundant, and indicate that several HGT events involving Pif1-like genes have occurred between these taxa (fig. 3). Although horizontally transferred sequences represent a relatively small fraction of eukaryotic genomes, in prokaryotes, HGT has long been considered a primary source of new genes and a major driver of evolution. These gene exchanges are not limited to closely related organisms, as they have been shown to cross prokaryotic domains and sometimes occur between bacteria, archaea and viruses (reviewed in Koonin [2016]). Hence, based on our phylogenetic analysis, it is reasonable to conclude that Pif1-like helicases are also members of the large set of gene families that have been horizontally transferred among prokaryotic organisms. Regardless of the particular explanations for each case, the frequent grouping of relatively distant taxa observed in the Eukaryotic and Prokaryotic major clades indicates that, in addition to ordinary vertical inheritance of genes, other events (e.g., HGTs and gene duplications) have shaped the evolution of genomic Pif1 helicases extensively.

Other interesting results were also revealed by the phylogenetic analysis. For instance, the TraA and Myoviridae clades formed sister groups with good branch support (fig. 3; supplementary fig. S2, Supplementary Material online). This result suggests a closer than expected relationship between replicons with completely distinct modes of propagation, underscoring the highly dynamic modularity that is typical of MGEs. Finally, as previously indicated in our Blast results, a protein annotated as belonging to the archaeon genus Methanothrix (TFH49976.1) grouped with TraA relaxases from Proteobacteria species, more specifically in the Desulfobacteraceae family (Desulfobacteraceae bacterium and Desulfosarcina cetonica) (fig. 3; supplementary fig. S2, Supplementary Material online). To verify whether this TraA gene derives from an HGT event or misannotation, we first used its protein sequence (TFH49976.1) as a query in separate Blastp searches against bacteria and archaea in the nr database. In this case, the query was significantly more similar to bacterial than archaeal sequences. We also used the nucleotide sequence corresponding to the protein (accession number: SPBB01000211.1) as a query in Blastn searches against bacteria and archaea in the nucleotide collection (nr/nt) and Whole Genome Shotgun (WGS) contigs databases. In this case, no hits with significant similarity were found in archaea. The query displays a significant identity (up to 75%) to bacterial genes, although limited to short stretches that cover up to 15% of the query length. Furthermore, the contig corresponding to the query only contains the TraA gene without flanking sequences that could be used to determine if this gene was integrated into an archaeal genome. Therefore, this putatively archaeal TraA gene is significantly more similar to bacterial than archaeal sequences, both at the amino acid and nucleotide level. Because this sequence is part of a metagenome assembly (BioSample: SAMN11127048), the possibility of misannotation or contamination in this case is very likely. Together, our analyses indicate that this TraA gene is likely from a bacterial plasmid misannotated as belonging to an archaeon. Regardless of those considerations, knowing the host species of this protein sequence does not change the interpretation of our results.

NMDS Analysis

The estimated evolutionary divergence between sequences were used to represent their distances in two dimensions with nonmetric multidimensional scaling (NMDS) analysis. By doing so, we intended to visualize their spatial arrangement without assuming cladistic relationships, and also verify if their distribution replicates the overall topology observed in the phylogeny.

The arrangement of Pif1-like helicases in the resulting NMDS ordination showed an overall segregation of proteins into seven major clusters (fig. 4). It also displayed a large divergence between Hel domains from the two major clades previously observed in our phylogeny (fig. 3), with *Helentron* and *Helitron2* sequences forming a single group distinctly segregated from *Helitron* variant sequences. In addition, *Helitron* Pif1-like domains from all variants did not appear to be more closely associated with any other specific major group, being roughly equidistant from genomic and viral helicases found in prokaryotes and eukaryotes (fig. 4).

Pif1 helicases from the Eukaryotic and Prokaryotic major groups formed two separate, albeit closely related clusters. Although genomic Pif1 helicases in the Eukaryotic group showed a tendency for clustering with sequences from more closely related taxa, in the Prokaryotic group, sequences from bacteria and archaea displayed a highly interspersed distribution. In both major groups viral sequences were mostly scattered among genomic Pif1 helicases (fig. 4). These distinct arrangements in the Eukaryotic and Prokaryotic major groups confirm the taxonomic incongruences and complex evolutionary history of genomic Pif1 helicases indicated by the phylogenetic analysis.

In sum, the resulting NMDS ordination recapitulates the main features observed in the phylogeny, that is, the segregation of seven major clades, the distant relationship between Hel domains from *Helitrons* and genomic helicases, and the indication of multiple HGT events involving Pif1-like helicases from eukaryotes, prokaryotes, and viruses.

Reassessing the Classification and Number of Pif1 Genes in Eukaryotes

As we have mentioned, Blastp searches on eukaryotic genomes using Pif1 helicases as queries often result in multiple significant hits. Because *Helitrons* are pervasive in most eukaryotic groups and their transposase includes a Pif1-like Hel domain, it is always possible that some of those hits constitute *Helitron* coding sequences, instead of genomic helicases. For example, during our preliminary analyses we



Fig. 4. NMDS plot of Pif1-like helicases. NMDS ordinations representing Euclidean distances between Pif1-like helicase sequences in two dimensions.

performed a Blastp search to identify putative genomic Pif1 helicases in the fungus *Rhizophagus clarus*, using the human Pif1 domain (6HPH_A) as a query, and found many candidate genes, together with RepHel sequences. However, a more detailed inspection revealed that some putative genomic Pif1 helicases are in fact Hel domains from *Helitron* coding sequences lacking the Rep domain in the same ORF (supplementary fig. S1, Supplementary Material online). Thus, without more careful analyses, the structural resemblance between genomic and *Helitron*-derived Pif1 domains can hinder the proper identification of sequences from this protein family. Indeed, to avoid classifying Hel domains as genomic Pif1 helicases, we excluded all species with *Helitrons* in their genomes from our initial Blast searches.

Although some eukaryotes are thought to have multiple genomic Pif1 helicases (Bochman et al. 2010, 2011; Harman and Manna 2016), most species from this domain of life apparently encode one or two Pif1 genes (Bochman et al. 2010). Considering that distantly related eukaryotes like *Schizosaccharomyces pombe* and humans only need one Pif1 helicase to carry out genomic functions, species with supposedly multiple Pif1 paralogs should be evaluated carefully. Thus, we reassessed three cases in the literature referring to genomic Pif1 genes from eukaryotes, which could have included *Helitron*-derived sequences inadvertently.

In the first example, Arabidopsis thaliana was described as having three genomic Pif1 helicases (CAB91581, NP_190738, and CAB63155) (Bochman et al. 2010). After examining the structure and sequence of these proteins we found that all of them are either RepHel proteins or Pif1-like sequences with significant identity to *Helitron* transposases (supplementary table S2, Supplementary Material online). Interestingly, a phylogeny of Pif1 sequences presented in the same work (Figure 1 in Bochman et al. 2010) displays a single Pif1 helicase from Oryza sativa (ABB47755) grouped together with the three A. thaliana proteins mentioned above. Because these three proteins were shown to be derived from RepHel transposases, and Helitrons are known to be abundant in the genomes of A. thaliana and O. sativa (Yang and Bennetzen 2009; Xiong et al. 2014), we examined this Pif1-like sequence from O. sativa. After inspecting its structure, we found that this O. sativa Pif1-like protein represents a RepHel transposase containing both of its major domains (supplementary table S2, Supplementary Material online). Hence, all these four proteins classified as genomic Pif1 helicases from A. thaliana and O. sativa constitute either RepHel transposases or Pif1-like Hel domains from Helitrons.

In the second example, the fungal pathogen of insects *Metarhizium robertsii* ARSEF 23 (formerly *M. anisopliae* ARSEF 23) was described as the eukaryote harboring the largest number of Pif1 genes, with 23 paralogs (Bochman et al. 2011). We conducted a Blastp search on the genome of this species using the human Pif1 domain (6HPH_A) and the *S. cerevisiae* Pif1 (NP_013650.1) as queries and found that, although *M. robertsii* appears to have up to 25 proteins with some similarity to Pif1 helicases, only 16 of them cannot be readily classified as RepHel transposases, that is, do not contain a Rep domain sequence. Of these 16 proteins, 11 either display significant similarity to RepHel transposases or belong to a cryptic RepHel ORF (truncated transposases with a Rep sequence upstream the Pif1 ORF), and one does not correspond to a Pif1 helicase (supplementary table S3,

7

MBE

Supplementary Material online). Hence, only four helicases from *M. robertsii* could represent genomic Pif1 candidates, with the other 20 Pif1-like sequence clearly being derived from *Helitron* transposases.

In the third example, it was suggested based on in silico analyses that A. thaliana could have up to 11 Pif1 genes (Knoll and Puchta 2011), with this large number of paralogs being attributed to Helitrons capturing and multiplying genomic Pif1 sequences. However, after inspecting all A. thaliana Pif1-like proteins on GenBank, retrieved after a Blastp searches using the human Pif1 domain (6HPH_A) and the S. cerevisiae Pif1 (NP_013650.1) as queries, we found that all of them either represent RepHel proteins directly or derive from Helitron transposases (supplementary table S4, Supplementary Material online). Although we anticipated that some sequences would derive from Helitrons, the fact that all retrieved A. thaliana Pif1-like proteins appear to represent RepHel transposases directly or indirectly was unexpected, considering the widespread distribution of genomic Pif1 helicases in eukaryotes. To investigate whether this apparent lack of genomic Pif1 homologs is exclusive from A. thaliana, we conducted a Blastp search using the same method on O. sativa, which is estimated to have diverged from A. thaliana \sim 163 Ma (Li et al. 2019). Like what was observed in A. thaliana, we found many Pif1-like sequences in O. sativa, with all results representing RepHel transposases directly or indirectly (supplementary table S5, Supplementary Material online).

Given the distant relationship between A. thaliana and O. sativa, we tried to estimate when genomic Pif1 helicases could have been lost during the evolution of these land plant lineages. To do that, we conducted a series of Blastp searches on taxonomic ranks above A. thaliana and O. sativa using the human Pif1 domain (6HPH_A) and the yeast Pif1 (NP_013650.1) as queries. Interestingly, genomic Pif1 homologs appear to have been lost in Brassicales and commelinids, the taxonomic groups from which A. thaliana and O. sativa belong, respectively (fig. 5). The best hits within these groups corresponded to RepHel proteins (supplementary table S6, Supplementary Material online). Conversely, the best hits from searches in taxa outside Brassicales (malvids) and commelinids (Liliopsida) were Pif1 proteins with low similarity to RepHel transposases, despite some of the species with putative genomic Pif1 helicases also having Helitron proteins (supplementary table S6, Supplementary Material online). To further confirm the absence of genomic Pif1 homologs in the mentioned groups, we first used the best hits from searches in malvids (EOX92974.1) and Liliopsida (MQL92731.1) as queries in Blastp searches against Brassicales and commelinids, respectively. The results still indicated a lack of genomic Pif1 homologs in Brassicales and commelinids, as the best hits also corresponded to Helitron sequences (supplementary table S7, Supplementary Material online). Additionally, we conducted Blastn searches using the nucleotide sequences corresponding to EOX92974.1 (CM001879.1) and MQL92731.1 (NMUH01001479.1) as queries against Brassicales and commelinids, respectively. Although the MBE

search against commelinids did not retrieve hits with significant similarity to the genomic Pif1 from Liliopsida, the result from Brassicales revealed a hit in *Bretschneidera sinensis* (JACXJD010000007.1) with 74% identity to the genomic Pif1 nucleotide sequence from malvids. This hit from *B. sinensis* translates to an ORF that appears to be intact, therefore representing a Pif1 gene that has not been annotated yet, which explains its absence in Blastp results. Interestingly, *B. sinensis* (family Akaniaceae) belongs to the most basal clade from Brassicales (Edger et al. 2018), indicating that genomic Pif1 homologs were probably lost shortly after the origin of this order and before the major radiation that gave rise to most extant families of Brassicales.

Although regions flanking genomic Pif1 helicases from malvids and Liliopsida up to tens of kilobase pairs on both sides display similarity to Brassicales and commelinids sequences, this similarity covers only limited portions of their length, as indicated by Blastn searches. Because this observed similarity is not contiguous over the whole span of flanking sequences, it is not possible to define whether they correspond to homolog regions, and therefore we could not determine what caused Pif1 genes to be lost in Brassicales and commelinids. However, it is noteworthy that most of the genomic Pif1-flanking regions with significant identity to sequences from both groups correspond to TEs, particularly LTR retrotransposons, as determined by searches using the Censor tool in Repbase (Kohany et al. 2006). Although with the current data presented it is not possible to ascertain what caused genomic Pif1 helicases to be lost in Brassicales and commelinids, the presence of long TE sequences in the vicinity of those genes in the closest taxonomic groups could be related to these events. For instance, TEs flanking these Pif1 genes could have promoted ectopic recombinations between insertions, leading to the deletion of large chromosome segments in Pif1 gene loci (Kent et al. 2017). However, more extensive analyses would be necessary to pinpoint the precise boundaries of these deleted chromosomal segments and to describe the mechanisms responsible for those events. Nonetheless, our results indicate that at least two major groups of land plants appear to have lost genomic Pif1 homologs independently (fig. 5) and that usual functions performed by this gene might be carried out by different proteins in species from these taxa.

Discussion

The Evolutionary History of Helitrons Takes Shape

Because Pif1 helicases are known to be typically eukaryotic proteins (Bochman et al. 2010), and Hel domains found in some RepHel transposases have introns, it has been suggested that an *Helitron* ancestor likely captured a Pif1 gene from its eukaryotic host (Kapitonov and Jurka 2001, 2007; Thomas and Pritham 2015). However, our results indicate that *Helitrons* already encoded a Hel domain before invading eukaryotic genomes (fig. 6), as genomic Pif1 helicases from prokaryotes and eukaryotes formed sister groups in our analyses, with Pif1-like Hel domains being only distantly related to

Pif1 Helicases and the Origin of Helitrons · doi:10.1093/molbev/msab334



Fig. 5. Cladogram of plant groups that appear to have lost genomic Pif1 helicases. Only major clades are represented, with Poales and Brassicales indicating the orders of *O. sativa* and *A. thaliana*, respectively. Red bars mark the two branches that lack sequences with significant similarity to genomic Pif1 helicases. Phylogeny adapted from Li et al. (2019).



FIG. 6. A hypothesis for the evolution of *Helitrons*. We propose that *Helitrons* descend from prokaryotic plasmid-like elements (first box) that invaded eukaryotic cells during their early evolution. After invading eukaryotes, *Helitrons* shifted to a predominantly transposon-like mode of propagation. During their subsequent adaptation to specific hosts, *Helitrons* diverged into distinct variants (*Helitrons, Helentrons, and Helitron2*) and captured additional domains. Arrows represent major steps during the evolution of *Helitrons*.

MBE

them. Nonetheless, in addition to a RepHel with its archetypal double-domain structure, Helentrons also have an AP endonuclease domain in their transposase (fig. 1), which was probably captured from a non-LTR retrotransposon residing in the same eukaryotic host (Thomas and Pritham 2015). The capture of an AP endonuclease gene likely marked the evolutionary origin of Helentrons from Helitron2-like ancestors, which also gave rise to the Helitron2 variant. Our identification of an intermediate Hel domain from S. microadriaticum branching externally to Helentron and Helitron2 sequences constitute the first direct evidence for a Helitron2-like element as the ancestor of both variants. Besides the AP endonuclease from Helentrons, several other domains have been incorporated to specific Helitron lineages during their evolution in eukaryotic genomes (Thomas and Pritham 2015) (fig. 6). However, the function of AP endonucleases and other coding sequences captured by Helitrons from eukaryotes have not been determined yet.

Although the evolutionary proximity of Helentron and Helitron2 lineages was expected (Thomas and Pritham 2015; Heringer and Kuhn 2018), our results indicating that Hel domains from the Helitron variant form a distinct group from the Helentron and Helitron2 variants (figs. 3 and 4) contrasts with the monophyletic distribution previously observed for Helitron Rep domains (Poulter et al. 2003; Heringer and Kuhn 2018). Assuming the more parsimonious scenario in which Helitrons constitute a monophyletic group, the resulting paraphyletic distribution of Hel domains might have been caused by faster evolutionary rates that occurred on this protein region. The same topology was not observed for Rep domains in previous studies, probably due to a higher tendency for amino acid sequence conservation in this portion of Helitron transposases. If Hel domains evolved under less constrained evolutionary pressures or went through a stronger positive selection imposed by their hosts, these processes could have potentially masked their monophyletic nature. Furthermore, the widespread distribution of Helitrons in eukaryotes (Thomas and Pritham 2015) and the overall similarity between RepHel and host phylogenies, indicate that Helitrons began to diverge before the emergence of most eukaryotic kingdoms (Poulter et al. 2003). As time estimates of major eukaryote radiations date back to approximately 1 billion years ago (Douzery et al. 2004; Berney and Pawlowski 2006), the first Helitron lineage divisions likely have a similar age. Thus, a rapid evolution of Hel domains that occurred through a very long period of time might have contributed to blur the monophyletic nature of Helitrons in our analyses.

An independent example supporting the hypothesis that each domain from RepHel proteins have evolved under distinct evolutionary pressures can be viewed in the phylogenies of *Helitron* Rep and Hel domains inferred by Poulter et al. (2003), which present distinct topologies. In their Rep domain phylogeny, *Helitron* sequences from the fungus *Phanerochaete chrysosporium* clustered with *Helentrons*, instead of *Helitrons*. Conversely, in the Hel domain phylogeny, all elements segregated into variant-specific clades, indicating that distinct *Helitron* variants display a more pronounced sequence divergence in this region. Furthermore, in the Hel

MBE

phylogeny, *Helitron* clades were connected by relatively longer branches when compared with the Rep domain tree, similarly to the observed between our results presented here for Hel domains (supplementary fig. S2, Supplementary Material online) and on our previous study involving Rep domains (Heringer and Kuhn 2018). It is worth mentioning that, in contrast to our phylogeny, the one presented by Poulter et al. (2003) did not display a polyphyletic distribution for Hel domains. The reason for that might be related to the smaller sample size and diversity of *Helitrons* used in the latter analysis when compared with the one presented here.

Altogether, these observations suggest that each domain from RepHel transposases has evolved under distinct evolutionary rates. These differences could be derived from selective pressures that constrained the Rep amino acid sequence to a higher degree, and/or favored a more rapid evolution of the Hel domain to optimize its interaction with host components. Hence, a very early radiation of *Helitrons*, combined with relatively faster evolutionary rates that have occurred in Hel domains since they first invaded eukaryotes, probably explain the spurious paraphyletic distribution between major *Helitron* groups in our results. In this case, the observed topology could represent a result of long-branch attraction (Bergsten 2005).

In summary, our phylogenetic and NMDS analyses indicate that RepHel proteins evolved independently from genomic Pif1 helicases found in prokaryotes and eukaryotes. Thus, in spite of previous hypotheses about the origins of Hel domains, it is unlikely that a *Helitron* ancestor captured a Pif1 gene from its eukaryotic host. Instead, we suggest that, before entering eukaryotic cells, *Helitrons* already encoded RepHel proteins, branching into two major lineages after they invaded eukaryotic genomes (fig. 6). From there on, Hel domains probably evolved under relatively faster rates, which could explain their distribution into marked separate groups, in contrast to what was observed in analyses of Rep domains (Poulter et al. 2003; Heringer and Kuhn 2018).

Helitrons May Be Descendants of Plasmid-Like Elements

Although it seems clear that neither Rep nor Hel domains have originated from genomic proteins, the ancestor of Helitrons probably resided within a prokaryotic cell. If this ancestor already had a transposon-like mode of propagation, it is conceivable that their descendants (or their remnants) could still reside in genomes of some unknown prokaryote lineages. However, even assuming the hypothesis of a transposon ancestor as correct, it is unlikely that such elements would be found, as sequences that do not benefit cellular functioning directly (like TEs) are subject to extremely rapid turnover rates in prokaryotes (Sela et al. 2016; Wolf et al. 2016). A second possibility is that prokaryotic ancestors of Helitrons had a predominantly plasmid-like mode of replication before they became eukaryotic TEs. This scenario not only agrees with the current lack of Helitron-like sequences in prokaryotes, but with the close relationship found between Rep domains from Helitrons and RC bacterial plasmids (Heringer and Kuhn 2018; Kazlauskas et al. 2019) and the fact that *Helitrons* generate plasmid-like intermediates during transposition (Grabundzija et al. 2018).

It is worth mentioning that a TraA relaxase was the only protein from a MGE retrieved in our Blast searches using Hel domains as queries. Similarly to RepHel transposases, TraA and other plasmid relaxases possess Rep-like and helicase domains within the same protein (Pérez-Mendoza et al. 2006; Chandler et al. 2013). Although Rep-like domains found in relaxases display an inverted orientation of their main catalytic motifs when compared with RepHel transposases, both enzymes have an overall similar architecture, consisting of a Rep followed by a helicase domain. In addition, despite their inverted orientation, the 3D topology of these motifs in relaxases and RCR proteins is essentially the same (Chandler et al. 2013). Interestingly, the cryo-EM structure of the RepHel in complex with the Helitron 5'-end ssDNA was solved only recently, revealing an even higher degree of organizational similarity with relaxases, particularly with Tral (Kosek et al. 2021). As mentioned by the authors, the structural similarity between these two classes of proteins does not imply a close evolutionary relationship, which is also supported by our results and previous studies involving the Rep domain (Heringer and Kuhn 2018; Kazlauskas et al. 2019). If these structural resemblances are most likely the result of convergent evolution, they would suggest the existence of functional parallels between relaxases and RepHel transposases. Nonetheless, the fact that a group of relaxases was retrieved in our searches by sequence similarity with Hel domains from Helitrons could still indicate a distant evolutionary relationship between these proteins.

Based on these considerations, we propose that *Helitrons* descend from prokaryotic plasmid-like elements that shifted to a transposon mode of propagation after invading eukaryotic cells (fig. 6). Importantly, a transition from an RCR plasmid to an RC TE would likely not require major adaptations, as the replicative processes employed in both types of MGEs work by the same basic enzymatic steps, only differing in the number of DNA substrates and type of final products involved (Chandler et al. 2013; Wawrzyniak et al. 2017).

What Is the Function of Pif1 Helicases in Helitrons?

Experimental assays revealed that Helitrons have to generate dsDNA circle intermediates in order to transpose, as ssDNA circular elements transfected into human cells were not viable substrates for host genome integration (Grabundzija et al. 2018). The formation of dsDNA intermediates could be achieved by the concomitant synthesis of leading and lagging strands while the element's leading strand is being "peeledoff," or by the addition of a short lagging strand primer on the unwound leading strand before an ssDNA circle is formed. In either case, these processes would require the recruitment of replication fork and DNA repair machinery components (Grabundzija et al. 2018), both of which Pif1 helicases are part of Bochman et al. (2010) and Muellner and Schmidt (2020). For instance, Pif1 stimulates the activity of DNA polymerase δ (Pol δ) during DNA repair and replication (Pike et al. 2009; Wilson et al. 2013; Koc et al. 2016) through its interaction with the proliferating cell nuclear antigen (PCNA) (Wilson et al. 2013; Buzovetsky et al. 2017; Dahan et al. 2018). In addition, Pif1 has a role in fork convergence, resolving the stalling of these structures, which are expected to occur in the final stages of linear and circular DNA replication (Deegan et al. 2019). Another relevant feature of Pif1 helicases is their preference for binding and unwinding forked structures (dsDNA with ssDNA overhangs) (Ramanagoudr-Bhojappa et al. 2013; Li et al. 2016), which are substrates expected to be formed in the first stages of RCT, when RepHel nicks the *Helitron*'s leading strand in its 5'-end (Dias et al. 2016; Grabundzija et al. 2016, 2018).

The combination of those Pif1 attributes suggests that the Hel domain could aid in the RepHel association to forked DNA structures during the initial steps of transposition and help to recruit replication machinery components from hosts (e.g., PCNA and Pol δ). Although prokaryotic RC TEs, which are thought to transpose similarly to Helitrons, do not encode helicases, it is possible that a Hel domain merged to a Rep protein confers mechanistic advantages for RCT in eukaryotic cells and maybe is essential in this environment. Indeed, it has been shown that a mutation in the Walker A motif from Hel domains causes Helitrons to lose their transposition activity in cells (Grabundzija et al. 2016). In addition, the RepHel cryo-EM structure reveals a considerable interface between the catalytic portion of Rep and the Hel domain, suggesting that they act in conjunction to unwind dsDNA and generate sufficient ssDNA to allow strand cleavage as transposition starts (Kosek et al. 2021). Thus, it is conceivable that a Hel domain also favored the invasion and colonization of eukaryotic genomes by Helitrons, which would explain their pervasiveness in this domain of life that lacks other groups of RC TEs.

Additionally, the Hel domain could facilitate the final stages of transposition, when the RepHel associated with a circular intermediate binds its target site before integration. In contrast to prokaryotic RC TE insertions, which are guided by site specificity (Garcillán-Barcia et al. 2002), Helitrons integrate between AT, TT, or TC dinucleotides, depending on the variant, with no preference for unique sequences (Thomas and Pritham 2015). Hence, the RepHel in complex with a Helitron intermediate could initially bind its target site by associating with specific DNA or chromatin structures, instead of using sequence guided recognition. In this case, an initial contact would be favored by the known affinity of Pif1 helicases to DNA secondary structures typically found in recombination sites and gene promoters (Bochman et al. 2012; Byrd and Raney 2015; Muellner and Schmidt 2020). Indeed, experimental assays revealed that active Helitrons appear to preferentially target highly expressed gene regions (Grabundzija et al. 2016). After a structure-based association mediated also by Hel, the Rep domain would be able nick the recipient strand at a nearby AT, TT or TC dinucleotide site, before transferring an ssDNA intermediate to the host's chromosome, forming a heteroduplex and completing transposition (Kapitonov and Jurka 2007; Thomas and Pritham 2015; Dias et al. 2016).

Taken together, these features of Pif1 helicases and *Helitrons* appear to agree with a scenario in which Hel domains play a more sophisticated role during RCT, beyond simply unwinding double-stranded DNA elements. The

presence of a Pif1-like Hel domain in *Helitron* transposases may have provided an advantage over the recruitment of host helicases, by concatenating the processes of DNA binding, leading strand nicking, and peeling-off, together with the formation of circular dsDNA intermediates, all conducted by the same enzyme. In addition, Hel domains could aid the association between RepHel-dsDNA intermediates and target sites on host chromosomes.

Helitrons Can Hamper the Identification of Eukaryotic Pif1 Helicases

The abundance of *Helitrons* in eukaryotic genomes, together with the general similarities between *Helitron* Pif1-like Hel domains and genomic Pif1 helicases from eukaryotes, make their distinction by in silico methods complicated. Our reevaluation of three examples in the literature describing Pif1 proteins from *A. thaliana, O. sativa,* and *M. robertsii* demonstrated how these problems have affected the classification and number estimation of genomic Pif1 helicases in eukaryotic species. In these cases, most, or all putative genomic Pif1 helicases described were shown to represent *Helitron*-derived sequences.

Interestingly, during our searches for genomic Pif1 candidates in A. thaliana and O. sativa we found that all Pif1-like proteins from these species either represent complete Helitron transposase sequences or Hel domains from broken RepHel ORFs. After investigating higher taxonomic ranks from which A. thaliana and O. sativa belong (Brassicales and commelinids, respectively), we found that both of them appear to have lost genomic Pif1 homologs independently (fig. 5). Even granting that Brassicales and commelinids may have genomic Pif1 homologs that went undetected in our searches, the fact that RepHel sequences represented the best hits to eukaryotic Pif1 helicases points to a similar evolutionary pattern in those distantly related groups. However, this issue should be further investigated to determine in more detail how the Pif1 family have evolved in land plants and if some of them have different proteins to perform the same functions of genomic Pif1 helicases.

Despite the examples described above, some eukaryotes have multiple bona fide genomic Pif1 helicases. As we have mentioned, kinetoplastids encode several Pif1 paralogs that likely participate in distinct functions related to their unique biology (Liu et al. 2009; Bochman et al. 2010). Furthermore, Helitron transposases are not found in kinetoplastid genomes. as indicated by our Blast searches and a previous analysis (Thomas and Pritham 2015). Hence, all Pif1 helicases found in this group might consist of genomic representatives derived from gene duplications. In addition to kinetoplastids, some amoebae also have multiple genomic Pif1 helicases, with Acanthamoeba castellanii encoding up to nine Pif1 genes (Harman and Manna 2016). Our Blast searches revealed that these amoebae species do not have RepHel sequences in their genomes, which confirms that these proteins indeed represent genomic Pif1 helicases. Thus, kinetoplastids and amoebae are the only eukaryotic groups so far in which there is solid evidence for species with more than two genomic Pif1 paralogs.

Altogether, it is clear that our knowledge about the distribution and number of genomic Pif1 helicases in eukaryotes is relatively limited to a small number of species. As we have shown, some of the attempts to identify genomic Pif1 proteins in eukaryotes have been hampered by the large amount of *Helitron* transposases found in this domain of life. It will be important to establish a reliable and efficient method to correctly discriminate between these two major groups of Pif1 helicases, before they are studied in large-scale analyses.

Conclusion

Although the similarity between Hel domains and genomic Pif1 helicases has been noted since the discovery of Helitrons 20 years ago, no study had explored their evolutionary connections. Despite previous suggestions that an Helitron ancestor likely acquired the Hel domain by capturing a Pif1 gene from its eukaryotic host, our results indicate that RepHel proteins already had their archetypal structure with two domains before invading eukaryotes. Furthermore, considering phylogenetic, structural, and mechanistic aspects of these elements, we propose that Helitron ancestors probably had a plasmid-like mode of replication in prokaryotic hosts, before invading eukaryotes and shifting into a transposon. Based on the known features of Pif1 helicases and RepHel proteins, we also hypothesize that Hel domains likely perform a more complex function during transposition, beyond simply unwinding Helitron double-stranded DNA.

In addition, our reassessment of the literature describing eukaryotic Pif1 helicases revealed that many of these examples actually represent complete or partial RepHel transposases from *Helitrons*, which are commonly abundant in eukaryotic genomes. This finding highlights the need for a careful inspection before classifying Pif1-like proteins as genomic helicases in eukaryotes, particularly in species that appear to harbor multiple Pif1-like genes. We also found that two distantly related groups of land plants appear to lack genomic Pif1 homologs, despite having multiple Pif1-like Hel domain sequences derived from *Helitrons*. This observation should be studied in more detail, as Pif1 helicases have been considered essential in many genomic processes that are conserved in all eukaryotes studied to date.

Materials and Methods

Selection of RepHel Sequences

We used RepHel protein sequences obtained in our previous study (Heringer and Kuhn 2018), belonging to the three main *Helitron* variants (*Helitron, Helentron,* or *Helitron*2) (Thomas and Pritham 2015), as initial queries in a series of Blastp searches on the nonredundant protein sequences (nr) database from GenBank (Sayers et al. 2019). With this strategy, we were able to retrieve a sample with a larger variety of RepHel representatives, thus enabling the generation of more accurate consensus sequences of each domain (Rep and Hel). Each one of the initial 13 *Helitron* protein sequences was used as a query to select an additional RepHel, which in turn, was used as a query to select another sequence in a second Blastp search round. In each of these searches the best hit, sorted

MBE

44

by Max Score, was selected, excluding sequences found in genomes of the same genus in a previous round. For the Helitron2 variant we applied four rounds of consecutive searches to increase the number of sequences, as it had a single representative in our previous analysis (Heringer and Kuhn 2018). To determine whether the additional RepHel sequences belonged to the same variant as the initial queries. we visually inspected their structure with the Conserved Domain Database (CDD) search tool (Lu et al. 2020), following the classification provided by Thomas and Pritham (2015). This classification considers differences in amino acids within conserved regions from the Rep domain and the presence or absence of specific domains in the RepHel protein. A total of 41 RepHel protein sequences were selected for further analyses: 18 from Helitrons, 18 from Helentrons, and 5 from Helitron2 elements. Sequences from Helitron and Helentron/ Helitron2 variants were aligned separately using the auto mode from the MAFFT online service (Katoh et al. 2019). Helentron and Helitron2 sequences were aligned as a single group because these variants are known to be closely related (Thomas and Pritham 2015; Heringer and Kuhn 2018). Rep and Hel domains from each protein were isolated and trimmed, keeping only well-defined conserved regions among aligned sequences. These conserved regions were used to generate consensus sequences of each domain from Helitron and Helentron/Helitron2 variants, considering the most common amino acid in each site (supplementary data S1, Supplementary Material online), using the Advanced Consensus Maker tool from the HIV Database (https://www.hiv.lanl.gov/content/sequence/CONSENSUS/ AdvCon.html; last accessed November 16, 2021).

Stepwise Search and Selection of Helicase Protein Sequences

The Hel domain consensus sequences of Helitron and Helentron/Helitron2 variants (supplementary data S3, Supplementary Material online) were used as queries in Blastp searches against the nr database from GenBank (Sayers et al. 2019), which includes all available annotated proteins for a given taxa. A sample of protein sequences representing a wide variety of organisms were retrieved from distinct taxonomic levels, depending on their number of resulting hits in preliminary Blastp searches. For example, in eukaryotes, searches were conducted from the kingdom down to the class level, as this domain displayed a large number of significant results distributed heterogeneously across thousands of genomes. Conversely, in bacteria we conducted searches at the phylum level, and in archaea the whole sample was retrieved at the domain level itself. The best hits (sorted by Max Score) from Blastp searches using consensus sequences of both Helitron and Helentron/Helitron2 variants were selected. Each species containing best hits had one or two protein sequence representatives selected, depending on whether searches using different variant consensuses retrieved the same or different best hits, respectively. To verify if Helitrons were present in the genomes of species containing selected hits, we carried out a second round of searches in these taxa, this time using Rep consensus sequences as queries. Blastp searches were conducted against the nr database and tBlastn searches were conducted against the WGS contigs database. Because the aim of our study was to investigate the relationship between Hel domains from Helitrons and genomic Pif1 helicases, taxa containing hits corresponding to Rep sequences in any of the two searches (Blastp or tBlastn) were excluded at this stage. By doing so, we expected to have avoided the inclusion of helicases derived from Helitrons during the retrieval of putative genomic helicases, which could result in false phylogenetic inferences. Using these criteria, we were able to select 76 Pif1-like sequences from a wide variety of organisms lacking Rep sequences in their genomes. To expand our sample, we used Hel domain consensus sequences and the S. cerevisiae Pif1 (NP_013650.1) as queries in Blastp searches against the same groups of organisms from the previous analysis, this time without filtering taxa with Rep sequences in their genomes and including eukaryotic and prokaryotic viruses. Because Pif1-like proteins selected in the initial searches could be more readily identified as either genomic or Helitron-derived helicases, they were used to aid in the classification of sequences retrieved without the Rep-filtering procedure by their relationship revealed later in the phylogenetic analysis.

Alignment and Isolation of Helicase Domains

Helicase sequences from each major taxon group (Eukaryota, Bacteria, Archaea, plasmids, eukaryotic, and prokaryotic viruses) were aligned separately with the Hel domain consensus sequences from Helitrons and Helentrons/Helitron2 using the auto mode from the MAFFT online service (Katoh et al. 2019) in order to identify a common region among them. Sequences that aligned poorly or displayed large gaps on conserved regions were excluded using the MAFFT data set refinement tool also available in the MAFFT online service (Katoh et al. 2019). Segments extending upstream and downstream the central conserved regions were visualized using MEGAX (Kumar et al. 2018) and trimmed to avoid spurious alignments between nonrelated portions of proteins. This procedure is important considering that a large majority of prokaryotic and eukaryotic proteins contain multiple domains that have evolved through modular rearrangements (Bornberg-Bauer et al. 2005; Wang and Caetano-Anollés 2009). Even among genomic Pif1-like domains from eukaryotes, there are low levels of sequence and size similarity in their N- and C-terminal regions extending beyond a conserved core (Boule and Zakian 2006). Thus, when conducting a phylogenetic analysis of highly divergent protein sequences, it is preferable to only consider limited domain regions as evolutionary units, because flanking segments can evolve through distinct selective constraints. A total of 310 helicases from Helitrons (65 sequences), eukaryotic (89 sequences) and prokaryotic organisms (56 sequences), plasmids (10 sequences), eukaryotic viruses (48 sequences), and prokaryotic viruses (42 sequences) were selected for the next step of our analyses (supplementary table S1, Supplementary Material online). Trimmed helicase domains from all taxa, including Helitrons, were aligned using the E-INS-i method combined with mafft-homologs in the MAFFT online service (Katoh et al. 2019). The final alignment containing all sequences used in the following analyses are available in supplementary data S2, Supplementary Material online.

Phylogenetic and NMDS Analyses

The best-fit evolutionary model for the alignment (LG + G + I) was selected using the smart model selection in PhyML (Lefort et al. 2017). The maximum likelihood phylogeny of aligned amino acid sequences was inferred with the SPR method of tree topology search, six random plus one parsimony starting trees and six substitution rate categories across sites modeled with estimated gamma-shaped distribution parameter and proportion of invariant sites. Branch supports were estimated using the approximate likelihood ratio test (aLRT) with the nonparametric Shimodaira-Hasegawa correction (SH-aLRT). All these procedures were conducted on PhyML 3.1 (Guindon et al. 2010). Branches with <0.7 SHaLRT statistical support were collapsed using TreeGraph 2 (Stöver and Müller 2010) and the final tree visualized using FigTree v.1.4.2 (http://tree.bio.ed.ac.uk/software/figtree/; last accessed November 16, 2021). For the NMDS analysis, pairwise evolutionary distances between aligned sequences were estimated with the JTT matrix-based model and the rate variation among sites modeled with a gamma distribution on MEGAX (Kumar et al. 2018). NMDS ordinations with Euclidean distances of the sequences represented in two dimensions were generated using the R package vegan v2.5-6 (Dixon 2003). The NMDS analysis and plotting were executed in RStudio v1.3.959 (RStudio Team 2020) with R v4.0.0 (R Core Team 2020). All the methodology described heretofore is represented as a schematic workflow in figure 2.

Search and Classification of Pif1-Like Proteins in Eukaryotic Species

To reexamine selected examples from the literature describing genomic Pif1 helicases, which could in fact constitute RepHel-derived sequences, we inspected the structure of those proteins using the CDD search tool (Lu et al. 2020). To reassess the description of species containing multiple genomic Pif1 helicases we conducted Blastp searches in the protein sequences from the corresponding taxa available in the nr database from GenBank (Sayers et al. 2019) using the human Pif1 domain (6HPH_A) and S. cerevisiae Pif1 protein (NP_013650.1) as queries. In order to verify if the resulting sequences corresponded to RepHel transposases, all hits had their structural features inspected with the CDD search tool (Lu et al. 2020). Hits that did not included a conserved Rep domain identified by the CDD search tool were used as queries in a second round of Blastp searches against the nr database from GenBank to check if they might constitute Hel domains from broken Helitron transposases (Hel domains highly similar to RepHel proteins) or cryptic RepHel proteins (truncated transposase with a Rep sequence upstream the Pif1 ORF). If the best hits (sorted by Max Score) from this second round of searches corresponded to RepHel proteins, queries were considered as derived from Helitrons. In contrast, if the resulting best hits did not correspond to RepHel sequences, queries were classified as putative genomic Pif1 helicases.

Supplementary Material

Supplementary data are available at *Molecular Biology and Evolution* online.

Acknowledgments

We would like to thank Dr Guilherme B. Dias for the helpful comments on an earlier version of the manuscript and three anonymous reviewers whose comments and suggestions helped to improve several aspects of the paper. We are also grateful to Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (fellowship 308386/2018-3 to G.C.S.K.) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior—Brasil (CAPES) (doctoral fellowship to P.H.).

Data Availability

The data underlying this article are available in the article and in its supplementary material.

References

- Alt-Mörbe J, Stryker JL, Fuqua C, Li PL, Farrand SK, Winans SC. 1996. The conjugal transfer system of *Agrobacterium tumefaciens* octopinetype Ti plasmids is closely related to the transfer system of an IncP plasmid and distantly related to Ti plasmid vir genes. J Bacteriol. 178(14):4248–4257.
- Barreat JGN, Katzourakis A. 2021. Paleovirology of the DNA viruses of eukaryotes. *Trends Microbiol*. https://doi.org/10.1016/j.tim.2021.07. 004.
- Bergsten J. 2005. A review of long-branch attraction. *Cladistics* 21(2):163-193.
- Berney C, Pawlowski J. 2006. A molecular time-scale for eukaryote evolution recalibrated with the continuous microfossil record. Proc R Soc B. 273(1596):1867–1872.
- Bochman ML, Sabouri N, Zakian VA. 2010. Unwinding the functions of the Pif1 family helicases. DNA Repair 9(3):237–249.
- Bochman ML, Judge CP, Zakian VA. 2011. The Pif1 family in prokaryotes: what are our helicases doing in your bacteria? *Mol Biol Cell*. 22(12):1955–1959.
- Bochman ML, Paeschke K, Zakian VA. 2012. DNA secondary structures: stability and function of G-quadruplex structures. Nat Rev Genet. 13(11):770–780.
- Bornberg-Bauer E, Beaussart F, Kummerfeld SK, Teichmann SA, Weiner J. 2005. The evolution of domain arrangements in proteins and interaction networks. *Cell Mol Life Sci.* 62(4):435–445.
- Boule JB, Zakian VA. 2006. Roles of Pif1-like helicases in the maintenance of genomic stability. *Nucleic Acids Res.* 34(15):4147–4153.
- Buzovetsky O, Kwon Y, Pham NT, Kim C, Ira G, Sung P, Xiong Y. 2017. Role of the Pif1-PCNA complex in Pol δ-dependent strand displacement DNA synthesis and break-induced replication. *Cell Rep.* 21(7):1707–1714.
- Byrd AK, Raney KD. 2015. A parallel quadruplex DNA is bound tightly but unfolded slowly by Pif1 helicase. J Biol Chem. 290(10):6482-6494.
- Cavalier-Smith T, Chao EE, Snell EA, Berney C, Fiore-Donno AM, Lewis R. 2014. Multigene eukaryote phylogeny reveals the likely protozoan ancestors of opisthokonts (animals, fungi, choanozoans) and Amoebozoa. *Mol Phylogenet Evol.* 81:71–85.
- Chandler M, De La Cruz F, Dyda F, Hickman AB, Moncalian G, Ton-Hoang B. 2013. Breaking and joining single-stranded DNA: the HUH endonuclease superfamily. *Nat Rev Microbiol.* 11(8):525–538.
- Dahan D, Tsirkas I, Dovrat D, Sparks MA, Singh SP, Galletto R, Aharoni A. 2018. Pif1 is essential for efficient replisome progression through

MBE

lagging strand G-quadruplex DNA secondary structures. *Nucleic Acids Res.* 46(22):11847–11857.

- Deegan TD, Baxter J, Bazán MÁO, Yeeles JT, Labib KP. 2019. Pif1-family helicases support fork convergence during DNA replication termination in eukaryotes. *Mol Cell*. 74(2):231–244.
- Dias GB, Heringer P, Kuhn GCS. 2016. Helitrons in *Drosophila*: chromatin modulation and tandem insertions. *Mob Genet Elements* 6(2):e1154638
- Dixon P. 2003. VEGAN, a package of R functions for community ecology. J Veg Sci. 14(6):927–930.
- Douzery EJ, Snell EA, Bapteste E, Delsuc F, Philippe H. 2004. The timing of eukaryotic evolution: does a relaxed molecular clock reconcile proteins and fossils? *Proc Natl Acad Sci U S A*. 101(43):15386–15391.
- Edger PP, Hall JC, Harkess A, Tang M, Coombs J, Mohammadin S, Schranz ME, Xiong Z, Leebens-Mack J, Meyers BC, et al. 2018. Brassicales phylogeny inferred from 72 plastid genes: a reanalysis of the phylogenetic localization of two paleopolyploid events and origin of novel chemical defenses. Am J Bot. 105(3):463–469.
- Fan L, Wu D, Goremykin V, Xiao J, Xu Y, Garg S, Zhang C, Martin WF, Zhu R. 2020. Phylogenetic analyses with systematic taxon sampling show that mitochondria branch within Alphaproteobacteria. *Nat Ecol Evol.* 4(9):1213–1219.
- Feschotte C, Wessler SR. 2001. Treasures in the attic: rolling circle transposons discovered in eukaryotic genomes. Proc Natl Acad Sci U S A. 98(16):8923–8924.
- Garcillán-Barcia MP, Bernales I, Mendiola MV, De La Cruz F. 2002. IS91 rolling-circle transposition. In: Craig N, Craigie R, Gellert M, Lambowitz A, editors. Mobile DNA II. Washington, DC: ASM Press. p. 891–904.
- Grabundzija I, Messing SA, Thomas J, Cosby RL, Bilic I, Miskey C, Gogol-Döring A, Kapitonov V, Diem T, Dalda A, et al. 2016. A Helitron transposon reconstructed from bats reveals a novel mechanism of genome shuffling in eukaryotes. *Nat Commun.* 7:10716.
- Grabundzija I, Hickman AB, Dyda F. 2018. Helraiser intermediates provide insight into the mechanism of eukaryotic replicative transposition. Nat Commun. 9(1):1278.
- Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New algorithms and methods to estimate maximumlikelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol. 59(3):307–321.
- Harman A, Manna S. 2016. Identification of Pif1 helicases with novel accessory domains in various amoebae. *Mol Phylogenet Evol.* 103:64–74.
- Heringer P, Kuhn GCS. 2018. Exploring the remote ties between Helitron transposases and other rolling-circle replication proteins. Int J Mol Sci. 19(10):3079.
- Husnik F, McCutcheon JP. 2018. Functional horizontal gene transfer from bacteria to eukaryotes. Nat Rev Microbiol. 16(2):67–79.
- Kapitonov VV, Jurka J. 2001. Rolling-circle transposons in eukaryotes. Proc Natl Acad Sci U S A. 98(15):8714–8719.
- Kapitonov VV, Jurka J. 2007. Helitrons on a roll: eukaryotic rolling-circle transposons. *Trends Genet*. 23(10):521–529.
- Katoh K, Rozewicki J, Yamada KD. 2019. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief Bioinform*. 20(4):1160–1166.
- Kazlauskas D, Varsani A, Koonin EV, Krupovic M. 2019. Multiple origins of prokaryotic and eukaryotic single-stranded DNA viruses from bacterial and archaeal plasmids. *Nat Commun.* 10(1):3425.
- Kent TV, Uzunović J, Wright SI. 2017. Coevolution between transposable elements and recombination. *Phil Trans R Soc B*. 372(1736):20160458.
- Knoll A, Puchta H. 2011. The role of DNA helicases and their interaction partners in genome stability and meiotic recombination in plants. J Exp Bot. 62(5):1565–1579.
- Koc KN, Singh SP, Stodola JL, Burgers PM, Galletto R. 2016. Pif1 removes a Rap1-dependent barrier to the strand displacement activity of DNA polymerase δ . Nucleic Acids Res. 44(8):3811–3819.
- Kohany O, Gentles AJ, Hankus L, Jurka J. 2006. Annotation, submission and screening of repetitive elements in Repbase: repbaseSubmitter and Censor. BMC Bioinf. 7:474.

- Koonin EV. 2016. Horizontal gene transfer: essentiality and evolvability in prokarvotes, and roles in evolutionary transitions. F1000Res. 5:1805.
- Kosek D, Grabundzija I, Lei H, Bilic I, Wang H, Jin Y, Peaslee GF, Hickman AB, Dyda F. 2021. The large bat Helitron DNA transposase forms a compact monomeric assembly that buries and protects its covalently bound S'-transposon end. *Mol Cell*. 81(20):4271–4286.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol.* 35(6):1547–1549.
- Lefort V, Longueville JE, Gascuel O. 2017. SMS: smart model selection in PhyML. Mol Biol Evol. 34(9):2422–2424. Available from: http://www. atgc-montpellier.fr/phyml/. Accessed November 16, 2021.
- Li J-H, Lin W-X, Zhang B, Nong D-G, Ju H-P, Ma J-B, Xu C-H, Ye F-F, Xi XG, Li M, et al. 2016. Pif1 is a force-regulated helicase. *Nucleic Acids Res.* 44(9):4330–4339.
- Li HT, Yi TS, Gao LM, Ma PF, Zhang T, Yang JB, Gitzendanner MA, Fritsch PW, Cai J, Luo Y, et al. 2019. Origin of angiosperms and the puzzle of the Jurassic gap. *Nat Plants* 5(5):461–470.
- Liu B, Wang J, Yaffe N, Lindsay ME, Zhao Z, Zick A, Shlomai J, Englund PT. 2009. Trypanosomes have six mitochondrial DNA helicases with one controlling kinetoplast maxicircle replication. *Mol Cell* 35(4):490–501.
- Lu S, Wang J, Chitsaz F, Derbyshire MK, Geer RC, Gonzales NR, Gwadz M, Hurwitz DI, Marchler GH, Song JS, et al. 2020. CDD/SPARCLE: the conserved domain database in 2020. Nucleic Acids Res. 48(D1):D265–D268. Available from: https://www.ncbinlm.nih.gov/ Structure/cdd/. Accessed November 16, 2021.
- Martijn J, Vosseberg J, Guy L, Offre P, Ettema TJ. 2018. Deep mitochondrial origin outside the sampled alphaproteobacteria. *Nature* 557(7703):101–105.
- Muellner J, Schmidt KH. 2020. Yeast genome maintenance by the multifunctional PIF1 DNA helicase family. *Genes* 11(2):224.
- Pérez-Mendoza D, Lucas M, Munoz S, Herrera-Cervera JA, Olivares J, de la Cruz F, Sanjuán J. 2006. The relaxase of the *Rhizobium etli* symbiotic plasmid shows nic site cis-acting preference. J Bacteriol. 188(21):7488–7499.
- Pike JE, Burgers PM, Campbell JL, Bambara RA. 2009. Pif1 helicase lengthens some Okazaki fragment flaps necessitating Dna2 nuclease/helicase action in the two-nuclease processing pathway. J Biol Chem. 284(37):25170-25180.
- Poulter RT, Goodwin TJ, Butler MI. 2003. Vertebrate helentrons and other novel Helitrons. *Gene.* 313:201–212.
- R Core Team. 2020. R: A language and environment for statistical computing. Vienna (Austria): R Foundation for Statistical Computing, Available from: https://www.R-project.org/. Accessed November 16, 2021.
- Ramanagoudr-Bhojappa R, Chib S, Byrd AK, Aarattuthodiyil S, Pandey M, Patel SS, Raney KD. 2013. Yeast Pif1 helicase exhibits a one-basepair stepping mechanism for unwinding duplex DNA. J Biol Chem. 288(22):16185–16195.
- Roger AJ, Muñoz-Gómez SA, Kamikawa R. 2017. The origin and diversification of mitochondria. Curr Biol. 27(21):R1177–R1192.
- RStudio Team. 2020. RStudio: integrated development for R. RStudio. Boston: PBC. Available from: http://www.rstudio.com/. Accessed November 16, 2021.
- Sayers EW, Cavanaugh M, Clark K, Ostell J, Pruitt KD, Karsch-Mizrachi I. 2019. GenBank. Nucleic Acids Res. 47(D1):D94–D99. Available from: https://www.ncbi.nlm.nihgov/genbank/. Accessed November 16, 2021.
- Sela I, Wolf YI, Koonin EV. 2016. Theory of prokaryotic genome evolution. Proc Natl Acad Sci U S A. 113(41):11399–11407.
- Stöver BC, Müller KF. 2010. TreeGraph 2: combining and visualizing evidence from different phylogenetic analyses. BMC Bioinf. 11:7.
- Thomas J, Pritham EJ. 2015. Helitrons, the eukaryotic rolling-circle transposable elements. Microbiol Spectr. 3(4):MDNA3-0049-2014.
- Thomas J, Vadnagara K, Pritham EJ. 2014. DINE-1, the highest copy number repeats in *Drosophila melanogaster* are non-autonomous endonuclease-encoding rolling-circle transposable elements (Helentrons). *Mob Dna*. 5:18.

MBE

- Van Etten J, Bhattacharya D. 2020. Horizontal gene transfer in eukaryotes: not if, but how much? Trends Genet. 36(12):915-925.
- Wang M, Caetano-Anollés G. 2009. The evolutionary mechanics of domain organization in proteomes and the rise of modularity in the protein world. *Structure* 17(1):66–78.
- Wawrzyniak P, Płucienniczak G, Bartosik D. 2017. The different faces of rolling-circle replication and its multifunctional initiator proteins. *Front Microbiol.* 8:2353.
- Wilson MA, Kwon Y, Xu Y, Chung WH, Chi P, Niu H, Mayle R, Chen X, Malkova A, Sung P, et al. 2013. Pif1 helicase and Pol δ promote

recombination-coupled DNA synthesis via bubble migration. *Nature* 502(7471):393–396.

- Wolf YI, Makarova KS, Lobkovsky AE, Koonin EV. 2016. Two fundamentally different classes of microbial genes. *Nat Microbiol.* 2:16208. Xiong W, He L, Lai J, Dooner HK, Du C. 2014. HelitronScanner uncovers a
- Xiong W, He L, Lai J, Dooner HK, Du C. 2014. HelitronScanner uncovers a large overlooked cache of Helitron transposons in many plant genomes. *Proc Natl Acad Sci U S A*. 111(28):10263–10268.
- Yang L, Bennetzen JL. 2009. Structure-based discovery and description of plant and animal Helitrons. *Proc Natl Acad Sci U S A*. 106(31):12832–12837.

Downloaded from https://academic.oup.com/mbe/article/39/1/msab334/6440065 by guest on 27 January 2022

5. CAPÍTULO 3

Multiple horizontal transfers of a *Helitron* transposon associated with a Bracovirus

Pedro Heringer and Gustavo C. S. Kuhn

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, CEP 31270-901, Brazil.

Multiple horizontal transfers of a *Helitron* transposon associated with a Bracovirus

Pedro Heringer and Gustavo C. S. Kuhn*

Departamento de Genética, Ecologia e Evolução, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte, MG, CEP 31270-901, Brazil.

* Corresponding author: Gustavo C. S. Kuhn, gcskuhn@ufmg.br

Abstract

In a previous study we found that a *Helitron* transposon became integrated as a segment in the genome of a symbiotic *Cotesia vestalis* bracovirus (CvBV) from the parasitoid wasp *C. vestalis*. We presented evidence that this *Helitron*, named Hel_c35, initially invaded the *C. vestalis* genome through a horizontal transfer (HT) event from a dipteran species and was later transferred horizontally from *C. vestalis* to a lepidopteran species. We have also anticipated that, as more species would have their genomes sequenced, more HT events involving Hel_c35 might be detected. Here, we investigated the evolution of Hel_c35 in arthropods using a more updated data set to reassess our previous findings. Most species (95%) in the present analysis had their genomes sequenced only after our initial study was published, thus representing new descriptions of taxa harboring Hel_c35. Our results expand considerably the number of putative HTs involving Hel_c35 and suggest that several recent HTs took place in Europe, probably from *C. vestalis* to other insects. We argue that many of these HT events were likely favored by the behavior of this wasp and the stability conferred to Hel_c35 DNA circles by CvBV particles.

Introduction

Horizontal Transfer (HT) events are defined as the exchange of DNA segments between organisms without the involvement of vertical inheritance (Wallau et al. 2018, Van Etten and Bhattacharya 2020). Although HTs are major drivers of evolutionary change in prokaryotes, they are considerably less frequent in eukaryotes, especially in multicellular organisms (Husnik and McCutcheon 2018, Van Etten and Bhattacharya 2020).

In contrast to most genomic components, transposons are DNA segments capable of moving from a locus to another and, as a consequence, they can be found in multiple copies on most eukaryotic genomes, thus being one of the genetic entities most likely to be involved in successful HTs among eukaryotes. Indeed, as the number of eukaryotic sequenced genomes has increased considerably in the last few decades, the number of described examples of horizontal transposon transfers (HTTs) between eukaryotes has also increased, as well as the availability of new bioinformatic methods to detect those events (Schaack et al 2010, Wallau et al. 2018).

We have previously described a *Helitron* transposon from the parasitoid wasp *Cotesia vestalis*, which was found to represent one of the circular segments of the symbiotic virus *C*. *vestalis* bracovirus (CvBV) (Heringer et al. 2017). This *Helitron* was named Hel_c35, as it was first characterized from the CvBV segment 35 (HQ009558.1). The Hel_c35 has 5,294 bp and appears to be autonomous, containing a 4,538 bp gene encoding its transposase (AEE09607.1) consisting of 1,384 amino acids. In the same work, we showed that, not only this CvBV *Helitron* originated after a HTT event (from a *Drosophila* species to *C. vestalis*), but also that this transposon was later transferred horizontally from *C. vestalis* to the domestic silk moth (*Bombyx mori*). Those HTTs were probably facilitated by the close interactions between *C. vestalis* and its potential hosts, which are mediated by CvBV and a fundamental part of this wasp's life cycle. However, as we anticipated in our study, any HT analysis is subject to a different interpretation in the future as more species with sequenced genomes become available (Heringer et al. 2017).

Here, we reassessed our earlier propositions using an updated data set that includes genomes sequenced more recently, providing both a larger and more diverse sample of species. Our results reveal that Hel_c35 elements can be found in a considerably wider range of arthropod species from different orders than it was previously suggested. Likewise, our analysis indicates that presence of Hel_c35 sequences in a large number of species are most likely the result of HT events. In particular, the investigation of sequences more similar to Hel_c35 elements from *C. vestalis* suggests that several recent putative HTs took place in Europe and were probably facilitated by the parasitoid behavior of this wasp, together with the association between Hel_c35 and CvBV.

Results and Discussion

We Blastn searched sequences similar to Hel_c35 (> 80% identity covering > 70% of the query) in all arthropod genomes available on GenBank (Sayers et al. 2019) using the complete CvBV *Helitron* sequence as a query. A total of 285 sequences from 117 species were retrieved for further analyses (Table S1). Although the vast majority of taxa consisted of Lepidoptera species, several different insect orders and two spider species were found to harbor Hel_c35 sequences.

After aligning all the retrieved Hel_c35 sequences, we conducted a phylogenetic analysis using the Maximum Likelihood method. The resulting phylogeny shows that Hel_c35 sequences from specific taxa (insect order or Lepidoptera superfamily) are mostly scattered across different branches, instead of representing the overall topology expected from their evolutionary relationships (Fig. 1). In addition, although several lepidopterans from the same superfamily grouped together, many of those clades contain species from distinct families. At the same time, taxa from the same family were found in separate clades, even though they were grouped with species from the same superfamily (Fig. S1, Table S1).

Despite the diversity and incongruent topology observed in the resulting phylogeny, its Hel_c35 sequences have > 80% sequence identity, what would place its earliest origin at ~ 33 million years ago (MYA), assuming that this transposon evolves neutrally. This diverge time is at least 15 times more recent than the one estimated for the split between arachnids and insects (> 500 MYA) (Kumar et al. 2017) and several times more recent than the estimated time of divergence between most insect orders (Misof et al. 2014). The patchy distribution of taxa, together with the marked deviation between observed and expected divergence times among sequences, strongly indicate that Hel_c35 has been involved in multiple HTT events during its evolution.

Given the large number of sequences included in our phylogeny, we decided to focus our analysis in the main clade containing the CvBV Hel c35 sequence (zoomed in clade on Fig. 1). This well supported clade (SH-aLRT branch support = 0.95) (see also Fig. S1 for support values) contains species from seven insect orders, along with a variety of lepidopteran species from 6 different superfamilies. Similarly to the phylogeny as a whole, most of this clade topology does not reflect the evolutionary relationships between species. Moreover, the estimated evolutionary distances between many sequences in this clade (Table S2) also strongly deviate from their expected divergence times. For example, the cat flea Ctenocephalides felis and Drosophila ficusphila were the two species with the largest number of pairwise differences per site (0.0751) between their Hel c35 sequences. Using a conservative assumption of one generation per year for all species, this clade would have originated ~ 12.5 MYA, which strongly contrasts with the estimated divergence time between most taxa included in this clade. For instance, C. felis and D. ficusphila are estimated to have diverged > 200 MYA, and all Lepidoptera species are estimated to have diverged from Gryllus *bimaculatus* (Orthoptera) > 300 MYA (Kumar et al. 2017). In both examples, if Hel_c35 has been exclusively evolving neutrally and being inherited vertically, no sequence homology would be expected in Hel c35 copies between groups. This contrasts strikingly with the observed sequence nucleotide identity > 92% between all sequences in this clade.



Figure 1. Phylogeny of Hel_c35 sequences. Maximum Likelihood phylogeny including all 285 Hel_c35 sequences retrieved from arthropod genomes is represented on the left. A clade containing sequences closely related to the CvBV Hel_c35 is featured on the right. Lepidoptera species from different superfamilies are represented by different colors. Non-lepidopteran arthropods are represented in black. Branches with < 0.7 SH-aLRT statistical support were collapsed. The same phylogeny with branch supports and all taxa names is shown on Fig. S1.

A deviation from the expected pairwise nucleotide differences per site between species is even more pronounced in the clade comprising taxa with sequences more closely related to the CvBV Hel_c35 (zoomed in clade on Fig. 2). All sequences in this proximal clade have > 99% identity between each other, even though they include species from 3 insect orders that diverged up to > 300 MYA (e.g., Hymenoptera and Lepidoptera) and 6 Lepidoptera superfamilies that diverged up to > 100 MYA (e.g., Bombycoidea and Tortricoidea) (Kumar et al. 2017). Considering the largest value of pairwise nucleotide differences per site among taxa in this clade, which is found between *Apotomis turbidana* and *Habrosyne pyritoides*

(0.009830), its earliest origin would be \sim 1.64 MYA, in contrast to the estimated divergence time for some species included, which are higher by up to two orders of magnitude.

Some of the most conspicuous examples of recent HTTs are shown on the clade containing the Hel_c35 sequences from *C. vestalis* (including CvBV), *Pararge aegeria* and *Pyrgus malvae* (Fig. 1). The phylogenetic relationships between these three species are represented as a polytomy containing sequences with > 99.95% identity, what puts its earliest date of origin at 0.068 MYA (68 thousand years ago). Considering that *P. aegeria* and *P. malvae* diverged > 70 MYA and these two Lepidoptera species have diverged from *C. vestalis* > 300 MYA, these values are at least three orders of magnitude higher than the maximum estimated divergence time for Hel_c35 sequences in this clade.

Even though the phylogenetic topology and level of identity between Hel_c35 sequences strongly suggest the occurrence of multiple HTTs, these events also require some degree of geographic overlap between species to be inferred (Loreto et al. 2008). To verify if the geographical distribution of the analyzed species provides further evidence for HTT events, we represented our phylogeny by color coding the taxa according to the geographical locations where the species were sampled. Sample locations were assigned into one of seven regions defined by their biogeographic realm, bioregions and/or expected migration barriers. Several topological incongruencies consisting of distantly related taxa grouping together on Figure 1 represent species sampled in the same region (Fig. 2 and Fig. S2), indicating that, in some cases, the geographical distribution of species appears to better explain the phylogenetic relationships of their Hel_c35 sequences.

Interestingly, 11 from the 14 species belonging to the CvBV immediate clade correspond to samples from Europe (zoomed in clade on Fig. 2). From those 11 species, nine were derived from the island of Great Britain (Table S1). The remaining two species were collected in Romania (*P. malvae* and *Fabriciana adippe*) but can also be found in Great Britain (Butterfly Conservation 2022a, 2022b). Although the *C. vestalis* samples used in our analysis derive from East Asia (China and South Korea), this wasp species can also be found in several European countries (Furlong et al. 2013), including Great Britain (Broad et al. 2016). Hence, 12 out of 14 species in this clade containing the CvBV Hel_c35 sequence overlap geographically in Great Britain, indicating this island as the most probable region where those HTT events occurred.



Figure 2. Geographical distribution of arthropod species containing Hel_c35. The same phylogeny of Hel_c35 sequences from Fig. 1 is represented, but with colors corresponding to the geographical location where the species were sampled (Table S1). A clade with species containing sequences closely related to CvBV Hel_c35 is featured expanded on the right. The same phylogeny with branch supports and all taxa names is shown on Fig. S2.

Although it is difficult to infer the direction of HTTs, the diversity of Lepidoptera superfamilies at the base of most clades suggests that species in this order are the earliest donors of horizontally transferred Hel_c35 sequences. However, considering the large number of potential HTTs in the presented phylogeny, it is also possible that Lepidoptera species could have received Hel_c35 sequences by secondary HTT events. For example, a HTT from a lepidopteran to a dipteran, which later transferred this transposon to another Lepidoptera species. The diversity and broad distribution of dipterans in the phylogeny (Fig. 1 and Fig. S1) indicate that species from this order were also basal donors of Hel_c35 elements. Nonetheless, because of mechanical and physiological constraints, direct HTs between insects should be

considered rare events. In those cases, it is reasonable to expect the involvement of species like *C. vestalis* as likely HT vectors or intermediates, due to their life history which is thought to facilitate those events (Schaack et al 2010, Wallau et al. 2018). That is particularly relevant for the putative HTTs in the clades more closely related to the CvBV Hel_c35 (Fig. 1). This *Helitron* appears to be autonomous and its copies are likely protected by a viral capsid and envelope when injected every time *C. vestalis* lay eggs in its potential hosts (Heringer et al. 2017). Hence, we suggest a preferred direction for those specific HTT events, which is from the parasitoid to other species.

Considering the topology revealed by our phylogenetic analysis, the geographical distribution of the species and their natural history, we suggest the following hypothesis to explain the putative HTTs involving sequences more closely related to the *C. vestalis* Hel_c35 element. The originally Palearctic/eastern Asian distribution of *C. vestalis* (Hiroyoshi et al. 2017) and several other lepidopterans and dipterans harboring closely related Hel_c35 sequences (Heringer et al. 2017) indicates that *C. vestalis* acquired Hel_c35 by HT from an insect species within those orders, less than 12.5 MYA. In our previous work (Heringer et al. 2017) we suggested a drosophilid as the most probable donor of the *C. vestalis* Hel_c35, given the evidence available at the time. Although our results showing eastern Asian drosophilids near the base of the CvBV Hel_c35 clade provide some support for that hypothesis, we cannot reject that lepidopterans from the same geographical region could also have been potential donors. In any case, after this HTT event, a Hel_c35 sequence became one of CvBV segments, which in turn facilitated other HTTs from *C. vestalis* to multiple species from several insect orders (Fig. 3).

Lepidoptera species are overrepresented in our phylogeny, what could indicate a genome sequencing bias favoring this order. On the other hand, this could likewise be a consequence of lepidopterans being more frequently attacked by parasitoid wasps. This feature might be particularly relevant to explain the putative HTTs indicated in the immediate clade containing the CvBV Hel_c35 sequence (Fig. 1). Despite being considered a specialist parasitoid of the diamondback moth (*Plutella xylostella*), *C. vestalis* is known to attack lepidopterans from at least ten different families within eight superfamilies (Hiroyoshi et al. 2017). In view of the high diversity of lepidopteran larvae that can be targeted by *C. vestalis*, it is reasonable to expect that unspecific attacks to larvae from other insect orders could also occur in some conditions, even if rarely. In fact, the diversity of insect orders found in the main clade containing *C. vestalis*/CvBV in itself might be considered as evidence for the occurrence of those unspecific attacks. As we previously suggested (Heringer et al. 2017), the detection of HTs involving parasitoid wasps and species outside the known range of hosts targeted by



those wasps could be used to indicate potential cryptic interactions to be confirmed in future ecological and behavioral studies.

Figure 3. Hypothesis for HTTs involving Hel_c35 sequences closely related to the one found in CvBV. Arrows represent the probable direction of HTTs and numbers indicate the order which most HTTs events in each geographical region occurred. The earliest event from a Diptera or Lepidoptera species to *C. vestalis* and CvBV (1) was followed by HTTs from CvBV to multiple insects from several orders, initially to species found in Southeast Asia (2) and more recently to species from Europe (3). Although most HTTs in 2 appear to have occurred earlier than those in 3, some European species are interspersed with, or more basal in relation to some Southeast Asian species, indicating that this chronological division is not clear cut.

Overall, the results presented here differ from our previous findings (Heringer et al. 2017) in some important aspects. Firstly, the single best hits from 24 species were retrieved in our earlier work, as opposed to the current analysis, in which 285 sequences from 117 species were included, even though we used a more stringent selection criteria in the latter. For instance, here we considered the same minimum query coverage (>70%) and identity (>80%) as previously, but using the whole CvBV Hel_c35 (5,294 bp) as a reference, as opposed to a region of ~ 838 bp only containing the Rep coding sequence. The sampled species in our former analysis belonged to five insect orders, with one spider species, in contrast to the current sample that comprises 117 species from eight insect orders and two spider families. Therefore, not only the resulting data set presented here is larger, but is also more diverse. It

is also worthwhile mentioning that using this more stringent sequence selection criteria, only five out of 24 species from the previous analysis were included in the current study. Only one of the new species included in the present data set (*Heliconius wallacei*) had its genome sequence already available before the previous study was conducted (November 2015), although we cannot explain the reason for this absence. The remaining 111 new species all had their genome sequences made available only after our previous work (Heringer et al 2017) was submitted (September 2017) and represent 95% of the current data set (Table S1).

The larger number of species in the present analysis revealed a more complex scenario regarding the evolutionary history of Hel_c35 sequences more closely related to the one found in *C. vestalis*. We previously suggested that East/Southeast Asia was probably the geographical region in which the most recent HTTs of Hel_c35 involving *C. vestalis* had occurred (Heringer et al. 2017). Although the evidence provided here still is consistent with a scenario in which the *C. vestalis* Hel_c35 originated from a HTT that probably occurred in East/Southeast Asia < 12.5 MYA, our current results also indicate that this *Helitron* was probably horizontally transferred more recently to multiple insect species in Europe in the last few million years. In spite of those significant differences, our results presented here confirm the previous hypothesis that, as new genome sequencing projects would become available, new HT events would probably be detected, resulting in new interpretations about the evolution of Hel_c35.

Given the large amount of putative HTTs involving *C. vestalis* as a donor of Hel_c35 sequences to other species, and the evidence for CvBV being an important promoter of these events, we consider that future sequencing *C. vestalis* and/or CvBV genomes from different lineages and geographical locations will be essential to confirm our proposed scenario. For instance, we expect that if Hel_c35 copies turn out to be absent in genomes from European lineages of *C. vestalis*, our main hypotheses regarding the direction and geographical location of the most recent HTTs would be refuted, at least partially. Likewise, an absence of Hel_c35 in CvBV genomes from outside East Asia would contradict our suggestion that CvBV has been a major HTT vector of Hel_c35 copies.

Materials and Methods

We Blastn searched all arthropod genomes available (as in October 2021) on the Whole Genome Shotgun (WGS) contigs database from GenBank (Sayers et al. 2019) using the Hel_c35 sequence from CvBV (HQ009558.1) as a query. In order to include only highly similar elements in our analysis, we downloaded all Blast aligned sequences from hits with > 80% sequence identity covering > 70% of the query. Those downloaded hits are sometimes

composed by multiple separate matches which, together, cover > 70% of the query, instead of continuous sequences with the minimum query cover size. Hence, to include only sequences covering > 70% (3,705 bp) of the query, we adapted a Biopython (Cock et al. 2009) script for that purpose and also to edit FASTA sequence descriptions in order to contain only the hit accession number, the sequence match range and the species name (Data S1). The resulting 285 sequences (Data S2) were aligned using the E-INS-i method in the MAFFT online service (Katoh et al. 2019). For the phylogenetic analysis, the best-fit evolutionary model (GTR+G+I) was selected using the Smart Model Selection (SMS) in PhyML (Lefort et al. 2017). The maximum likelihood phylogeny of sequences was inferred using the best topology from NNI and SPR methods, six random plus one parsimony starting trees and 10 substitution rate categories across sites, modelled with estimated gamma-shaped distribution parameter and a proportion of invariant sites. Branch supports were estimated using the approximate likelihood ratio test (aLRT) with the nonparametric Shimodaira-Hasegawa correction (SH-aLRT). The phylogenetic analysis procedures described above were conducted on PhyML 3.1 (Guindon et al. 2010). All branches with < 0.7 SH-aLRT statistical support were collapsed using TreeGraph 2 (Stöver and Müller 2010), with the final tree edited and visualized using FigTree v.1.4.2 (http://tree.bio.ed.ac.uk/software/figtree/; last accessed December 15, 2022). The species taxonomy and sample collection locations were obtained from their corresponding accession on GenBank (Sayers et al. 2019), and additional information about the geographical distribution of organisms included in our analysis was obtained from various Web sources. The average nucleotide differences per site between groups in the main clade containing CvBV Hel c35 (Table S2) was calculated using MEGA X (Kumar et al. 2018), and their divergence time estimated using the equation:

$$T = \frac{K}{2r}$$

in which *T* is the number of generations, *K* is the number of substitutions per site, and *r* is the rate of nucleotide substitution. We considered that *r* is equal to the mutation rate (μ), as expected for neutral mutations (Graur and Li 2000), and a value of μ equal to 3.0 x 10⁻⁹ for insect species (Liu et al. 2017). To obtain a conservative estimation for the maximum time of divergence between sequences we considered one generation per year for all insect species. Hence, in our equation, the value found for *T* is equal to the diverge time between species given in number of years.

References

Broad, G. R., Shaw, M. R., Godfray, H. C. J. (2016). Checklist of British and Irish Hymenoptera - Braconidae. Biodiversity Data Journal, 4:e8151.

Butterfly Conservation. (2022a). Grizzled Skipper, Pyrgus malvae. Available at: https://butterflyconservation.org/butterflies/grizzled-skipper (last accessed January 22, 2022).

Butterfly Conservation. (2022b). High Brown Fritillary, Fabriciana adippe. Available at: https://butterfly-conservation.org/butterflies/high-brown-fritillary (last accessed January 22, 2022).

Cock, P. J., Antao, T., Chang, J. T., Chapman, B. A., Cox, C. J., Dalke, A., ... & De Hoon, M. J. (2009). Biopython: freely available Python tools for computational molecular biology and bioinformatics. Bioinformatics, 25(11), 1422-1423.

Furlong, M. J., Wright, D. J., & Dosdall, L. M. (2013). Diamondback moth ecology and management: problems, progress, and prospects. Annual review of entomology, 58, 517-541.

Graur, D., and W.-H. Li. 2000. Fundamentals of Molecular Evolution, Ed. 2. Sinauer Associates, Sunderland, MA.

Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol. 59(3):307-321.

Heringer, P., Dias, G. B., & Kuhn, G. C. (2017). A horizontally transferred autonomous Helitron became a full polydnavirus segment in Cotesia vestalis. G3: Genes, Genomes, Genetics, 7(12), 3925-3935.

Hiroyoshi, S., Harvey, J. A., Nakamatsu, Y., Nemoto, H., Mitsuhashi, J., Mitsunaga, T., & Tanaka, T. (2017). Potential host range of the larval endoparasitoid Cotesia vestalis (= plutellae) (Hymenoptera: Braconidae). International Journal of Insect Science, 9. https://doi.org/10.1177/1179543317715623

Husnik, F., & McCutcheon, J. P. (2018). Functional horizontal gene transfer from bacteria to eukaryotes. Nature Reviews Microbiology, 16(2), 67-79.

Katoh, K., Rozewicki, J., & Yamada, K. D. (2019). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Briefings in bioinformatics, 20(4), 1160-1166.

Kumar, S., Stecher, G., Suleski, M., & Hedges, S. B. (2017). TimeTree: a resource for timelines, timetrees, and divergence times. Molecular biology and evolution, 34(7), 1812-1819.

Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. Molecular biology and evolution, 35(6):1547-1549.

Lefort V, Longueville JE, Gascuel O. 2017. SMS: smart model selection in PhyML. Mol Biol Evol. 34(9):2422-2424. (Available from: <u>http://www.atgc-montpellier.fr/phyml/</u>).

Liu, H., Jia, Y., Sun, X., Tian, D., Hurst, L. D., & Yang, S. (2017). Direct determination of the mutation rate in the bumblebee reveals evidence for weak recombination-associated mutation and an approximate rate constancy in insects. Molecular biology and evolution, 34(1), 119-130.

Loreto, E. L. S., Carareto, C. M. A., & Capy, P. (2008). Revisiting horizontal transfer of transposable elements in Drosophila. Heredity, 100, 545-554.

Misof, B., Liu, S., Meusemann, K., Peters, R. S., Donath, A., Mayer, C., ... & Zhou, X. (2014). Phylogenomics resolves the timing and pattern of insect evolution. Science, 346(6210), 763-767.

Sayers EW, Cavanaugh M, Clark K, Ostell J, Pruitt KD, Karsch-Mizrachi I. 2019. GenBank. Nucleic Acids Res. 47(D1):D94-D99. Available from: https://www.ncbi.nlm.nih.gov/genbank/. Accessed October 30, 2021.

Schaack, S., Gilbert, C., & Feschotte, C. (2010). Promiscuous DNA: horizontal transfer of transposable elements and why it matters for eukaryotic evolution. Trends in ecology & evolution, 25(9), 537-546.

Stöver BC, Müller KF. 2010. TreeGraph 2: combining and visualizing evidence from different phylogenetic analyses. BMC Bioinf. 11:7.

Van Etten, J., Bhattacharya, D. (2020). Horizontal gene transfer in eukaryotes: not if, but how much?. Trends in Genetics. 36(12):915–925.

Wallau, G. L., Vieira, C., & Loreto, É. L. S. (2018). Genetic exchange in eukaryotes through horizontal transfer: connected by the mobilome. Mobile DNA, 9:6.

6. DISCUSSÃO GERAL

Os resultados apresentados no Capítulo 1 sugerem que, a despeito de os *Helitrons* serem transposons exclusivamente eucarióticos, estes elementos pertencem a uma linhagem filogenética de replicons tipicamente procarióticos. Apesar de os *Helitrons* terem uma origem procariótica, modo de replicação por transposição e possuírem uma proteína com atividade enzimática semelhante às encontradas em transposons procarióticos que utilizam RCT, nossos resultados indicam que *Helitrons* não são parentes próximos destes últimos. Por outro lado, a hipótese de que *Helitrons* seriam descendentes ou mesmo teriam dado origem a vírus eucarióticos do tipo RCR também não é sustentada pelos resultados das nossas análises utilizando o domínio Rep.

Ao contrário, nossos dados indicam que *Helitrons* são parentes são mais proximamente relacionados a plasmídeos e vírus procarióticos, formando com estes um grupo filogenético composto por elementos circulares que se replicam por RCR e possuem duas tirosinas catalíticas no seu domínio Rep. Após sua publicação (Heringer & Kuhn 2018), estes resultados foram corroborados por um estudo independente que analisou as relações evolutivas entre proteínas Rep de elementos procarióticos e eucarióticos (Kazlauskas et al. 2019).

Já os resultados no Capítulo 2 argumentam contra a hipótese de que os *Helitrons* teriam adquirido seu domínio Hel após a captura de uma helicase Pif1 eucariótica. Apesar de helicases Pif1 serem tipicamente codificadas por genomas de eucariotos, esta família de proteínas também é encontrada em diversos genomas de arqueias e bactérias, além de vírus eucarióticos e procarióticos. A distribuição filogenética das proteínas analisadas demonstra que o domínio Hel evoluiu independentemente de da linhagem que deu origem a helicases Pif1 eucarióticas, indicando que *Helitrons* já possuíam uma transposase contendo seus dois domínios antes de invadirem seus primeiros hospedeiros eucariotos.

Sugerimos que *Helitrons* representam um grupo de plasmídeos procarióticos que, após invadirem organismos eucariotos, passaram a se replicar por transposição nos genomas de seus hospedeiros (Fig. 6 do Cap. 2). Esta hipótese se baseia no conjunto de dados revelados no presente trabalho e em outros estudos, sendo estas evidências apresentadas a seguir. Primeiramente, apesar de terem se tornado transposons, *Helitrons* geram intermediários de dsDNA circulares para se mover no genoma (Grabundzija et al. 2018). Além disso, estes elementos possuem em sua transposase um domínio Rep mais proximamente relacionado com proteínas de vírus circulares e plasmídeos (Cap. 1, Kazlauskas et al. 2019).

Por outro lado, o domínio helicase presente em relaxases TraA de plasmídios parece ser filogeneticamente relacionado à família Pif1, que inclui o domínio Hel (Cap. 2). Apesar de remota, esta relação sugere que *Helitrons* poderiam representar parentes distantes de plasmídeos atuais. Mesmo considerando que a similaridade entre a transposase RepHel e a relaxase TraA provavelmente resulta de convergência evolutiva, tal fato ainda indicaria a existência de paralelos entre os processos enzimáticos conduzidos por estas duas proteínas distintas. Recentemente, a estrutura da RepHel associada à extremidade 5' ssDNA do *Helitron* foi resolvida por crio-microscopia eletrônica, revelando que esta transposase apresenta uma estrutura tridimensional notavelmente similar a encontrada na relaxase Tral (Kosek et al. 2021). Assim como a semelhança na sequência de aminoácidos observada para o caso da relaxase TraA, a similaridade estrutural entre Tral e RepHel muito provavelmente resulta de convergência evolutiva pelo fato de ambas as proteínas desempenharem reações catalíticas análogas.

Apesar de possuírem características de plasmídeos, o conjunto de resultados apresentados nos dois primeiros capítulos indicam que cada um dos dois domínios principais da transposase RepHel se assemelha mais a proteínas encontradas em elementos genéticos móveis de grupos distintos. De um lado, o domínio Rep claramente pertence a um grupo de proteínas responsáveis pela replicação de plasmídeos e vírus procarióticos do tipo RCR (Cap. 1); do outro, o domínio Hel representa um dos clados mais basais de helicases Pif1 (Cap. 2). De fato, a divergência dos dois grandes grupos do domínio Hel (*Helitrons* e *Helentrons/Helitron2*) parece ser tão antiga quanto as principais radiações basais de proteínas semelhantes a helicases Pif1. A profundidade desta divergência evolutiva entre domínios Hel de *Helitrons* e *Helentrons/Helitron2* é tão acentuada que domínios Hel sequer formam grupos monofiléticos na nossa análise (Fig. 3 e Fig. S2 do Cap. 2).

Por fim, no Capítulo 3 exemplificamos a capacidade que os *Helitrons* possuem de se propagar horizontalmente cruzando a barreira das espécies, muitas vezes entre organismos de ordens ou mesmo classes diferentes. Em um trabalho anterior (Heringer et al. 2017) havíamos identificado um *Helitron*, denominado Hel_c35, que se tornou um dos segmentos do vírus simbionte *Cotesia vestalis* bracovirus (CvBV) associado à vespa parasitoide *C. vestalis*. Neste último estudo, também havíamos demonstrado que elementos Hel_c35 se encontravam distribuídos de forma desigual em genomas de diversas espécies de insetos em diferentes ordens, além de uma espécie de aracnídeo. Tal distribuição desigual e irregular já indicava que este *Helitron* estaria envolvido em vários eventos de HT. De fato, nossos resultados sugeriam que o próprio elemento Hel_c35 presente em CvBV teria se originado após a HT de um díptero para *C. vestalis*, seguida pela inserção deste *Helitron* no genoma proviral de CvBV. Além disso, nossas análises apontavam para um segundo evento de HT, de *C. vestalis* para a espécie de mariposa *Bombyx mori*.

Os resultados apresentados no Capítulo 3 descrevem a evolução de elementos Hel_c35 e sua distribuição em genomas de artrópodes utilizando uma amostra consideravelmente maior de espécies e análises mais robustas. Além de atualizar nossos achados anteriores (Heringer et al. 2017) ao revelar uma quantidade e diversidade consideravelmente maior de espécies com elementos Hel_c35, nossos resultados sugerem que esta família de *Helitrons* possivelmente está envolvida em dezenas de eventos de HT. Várias destas HTs estão associadas a espécies que contém sequências mais similares ao elemento Hel_c35 encontrado em *C. vestalis*, provavelmente foram transferidas horizontalmente desta vespa parasitoide para outras espécies de insetos e facilitada pela presença do *Helitron* Hel_c35 em partículas virais de CvBV.

7. CONCLUSÕES

Desde que os *Helitrons* foram descritos pela primeira vez em 2001, estes elementos têm se revelado cada vez mais como componentes genômicos importantes e versáteis em diversos grupos de organismos eucariotos. Algumas das características mais bem estabelecidas sobre os *Helitrons* nas últimas duas décadas dizem respeito a sua capacidade de ocupar frações consideráveis dos seus genomas hospedeiros, capturar, mobilizar e duplicar fragmentos cromossômicos. Apesar disso, informações sobre a sua origem e mecanismo de transposição permaneceram obscuras até recentemente. O objetivo central deste trabalho foi o de elucidar a origem e relações evolutivas destes elementos através do estudo de sua estrutura codificante, composta por dois domínios principais. Nossos resultados indicam que *Helitrons* representam transposons descendentes de plasmídeos procarióticos que invadiram o genoma dos seus primeiros hospedeiros eucarióticos em um período próximo à origem deste domínio da vida. Apesar do domínio catalítico central da sua transposase RepHel se assemelhar mais a proteínas encontradas em um grupo de plasmídeos e vírus bacterianos, *Helitrons* diferem destes últimos por codificarem um domínio helicase em sua transposase.

Em conjunto com dados revelados em outros estudos, nossos resultados sugerem que este domínio helicase não representa uma aquisição evolutiva posterior à invasão dos *Helitrons* em genomas eucarióticos. Ao contrário, a estrutura composta por dois domínios principais na transposase RepHel parece anteceder a origem dos *Helitrons* em eucariotos e ser indispensável para a transposição destes elementos. De fato, a similaridade estrutural entre a transposase RepHel e relaxases encontradas em plasmídeos indica que o domínio Hel desempenha uma função complexa que vai além da simples atividade típica de uma helicase. Neste cenário, os domínios Rep e Hel desempenhariam funções enzimáticas essenciais, complementares e necessariamente concatenadas nas principais etapas do processo de transposição dos *Helitrons*.

Para além dos aspectos fundamentais sobre a origem e mecanismo de transposição destes elementos, nosso estudo de uma família de *Helitrons* encontrada em artrópodes ilustra como a evolução destes transposons em genomas hospedeiros pode ser altamente complexa. Tal complexidade se dá pela capacidade dos *Helitrons* de invadir novas espécies por transferência horizontal, sendo que análises filogenéticas de suas sequências comumente resultam em topologias incongruentes com as relações evolutivas de suas espécies hospedeiras. A evolução desta família de *Helitrons* analisada no nosso último capítulo é particularmente notável não só por incluir múltiplos eventos de transferência horizontal entre

diferentes ordens de artrópodes, mas também pela associação entre um elemento desta família com o vírus simbiótico de uma vespa parasitóide.

Os aspectos revelados sobre os *Helitrons* neste trabalho, e em outros estudos recentes, sobre a sua origem, evolução, mecanismo de transposição e estrutura da sua transposase, abrem caminho para futuras investigações mais profundas sobre cada um destes temas. No campo das análises in silico, o aumento no número de espécies com genomas sequenciados poderá contribuir com cenários mais completos sobre a origem dos *Helitrons*, seja revelando variantes estruturalmente mais semelhantes à sua forma ancestral ou replicons evolutivamente mais próximos dos *Helitrons*. Já análises in vitro poderão confirmar se as similaridades estruturais entre transposases RepHel e relaxases de fato se traduzem em semelhanças funcionais. Por fim, a compreensão mais detalhada da estrutura e processos enzimáticos conduzidos por esta transposase única em genomas eucariotos cria novas possibilidades na investigação de ferramentas de engenharia genética.

8. REFERÊNCIAS

Bellas, C. M., & Sommaruga, R. (2021). Polinton-like viruses are abundant in aquatic ecosystems. Microbiome, 9:13.

Bochman, M. L., Sabouri, N., & Zakian, V. A. (2010). Unwinding the functions of the Pif1 family helicases. DNA repair, 9(3), 237-249.

Boule, J. B., & Zakian, V. A. (2006). Roles of Pif1-like helicases in the maintenance of genomic stability. Nucleic acids research, 34(15), 4147-4153.

Bourque, G., Burns, K. H., Gehring, M., Gorbunova, V., Seluanov, A., Hammell, M., ... & Feschotte, C. (2018). Ten things you should know about transposable elements. Genome biology, 19:199.

Chandler, M., De La Cruz, F., Dyda, F., Hickman, A. B., Moncalian, G., & Ton-Hoang, B. (2013). Breaking and joining single-stranded DNA: the HUH endonuclease superfamily. Nature Reviews Microbiology, 11(8), 525-538.

Chellapan, B. V., van Dam, P., Rep, M., Cornelissen, B. J., & Fokkens, L. (2016). Non-canonical Helitrons in Fusarium oxysporum. Mobile DNA, 7:27.

Dias, G. B., Heringer, P., & Kuhn, G. C. (2016). Helitrons in Drosophila: Chromatin modulation and tandem insertions. Mobile genetic elements, 6(2), e1154638.

Du, C., Fefelova, N., Caronna, J., He, L., & Dooner, H. K. (2009). The polychromatic Helitron landscape of the maize genome. Proceedings of the National Academy of Sciences, 106(47), 19916-19921.

Feschotte, C., & Wessler, S. R. (2001). Treasures in the attic: rolling circle transposons discovered in eukaryotic genomes. Proceedings of the National Academy of Sciences, 98(16), 8923-8924.

Grabundzija, I., Messing, S. A., Thomas, J., Cosby, R. L., Bilic, I., Miskey, C., ... & Ivics, Z. (2016). A Helitron transposon reconstructed from bats reveals a novel mechanism of genome shuffling in eukaryotes. Nature communications, 7:10716.

Grabundzija, I., Hickman, A. B., & Dyda, F. (2018). Helraiser intermediates provide insight into the mechanism of eukaryotic replicative transposition. Nature communications, 9:1278.

Heringer, P., Dias, G. B., & Kuhn, G. C. (2017). A horizontally transferred autonomous Helitron became a full polydnavirus segment in Cotesia vestalis. G3: Genes, Genomes, Genetics, 7(12), 3925-3935.

Heringer, P., & Kuhn, G. (2018). Exploring the remote ties between helitron transposases and other rolling-circle replication proteins. International journal of molecular sciences, 19(10), 3079.

Kapitonov, V. V., & Jurka, J. (2001). Rolling-circle transposons in eukaryotes. Proceedings of the National Academy of Sciences, 98(15), 8714-8719.

Kapitonov, V. V., & Jurka, J. (2007). Helitrons on a roll: eukaryotic rolling-circle transposons. TRENDS in Genetics, 23(10), 521-529.

Kazlauskas, D., Varsani, A., Koonin, E. V., & Krupovic, M. (2019). Multiple origins of prokaryotic and eukaryotic single-stranded DNA viruses from bacterial and archaeal plasmids. Nature communications, 10:3425

Koonin, E. V., & Dolja, V. V. (2014). Virus world as an evolutionary network of viruses and capsidless selfish elements. Microbiology and Molecular Biology Reviews, 78(2), 278-303.

Koonin, E. V., & Krupovic, M. (2017). Polintons, virophages and transpovirons: a tangled web linking viruses, transposons and immunity. Current opinion in virology, 25, 7-15.

Kosek, D., Grabundzija, I., Lei, H., Bilic, I., Wang, H., Jin, Y., ... & Dyda, F. (2021). The large bat Helitron DNA transposase forms a compact monomeric assembly that buries and protects its covalently bound 5'-transposon end. Molecular Cell, 81(20), 4271-4286.

Krupovic, M. (2013). Networks of evolutionary interactions underlying the polyphyletic origin of ssDNA viruses. Current opinion in virology, 3(5), 578-586.

Krupovic, M., Bamford, D. H., & Koonin, E. V. (2014). Conservation of major and minor jellyroll capsid proteins in Polinton (Maverick) transposons suggests that they are bona fide viruses. Biology direct, 9:6.

Krupovic, M., & Koonin, E. V. (2015). Polintons: a hotbed of eukaryotic virus, transposon and plasmid evolution. Nature Reviews Microbiology, 13, 105-115.

Muellner, J., & Schmidt, K. H. (2020). Yeast genome maintenance by the multifunctional PIF1 DNA helicase family. Genes, 11(2), 224.

Pritham, E. J., & Feschotte, C. (2007). Massive amplification of rolling-circle transposons in the lineage of the bat Myotis lucifugus. Proceedings of the National Academy of Sciences, 104(6), 1895-1900.

Schaack, S., Gilbert, C., & Feschotte, C. (2010). Promiscuous DNA: horizontal transfer of transposable elements and why it matters for eukaryotic evolution. Trends in ecology & evolution, 25(9), 537-546.

Thomas, J., & Pritham, E. J. (2015). Helitrons, the eukaryotic rolling-circle transposable elements. Microbiology spectrum, 3(4):MDNA3-0049-2014.

Thomas, J., Vadnagara, K., & Pritham, E. J. (2014). DINE-1, the highest copy number repeats in Drosophila melanogaster are non-autonomous endonuclease-encoding rolling-circle transposable elements (Helentrons). Mobile DNA, 5:18.

Van Etten, J., Bhattacharya, D. (2020). Horizontal gene transfer in eukaryotes: not if, but how much?. Trends in Genetics. 36(12):915–925.

Wallau, G. L., Vieira, C., & Loreto, É. L. S. (2018). Genetic exchange in eukaryotes through horizontal transfer: connected by the mobilome. Mobile DNA, 9:6.

Wawrzyniak, P., Płucienniczak, G., & Bartosik, D. (2017). The different faces of rolling-circle replication and its multifunctional initiator proteins. Frontiers in microbiology, 8, 2353.

Wells, J. N., & Feschotte, C. (2020). A field guide to eukaryotic transposable elements. Annual review of genetics, 54, 539-561.

9. ANEXOS

9.1 Material suplementar do Capítulo 1

Supplementary Material

Group	Sequence ID	Taxon name	Family/Group ^a	# of tyr ^b	Accession
Eukaryotic viruses					
0	MSV	Maize streak virus	Geminiviridae	1	AAF97764.1
	WDV	Wheat dwarf virus	Geminiviridae	1	CAA57625.1
	BMCTV	Beet mild curly top virus	Geminiviridae	1	AAC54875.1
	TYLCSV	Tomato yellow leaf curl Sardinia virus	Geminiviridae	1	CAA43466.1
	CLCGV	Cotton leaf curl Gezira virus	Geminiviridae	1	AAF97439.1
	SsHADV	Sclerotinia sclerotiorum hypovirulence associated DNA virus 1	Genomoviridae	1	YP_003104796.1
	PFFFGmV	Pacific flying fox faeces associated gemycircularvirus 12	Genomoviridae	1	AMH87729.1
	HPAGmV	Human plasma-associated gemycircularvirus	Genomoviridae	1	YP_009181996.1
	BBTV	Banana bunchy top virus	Nanoviridae	1	NP_604483.1
	FBNS	Faba bean necrotic stunt virus	Nanoviridae	1	YP_003104737.1
	SCSV	Subterranean clover stunt virus	Nanoviridae	1	Q9ICP7.1
	FBNY	Faba bean necrotic yellows C11 alphasatellite	Nanovirus-associated alphasatellite	1	NP_619565.1
	MVDC2	Milk vetch dwarf C2 alphasatellite	Nanovirus-associated alphasatellite	1	NP_619760.1
	PCV	Porcine circovirus 1	Circoviridae	1	NP_065678.1
	SGCV	Silurus glanis circovirus	Circoviridae	1	YP_009091696.1
	ZFCV	Zebra finch circovirus	Circoviridae	1	YP_009134739.1
	HSCycl	Cyclovirus PK5510 (H. sapiens)	Circoviridae	1	ADD62457.1
	DACycl	Dragonfly associated cyclovirus 1	Circoviridae	1	YP_009021893.1
	CACycl	Chicken associated cyclovirus 1 (NGchicken8)	Circoviridae	1	ADU77011.1
	DCircV	Diporeia sp. associated circular vírus	Unclassified	1	AGG39813.1
	SARCircV	Circovirus-like genome SAR-A	Unclassified	1	ACQ78172.2
	MpaCircV1	McMurdo Ice Shelf pond-associated circular DNA virus 1	Unclassified	1	AIF71501.1
	MpaCircV2	McMurdo Ice Shelf pond-associated circular DNA virus 2	Unclassified	1	AIF71504.1
	MpaCircV3	McMurdo Ice Shelf pond-associated circular DNA virus 3	Unclassified	1	AIF71507.1
	MpaCircV4	McMurdo Ice Shelf pond-associated circular DNA virus 4	Unclassified	1	AIF71509.1
	MpaCircV5	McMurdo Ice Shelf pond-associated circular DNA virus 5	Unclassified	1	AIF71512.1
	RsaCircV	Rodent stool-associated circular genome virus	Unclassified	1	AEM05803.1
	BcCircV	Bat circovirus ZS/China/2011	Unclassified	1	AEL87784.1
	CsalDNAV	Chaetoceros salsugineum DNA virus	d Bacilladnaviridae	1	YP_473359.1
	AcrBV1	Amphibola crenata associated bacilladnavirus 1	Bacilladnaviridae	1	YP_009345107.1
	AHEaBV	Avon-Heathcote estuary associated bacilladnavirus	Bacilladnaviridae	1	YP_009345097.1
	AAV2	Adeno-associated virus 2	Parvoviridae	2	YP_680422.1
	AAV5	Adeno-associated virus 5	Parvoviridae	2	YP_068408.1
	SLP	Slow loris parvovirus 1	Parvoviridae	2	YP_009111339.1
Bacterial viruses					
	phiX174	Enterobacteria phage phiX174	Microviridae	2	NP_040703.1
	phageNC3	Enterobacteria phage NC3	Microviridae	2	AAZ49040.1
	ERBP1	Eel River basin pequenovirus	Microviridae	2	YP_009126954.1
	P2	Escherichia virus P2	Myoviridae	2	NP_046795.1
	Sphage_RE2010	Salmonella phage RE-2010	Myoviridae	2	YP_007003504.1
	phiE122	Burkholderia virus phiE122	Myoviridae	2	YP_001111165.1
	phi_Lf	Xanthomonas phage Lf	Inoviridae	2	AAC54630.1
	SVTS2	Spiroplasma phage SVTS2	Inoviridae	2	AAF18311.2
	Rhizob_R404	Rhizobacter sp. Root404 (Inovirus Gp2 family protein)	Inoviridae	2	WP_056466193.1
	RSIBR1	Ralstonia virus RSIBR1	Inoviridae	2	ATW64834.1
	GkshoV_Hs	Gokushovirus WZ-2015a (H.sapiens)	Microviridae	2	ALS03579.1
	GkshoV_Bird	Gokushovirus WZ-2015a (Bird)	Microviridae	2	ALS03530.1
	GkshoV Marine	Marine gokushovirus	Microviridae	2	YP 008798246.1

Supplementary Table S1. Taxa information

		HRPV1	Halorubrum pleomorphic virus 1	Pleolipoviridae	2	YP_002791886.1
		HRPV2	Halorubrum pleomorphic virus 2	Pleolipoviridae	2	YP_005454258.1
		H_rubripr	Haloarcula rubripromontorii	Haloarculaceae	2	KOX95265.1
		SNJ1	Natrinema virus SNJ1	Sphaerolipoviridae	1	NC_003158.1
		H_inordinatus	Halopelagius inordinatus	Haloferacaceae	1	WP_092894117.1
		H_thailandensis	Halococcus thailandensis JCM 13552	Halococcaceae	1	EMA56448.1
		CN_piranensis	Candidatus Nitrosopumilus piranensis	Nitrosopumilaceae	1	AJM92193.1
		Therm_BRNA1	Thermoplasmatales archaeon BRNA1	unclassified Thermoplasmatales	1	WP_015491922.1
		Thaum_SCGC	Marine Group I thaumarchaeote SCGC AAA799-P11	unclassified Thaumarchaeota	1	WP_048071526.1
Prokaryot	ic TEs					_
		1591	Insertion sequence IS91 (Escherichia coli)	IS91 Group	2	S23782
		IS801	Insertion sequence IS801 (Pseudomonas savastanoi)	IS91 Group	2	P24607.1
		IS1294	Insertion sequence IS1294 (Escherichia coli)	IS91 Group	2	CAA07835.1
		ISCR1	Insertion sequence ISCR1 (Citrobacter freundii)	ISCR Group	1	AFL38296.1
		ISCR2	Insertion sequence ISCR2 (Klebsiella pneumoniae)	ISCR Group	1	SBN37579.1
		ISCR3	Insertion sequence ISCR3 (Pseudomonas aeruginosa)	ISCR Group	1	ATE47644.1
		15608	Insertion sequence IS608 (Helicobacter pylori)	IS200/IS605 Family	1	2A6M A
		Rhiz NXC24	IS200/IS605 insertion sequence (Rhizobium sp. NXC24)	IS200/IS605 Family	1	AVA22184.1
		ISDra2	Insertion sequence ISDra2 (Deinococcus radiodurans)	IS200/IS605 Family	1	WP 010887312.1
Plasmids						-
Euka	ryotic	pPpulchr	Pyropia pulchra (red algae) plasmid	Gemini_AL1	1	AAF36424.1
Bact	erial	pEcOYNIM	Onion yellows phytoplasma EcOYNIM_2000	Gemini_AL1	1	YP_006959597.1
		pPASb11	Candidatus Phytoplasma australiense plasmid pPASb11	Gemini AL1	1	- YP 001965310.1
		pPAPh2	Candidatus Phytoplasma australiense plasmid pPAPh2	Gemini AL1	1	YP 001965305.1
		pPaWBNy	Paulownia witches'-broom phytoplasma plasmid pPaWBNv-1	Gemini AL1	1	YP 001708784.1
		p4M	Bifidobacterium pseudocatenulatum plasmid p4M	Viral Rep	1	NP 613078.1
		pFTB14	Bacillus amyloliquefaciens plasmid pFTB14	Rep 1	1	P13963.1
		pUB110	Staphylococcus aureus plasmid pUB110	Rep 1	1	AAA88362.1
		pBC1	Bacillus coagulans plasmid pBC1	Rep 1	1	AAA98048.1
		pKYM	Shigella sonnei nlasmid nKYM	Rep 1	1	AAA98159 1
		p5K89	Stanbylococcus aureus plasmid nSK89	Rep_1	1	AAB02112.1
		pNost	Nostoc sp. plasmid ('nNost')	Rep_1	1	AAA25513.1
		nTD1	Trenonema denticola plasmid nTD1	Rep_1	1	AAA98363 1
		DAYWB	Aster vellows witches'-broom phytoplasma AYWB plasmid pAYWB-II	Rep_1	1	ABC65794 1
		pOYM	Onion vellows nhvtoplasma plasmid pOYM	Rep_2	1	YP_002600752.1
		pCPa	Candidatus Phytoplasma australiense plasmid pCPa	Rep_2	1	YP_001966814.1
		plm	Leuconostor mesenteroides plasmid replication protein	Rep_2	1	WP 002815993 1
		nla	Lactobacillus acidonbilus plasmid replication protein	Rep_2	1	WP_003549058.1
		p04504	Lactoroccus lactis plasmid pQ4504	Rep_2	1	AFU41945 1
		nSAP110B	Stanbylococcus enidermidis plasmid SAP110B	Rep_2	1	YP 006939186 1
		pMV158	Streptococcus agalactiae plasmid pMV158	Rep_2	1	VP_001586272.1
		pE194	Staphylococcus agriactiae plasmid pE194	Rep_2	1	P03858 2
		p2194	Myconlacma mycoides pADP201	Rep_2	1	NR 040420 2
		pAD6201	lactococcus lactis plasmid pW/01	Rep_2	1	NP 053450 1
		pPhacel	Dhare-placmid hubrid Dhard	Phage GPA	2	D10071 1
		prilasyi	nage-plasmu nyonu masyi		2	PAA07700 1
		plinnamed2	Eucobastarium puelostum suben polimeentum eleveid "	PHA00220	2	042405 1
		ponnamedz	rusobacterium nucleatum subsp. polymorphum plasmid "unnamed2"		2	ALQ43495.1
		pols	Leptolyngbya boryana plasmid pGL3	Unclassified	2	AAA25610.1

Unclassified

DUF1424

DUF1424

DUF1424

DUF1424

2

2

2

2

2

BAA34784.1

S06780

P17565.1

YP_232880.1

AKN10606.1

Archaeal viruses

pSA1

pHGN1

pGRB1

pHF2

pZMX201

Archaeal

Streptomyces cyaneus plasmid pSA1.1

Halobacterium salinarum plasmid pGRB1

Natrinema sp. CX2021 plasmid pZMX201

Halobacterium sp. plasmid pHGN1

Haloferax sp. Q22 plasmid pHF2
	pHK2	Haloferax lucentense DSM 14919 plasmid pHK2	DUF1424	2	YP_006961960.1
	pNB101	Natronobacterium sp. AS-7091 plasmid pNB101	DUF1424	2	NP_942603.1
	pML	Methanohalophilus mahii plasmid pML	DUF1424	2	NP_976268.1
	pTP2	Thermococcus prieurii plasmid pTP2	PHA00330	2	YP_007974244.1
Helitrons					
	Helen_A_aeg	Helitron-2_Aae (Aedes aegypti)	Helentron	2	Helitron-2_Aae ^g
	Helen_D_rer	Helitron-2_DR (Danio rerio)	Helentron	2	Helitron-2_DR
	Helen_D_kik	Helitron-1_DK (Drosophila kikkawai)	Helentron	2	Helitron-1_DK
	Helen_N_vec	Helitron-1_NV (Nematostella vectensis)	Helentron	2	Helitron-1_NV
	Helen_M_cir	Helitron-like sequence (Mucor circinelloides)	Helentron	2	EPB86818.1
	Helen_C_gig	Helitron-10_Cgi (Crassostrea gigas)	Helentron	2	Helitron-10_CGi ^g
	Hel2_F_oxy	FoHeli1 (Fusarium oxysporum)	Helitron2	2	FoHeli1 ^g
	Hel_A_tha	HELITRON1 (Arabidopsis thaliana)	Helitron	2	AAD15468.1
	Hel_c35	Hel_c35 (Cotesia vestalis bracovirus)	Helitron	2	AEE09607.1
	Hel_M_luc	HELIBAT1 (Myotis lucifugus)	Helitron	2	HELIBAT1
	Hel_A_nid	Helitron-1_AN (Aspergillus nidulans)	Helitron	2	XP_662882.1
	Hel_C_ele	HELITRON1_CE (Caenorhabditis elegans)	Helitron	2	NP_493834.1
	Hel_A_gam	HELITRON1_AG (Anopheles gambiae)	Helitron	2	HELITRON1_AG

Notes:

^a Plasmids were classified by their RCRE protein family. Helitrons were assigned to their structural variant according to Thomas and Pritham (2015). ^b Number of tyrosines in the catalytic core. The colors indicate the tyrosine group (Y1 = green, Y2 = red, Yx = blue), as shown in figures 2 and 3C. ^c Sequences representing unclassified viruses were sampled from Zawar-Reza et al. (2014). ^d Family proposed by Kazlauskas et al. (2017). ^e Viral sequence integrated in the genome of indicated taxon. ^f Translated ORF was obtained from nucleotide sequence, according to Wang et al. (2016). ^g Sequences retrieved from Repbase (Bao et al. 2015).



Supplementary Figure S1. Phylogenetic analysis of RCRE domain sequences. Same phylogeny as in Figure 2, with branch support numerical values displayed. Only values above 50% are shown.



Supplementary Figure S2. Phylogenetic and NMDS analysis of helicase sequences. (A) Phylogeny of helicase domain sequences inferred by the Neighbor Joining method (Poisson correction). (B) Phylogeny of helicase domain sequences inferred by the Maximum Likelihood method (LG+G+I). (C) NMDS of evolutionary divergence between helicase domain sequences with scaling representing euclidean distances for three dimensions (stress: 0.08666). See Table S2 for taxa information.

Prokaryotes	M_phaeus F_chilense	Myroides phaeus	WP 090404604 1
	M_phaeus F_chilense	Myroides phaeus	WP 090404604.1
	F_chilense		
		Flavobacterium chilense	WP_068841780.1
	C_lonarensis	Cecembia Ionarensis	WP_009185623.1
	P salivibrio	Pontimonas salivibrio	WP 104912779.1
	C Zambryskibact	Candidatus Zambryskibacteria	OHB14600.1
	Algoriphagus sp	Algoriphagus sp.	WP 100627322.1
	A bacterium	Alphaproteobacteria bacterium	OJV13697.1
	C Vogelbacteria	Candidatus Vogelbacteria	OHA59397 1
	Aalborg AAW1	SR1 bacterium Aalborg AAW-1	AKH32407 1
	Gulosibacter sp	Gulosibacter sp	WP 087008023.1
	Bacteroides sn	Bacteroides sp	CDC65823 1
	S novella	Starkeva povella	P7084937 1
Fungi	5_HOVENa	Starkeya novena	12004557.1
	P_parasitica	Parasitella parasitica	CEP10706.1
	G dilepis	Gymnopilus dilepis	PPQ64766.1
	C cinerea	Coprinopsis cinerea	XP 001829007.2
	H_opuntiae	Hanseniaspora opuntiae	OEJ83279.1
	T_phaffii	Tetrapisispora phaffii	XP_003684282.1
	E granulatus	Elaphomyces granulatus	OXV06635.1
	R clarus	Rhizophagus clarus	GBB91117.1
	T_mesenterica	Tremella mesenterica	XP_007002293.1
	A glauca	Absidia glauca	SAL95951.1
	Termitomyces sp	Termitomyces sp.	KNZ79783.1
	Leucoagaricus sp	Leucoagaricus sp.	KXN86260.1
	S_stellatus	Sphaerobolus stellatus	KIJ35046.1
	S_cerevisiae	Saccharomyces cerevisiae	NP_013650.1
Mammals			
	F_damarensis	Fukomys damarensis	XP_010639595.1
	H_glaber	Heterocephalus glaber	EHA98492.1
	S_boliviensis	Saimiri boliviensis	XP_010349962.1
	S_araneus	Sorex araneus	XP_004619712.1
	M_murinus	Microcebus murinus	XP_012614176.1
	H_sapiens	Homo sapiens	NP_079325.2
	S harrisii	Sarcophilus harrisii	XP 012398677.2
	M domestica	Monodelphis domestica	XP_007479627.1
	G variegatus	Galeopterus variegatus	XP_008566201.1
	C cristata	Condylura cristata	XP 004687737.1
	E edwardii	Elephantulus edwardii	XP 006899697.1
	O_afer	Orycteropus afer	XP_007956003.1
lelentron			-
	Helen_A_aeg	Helitron-2_Aae (Aedes aegypti)	Helitron-2_Aae ^b
	Helen_D_rer	Helitron-2_DR (Danio rerio)	Helitron-2_DR ^b
	Helen_D_kik	Helitron-1_DK (Drosophila kikkawai)	Helitron-1_DK ^b
	Helen_N_vec	Helitron-1_NV (Nematostella vectensis)	Helitron-1_NV ^b
	Helen_M_cir	Helitron-like sequence (Mucor circinelloides)	EPB86818.1
	Helen_C_gig	Helitron-10_Cgi (Crassostrea gigas)	Helitron-10_CGi b
Helitron2			
	Hel2_F_oxy	FoHeli1 (Fusarium oxysporum)	FoHeli1 ^b
Helitron	Hall A sha		11015460.4
	Hel_A_tha	HELIIKON1 (Arabidopsis thaliana)	AAD15468.1
	Hel_c35	Hel_c35 (Cotesia vestalis bracovirus)	AEE09607.1
	HeI_M_luc	HELIBATI (Myotis lucifugus)	HELIBAT1
	HOLD DID	Heutron-1 AN (Aspergulus pidulans)	VD 6679971
	Hel_A_IIId		NF_002882.1

Supplementary Table S2. Taxa used in the helicase domain analysis ^a

Notes: ^a Prokaryotic, fungal and mammalian sequences were retrieved from Genbank (Benson et al. 2017) by using Helitron sequences as a reference. ^b Sequences retrieved from Repbase (Bao et al. 2015).

References

Bao, W.; Kojima, K.K; Kohany, O. Repbase Update, a database of repetitive elements in eukaryotic genomes. *Mob. DNA* 2015, 6, 11, https://doi.org/10.1186/s13100-015-0041-9.

Benson, D.A.; Cavanaugh, M.; Clark, K.; Karsch-Mizrachi, I.; Lipman, D.J.; Ostell, J.; Sayers, E.W. GenBank. *Nucleic Acids Res.* 2017, 45:D37–D42, <u>https://doi.org/10.1093/nar/gkx1094</u>.

Kazlauskas, D.; Dayaram, A.; Kraberger, S.; Goldstien, S.; Varsani, A.; Krupovic, M. Evolutionary history of ssDNA bacilladnaviruses features horizontal acquisition of the capsid gene from ssRNA nodaviruses. *Virology* 2017, 504, 114-121, https://doi.org/10.1016/j.virol.2017.02.001.

Thomas, J.; Pritham, E.J. Helitrons, the eukaryotic rolling-circle transposable elements. *Microbiol. Spectr.* 2015, 3, MDNA3-0049-2014, <u>https://www.doi.org/10.1128/microbiolspec.MDNA3-0049-2014</u>.

Wang, Y.; Sima, L.; Lv, J.; Huang, S.; Liu, Y.; Wang, J.; Krupovic, M.; Chen, X. Identification, characterization, and application of the replicon region of the halophilic temperate sphaerolipovirus SNJ1. *J. Bacteriol.* 2016, 198:1952–1964, http://dx.doi.org/10.1128/JB.00131-16.

Zawar-Reza, P.; Argüello-Astorga, G.R.; Kraberger, S.; Julian, L.; Stainton, D.; Broady, P.A.; Varsani, A. Diverse small circular single-stranded DNA viruses identified in a freshwater pond on the McMurdo Ice Shelf (Antarctica). *Infect. Genet. Evol.* 2014, 26, 132-138, <u>https://doi.org/10.1016/j.meegid.2014.05.018</u>.

Supplementary Data S1. Trimmed amino acid sequences used in the alignment

>MSV

VNTFLTYPHCPENPEIVCQMIWELVGRWTPKYIICAQEAHKDGDMHLHALLQTEKPVRITDSRFFDIEGFHPNIQSAKSVNKVRDYILKEPL >WDV KYLFLTYPQCTLEPQYALDSLRTLLNKYEPLYIAAVRELHEDGSPHLHVLVQNKLRASITNPNALNLRMFHPNIQAAKDCNQVRDYITKEVD >BMCTV KNIFLTYPRCSVIKEDALEILKNIPCPSDKLFIRVSQEKHQDGSLHLHALIQFKGKAQFRNPRHFDITHFHPNFQGAKSASDVKQYIEKDGD >TYLCSV KNYFLTYPKCDLTKENALSQITNLQTPTNKLFIKICRELHENGEPHLHILIQFEGKYNCTNQRFFDLVSFHPNIQGAKSSSDVKSYIDKDGD >CLCGV KNYFLTFPKCSLTKEEALEQIQKISTASNKKYIKICRELHEDGQPHLHVLLQFEGKFKCQNQRLFDLVSFHPNIQGAKSSSDVKSYIDKDGD >SsHADV KYVLLTYAOCELDAFRVMDKLSLLGAECIIGREHHEDGGTHLHCFAEFGRKFRSRKADVFDVDGHHPNITSRGTPEKGYDYAIKDGD >PFFFGmV RYALLTYAQCDLDPFAVVNHLAELAAECIIGREDHADGGIHLHAFVDFGKKYRTRNTRTFDVEGYHPNISSRRTPEEGYDYAIKDGD >HPAGmV RFCIVTYSQTDFDADAIVRILHRDCRGCIVARESHLDGGTHYHAFVDYGTPRDWTNSRRWDVLGVHPNIKVSRTPFNAYAYVGKDKN >pPpulchr RLFFLTYPCGLTKELILRELRKIVVVVSKERESGDGYDHFHVLLEAKTKKNYKDPRCFDILGVHGKYETVRNRKRSLKYICKEGD >pEcOYNIM QNIFLTYSQCDLSKEEIKTFIINLCNEKKLQINYLIIGIENHQDKGKHHHVFFQLNKQFRTRDLTIFNIPKYSPHIEPIKDTTDVRNYVKKDGD >pPASb11 KDIFLTYSKCPLGKEKIHNHIKQLMESKNQKIAYIISNTENHQDKEIHTHVLFQLNKRCNLTSQRFFDLDGYHPKIENTRDVEKAIEYIKKDGD >pPAPh2 RDIFLTYSKCPLGKEKIHNHLKQLLASKKKEIKYIISNNENHQDKEIHTHVFIQLKKQIEITNQRFFDIEGYHPKIETARDVEKSVSYIKKDKD >pPaWBNy KDIFLTYSKCPLGKDKIHNHIKQLMASKKKEIQYLITNQENHKDKEIHSHVLFQLTKSATFNGERFFDIEGFHPEIEVARDIEKSISYIKKDGD >BBTV VCWMFTINNPTTLPVMRDEIKYMVYQVERGQEGTRHVQGYVEMKRRSSLKQMRGFFPGAHLEKRKGSQEEARSYCMKEDT >FBNY >MVDC2 KRWCFTLNYKTALERETFISLFSRDELNYFVCGDEIAPTTGQKHLQGYVSMKKLIRLGGLKKKFGSIAHWEIAKGDDFQNRDYCTKETL >FBNS ${\tt ICWCFTLNNPLSPIFLHESMKYLVYQTEQGESGNIHFQGYIEMKKRTSLAGMKRLIPGAHFEKRRGTQGEARAYAMKEES}$ >SCSV ICWCFTLNNPLAPLSLHESMKYLVYQTEAGDNGTIHYQGYVEMKKRTSLVQMKKLLPGAHLEKRRGSQGEARAYAMKEDS SPOU KRWVFTLNNPSEEEKNKIRELPISLFDYFVCGEEGLEEGTPHLQGFANFAKKQTFNKVKWYFGARCHIEKAKGTDQQNKEYCSKEGH >SGCV KRYVFTLNNYTTEEYARIDNVGADGLARYMITGKEVGENGTPHLQGFINLKVKKRFSQIKEMLGSRCHIEKARGTDLENRVYCSKEGS >ZFCV KRWVFTLNNPTEQEVESVKSLPPSEYHYAIVGKEKGEQGTPHLQGFLHLKKKVRLNQMKQLIPRAHFEIARGSDEDNEQYCSKEGD >HSCycl RRFCFTWNNYTELNYALCQEFIKKYCKYGIVGKELAPTTNTPHLQGFCNLQKPMRFSTIKKRLDNGIHIEKSMGSDTQNQTYCSKSGE >DACycl RRFVFTWNNYTPSDFETCITFLDNFCKYGIIGKEKCPTTQTPHIQGFCNLSKPMRFNNIKKHLHNSIHIEKANGSDEQNKIYCSKSGE

LLHKTK

TKFMYTQQLKYLNLSIEQLKNNLENDAYIQDFAMINHNKDLDENNQNVAEHLHVFIKLNQQKTIDYVADLVDDKAQYIEFFDKSNKSRNEQNGYLY

>pSAP110B

MHAVIELPSKRDLSFISTAIGIRPEQIEVPRGRYGRENMLAYLVHAKD

>pOA504 SVFGFTQQFKADMWDWADDEKAVCFPNGVPDTARIMKRVAERLYVYLIGDIKKANAPDRPHAKDLFKYSAIIHDKDMSFAWDTKTNSKVIVPKELH

>pLa

 ${\tt RQFMYTQDLDHLPFKKEDLKTLLEKSSAEEWAYILHDKDIGKNGKTIRPHFHVVMKFKDAKTISRVAKLFNDKQEYIEVWRNTIGNAYSYLIHETS$

RTFMYTQQLQHLPFQDVAAFQSRLENINVAEYAFIIHDQDTVDGHPVTSHIHAVLRYQNARSVDSVAKQVSDKAQYIEIWNGNYANAYAYLVHKTD

LLYLTHANR >pLm

CELVIKADLIKQTEIEKVLESKKKVIQSYAFILHDNDKYLNEKEAKENGKSVGDYKIPHWHIMLRFHQSQEFKYIAKWFNTTENFVSQIKGRFTDA

>pCPa

LMYMIHANR

CELVINKTLITKTKIETILETKKKAIQNYAYILHDKDIYQNEKEAQLNGKKVGDIKAPHWHIYLRFNYSQDTKHISQWFNTQENFVSKIKGRFSDA

>pOYM

CELVINANKITKSKIENILELKKKAIQNYAYILHDKDTYQNEKEAQLNGKKIGDLKSPHYHIYLRFNYAYDTKHIAQWFNTQDNFVSKIKGRFSDA LMYMTHANS

>pAYWB

TDWLLTIRRELPDGSERTVDDVVNALQGIFDAAIGQPEKGEGGYRHYQIFAQGKRQRFSTLKKKLTAAGLGDAHVEPRKGSVSEAVGYCSKEKT

>p4M

RSGLLTIHPPSSHPSWLKPETWFPQCDDILEIWCAKFEKGEDTGNLHVHIYFKLKHSNTIRFELLQKWITKHVTGFDFKPQRSATKNSTQCVVNYV LKPET

>AHEaBV

GRCIVTLFPPDSEPKWLDPSTYYTDPASVVKIWVGOFEITPETNOIHAHIYIEFHHKKRPKFNLFVKMFTDIGKHVNVKSPKKSNNTOROGAVNYC MKDET

>AcrBV1

SRCIVTFFPKDNDRRWLKPETYFGPNPDNFQCWCGQFEICPRTGALHAHIYFECVRSRRLRFVRTAALFRKYHHRVHIKKARTVSKKQRQSAINYV LDDAK

>CsalDNAV

RYWLLTIPYEHFTPYLPPNCAYIKGQLEQGSNTSYLHWQLVVYFSQKKSLNYVKLIFGDGIHCEPSKSKAAEEYVWKEDT

>BcCircV

RYYMLTIPYSLFTIPDPLPEGLVWLKGQPERGENGYEHWQLICCTRKKCRASAVKRLFCPQAHVELTRSAAADEYVWKDDT

>RsaCircV

>MpaCircV5

>MpaCircV4 KYWVFTWHGPPKDDEGNRASPALWPEPQFDADMMDALQYQMEIAPSTGKYHYQGAVAFKTRKRSDPLREALAIPGAWTQMMRGSDKDQVYTNKEET

RNFVFTWNNYSDASKTYLSTLACKYVAYAEEVAPTTGTRHLQGFIAFTNAKTIQQARSKLPGCHVETMNGSIAQSEDYCSKAGT

 $\texttt{RCVCVTIHVDNIFWELQKWNQSLTYGIGQLELGLNGSTHWQMYFENNTAISLTQWKQLLGCKRAHVETRKGTALLAIEYCKKEET$ >MpaCircV3

>MpaCircV2

>MpaCircV1 KHWQFTLNNPTQDERNVLAELGDQPTTQYLIYGDEVGASGTPHLQGHVSFVQRYRFNQVKNWVSPRAHLELVRLLRRHIEYCKKDGA

>SARCircV

>DCircV RNWVFTLNNYVDADRVVIGERLANDATYVCYQPEIGASGTPHLQGLVVFANPRTLGGVKRLISDRVHLEPMRGTFAEAHAYCSKDDT

>CACycl RRFVFTWNNYPIEAYDKCEKYLTKFCKYGIVGEEIAPETGTPHLQGFCNLHKPTRFSTIKKHLDNSIHIEKANGSDIDNQKYCSKSGI >pMV158

TFLLYPESIPSDWELKLETLGVPMAISPLHDKDKSSIKGQKYKKAHYHVLYIAKNPVTADSVRKKIKLLLGEKSLAMVQVVLNVENMYLYLTHESK

>pE194

TFVLYPESAKAEWLEYLKELHIQFVVSPLHDRDTDTEGRMKKEHYHILVMYEGNKSYEQIKIITEELNATIPQIAGSVKGLVRYMLHMDD

>pADB201

TLLVYPDSAPENWKEILDQNGVEYFGALHDKDVNPDGTIKKPHYHIVLAYSGPTTFNNVKTLCNTLNSPKPLPLDGVGGMWRYMTHKDN

>pWV01

GFLLYPDSIPNDWKEKLESLGVSMAVSPLHDMDEKKDKDTWNSSDVIRNGKHYKKPHYHVIYIARNPVTIESVRNKIKRKLGNSSVAHVEILDYIK GSYEYLTHESK

>pFTB14

GWIFLTLTVRNVKGERLKPQISEMMEGFRKLFQYKKVKTSVLGFFRALEITKNHEEDTYHPHFHVLLPVKRNYFGKNYIKQAEWTSLWKRAMKLDY TPIVDIRRVKGRVKIDAEQIESDVREAMMEQKAVLEISKYPVKDTD

>pUB110

RWLFLTLTVKNVYDGEELNKSLSDMAQGFRRMMQYKKINKNLVGFMRATEVTINNKDNSYNQHMHVLVCVEPTYFKNTENYVNQKQWIQFWKKAMK LDYDPNVKVOMIRPKNKYKSDIOSAIDETAKYPVKDTD

>pBC1

QWLFLTLTVRNTSPESLPETISAMFEGFNRLTKYKAFKTSVKGYFRALEVTKNRDPHSEWFGTYHPHFHVLLCVPSSYFKKKELYITEQEWTDLWK KAMKLDYTPIVHVQRVKPKEQLEDMETYEEQLKNAIREQNAILEVSKYPVKDTD

>pKYM

RWLFLTLTVRNCEIGELGTVLTAMNAAFKRMEKRKELSPVQGWIRATEVTRGKDGSAHPHFHCLLMVQPSWFKGKNYVKHERWVELWRDCLRVNYE PNIDIRAVKTKTGEVVANVAEQLQSAVAETLKYSVKPED

>pSK89

QFIFLTLTTPNVTDEHLESEIKNYNHAFQKMFKRKKVNAITKGYVRKLEITYNSKRDDYNPHFHVLMAVNKSYFKDTKAYISQKEWLNLWRDVTGI SEITQVHVQKIKQNSNKELYEMAKYSGKDSD

>pNost

RWLFVTLTVKNCAITDLRETLTWMNKSFKRFSELKAFPAEGYIKTVEVTRGKTPDGSAHPHFHVLMMVKPSYFGVGYLSQAKWVEMWRKSLRVDYK PILDVQSLNPQDSLIGLLAEVIKYSVKESD

>pTD1

DFIFITLTVKNCSADELPATLEMMTKGWRRLAMTAMCEFRRSFEGTFKALEITVNKKTGEYHPHYHILAAVKKGYFRKSNPDYISQENLIKLWQKV CKLDYEPNVDIRRVKNSTYKAVAEVAKYSVKATD

>AAV2

YEIVIKVPSDLDEHLPGISDSFVNWVAEKEWELPPDSDMDLNLIEQAPLTVAEKLQRDFLTEWRRVSKAPEALFFVQFEKGESYFHMHVLVETTGV KSMVLGRFLSQIREKLIQRIYRGIEPTLPNWFAVTKTRNGAGGGNKVVDECYIPNYLL

>AAV5

YEVIVRVPFDVEEHLPGISDSFVDWVTGQIWELPPESDLNLTLVEQPQLTVADRIRRVFLYEWNKFSKQESKFFVQFEKGSEYFHLHTLVETSGIS SMVLGRYVSQIRAQLVKVVFQGIEPQINDWVAITKVKKGGANKVVDSGYIPAYLL

>SLP

WELVIKLKYDWIEDLEGSDDPWYDWPEDEIDIYMAILGIKAIKAITRVLRERSKNKTCNYFGQIEQGGEFFHIHLLFEVDGFVSFLLGRMFETLRQ TLRNSVYFGYPFEVSSEIAITKVKTGGRNKVQDGSYIVNYLL

>SNJ1

HHSVISPPEELYIDAEFPEQELISVAQEFMEEIGMQGIALYHSWSGGDDHDDDIGEWKKRLFADRDWHGDVREELQHRPHVHLIGACPWFPMGDVT KLTHAETDWVIHRITGKRDGNSSVSLADMRSVARAVVYALSHCA

>H_inordinatus

HHVVFSPPRDWFLQAQDPLDKTFKLIGDILTNHFDAAGRVYYHGWSGGDDLEDDLGEWKNRLFEGRDWETDVRHELEPRPHFHAVVASPFIPGEGV TDRIHDETGWVIKRIADEKSKRSIDGIDALARVVTYCMSHTS

>H_thailandensis

 $\label{eq:construction} I \texttt{HAMFSPEQDWTISRVDGMRSESYELAQEAGVTGGGALL\texttt{HMWRTTDDLDGEFKKWKYRETYGQGWRQATEVAPHVHQIATAPEFEPEQGDWVAKRVRTLDAMRSLS\texttt{HPSSYEDVAGLAMYLLSHTA}$

>CN_piranensis

LHNIVSIPFELYLTKDGRKKLRAKAIKYLKEFDIDGGVMIDHPYRFSKDLESARLSPHLHLIVTGWLDGQKVKELYEKTGWIVTNVSTIETWNDCY NLSKYLLSHSA

>Therm BRNA1

VHVVVSPPQDLRFMRSKEGFRIMVNKVIRVLKDFQVDTGALVFHPWRQCGDRDGSFPSSSFVWRAGPHFHAVGYGYVPEDRIKEFHERTGWILKVV HDKSDVVSPTATLAYLLTHAG

>Thaum SCGC

IHLILAVPENQRELPVKLLRQRMSHILKLGNIKGGSVIFHPFRFSKTQHRWYASPHFHLVGFGKSSDIKNAFGRYGWYVKEAGERESVFQTFCYLL SHCG

>IS91

QHIVFTLPCQYWSLVFHNRWLLAEMSRIAADVILEICHOTDVEPGIFTVIHTWGRDQQWHPHIHLSTTAGGVTSGHTWKNLHFYARKVMSMWRYRI

TRLLSRKYPELVIPDELAVGNSKRDWNCFLDTYRRGWNVNISRVMDNATHVAVYFGSYLK

>IS801

QHLVFTLPDTLWPLFFYNRWLLDALFRLAADNLIYAAKRRGLRVGIFGALHTYGRRLNWHPHVHLSVTAGGLDEQGVWKNLSFHKEALRRRWMWLV RDYLLGQPLSQLTMPPPLAHILCESDWRRLILAGGQHWHIHLSKKTKNGRKTVNYLGRYLK

>IS1294

VHLVFTLPDTLWPVFESNRWLLNDVCRLAVENLLYAARKRGLEPGIFCAIHTYGRRLNWHPHVHVSVTCGGLNKHGQWKKLSFLKDAMRSRWMWNM RQLLLKAWSEGMAMPESLSHITTESQWRSLVLKGGKYWHVYMSKKTAGGRNTARYLGRYLK

>ISCR1

RQWVLSFPFQLRFLLARHPQLLSIVYRTLSTHLIKKAGYTKASAQTGSVTLIQRFGSALNLNVHYHMLFLDGVYAEDDYGKQRFHRKALAHTLSHR IARCMEKRTLTQLHGASVTYRIAVGPQQGRKVFTAGFSLHAGVMAEAHQRDKLERLCRYIS

>ISCR2

RQWVLSFPFQLRFLFASRPEILGIVYRVIATHLVKKAGHTHQVAKTGAVTLIQRFGSALNLNVHFHMLFLDGVYVEQSHGSARFRWKALTHTIAHR VGRYLERQPMTPLLGHSITYRIAVGSQAGRKVFTAGFSLHAGVAARADERKKLERLCRYIS

>ISCR3

RQWVLSFPYPLRFLFASKPEALGIVQRVIAGWLADQAGIDRASAQCGAVTLIQRFGSALNLNIHFHMLWLDGVYVEATRRELRLHRRALAATIAHR VCRHLTRKSMDGLRMSSITYRIATGRDAGCKVVTGGFSLHAGVAAEAHESHKLEKLCRYIT

>IS608

HNVVYSCKYHIVWCPKYRRKVLVGAVEMRLKEIIQEVAKELRVEIIEMQTDKDHIHILADIDPSFGVMKFIKRILRQEFNHLKTKLPTLWTNSCFI STVGGAPLNVVKQYIENQQN

>Rhiz NXC24

RIVVPDIPHHVTQRGNGRAQTFFCDDDYALYRDLLAHHCRAADVEVWGWVLMPNHVHLILVPADADGIRRALRVHRAYAGHIHARLRRTGHFWQGR FGCVPMDEEHLAAALRYVALNPV

>ISDra2

RGYVYQLEYHLIWCVKYRHQVLVGEVADGLKDILRDIAAQNGLEVITMEVMPDHVHLLLSATPQQAIPDFVKRRMFVAYPQLKEKLWGGNLWNPSY CILTVSENTRAQIQKYIESQHD

>phiX174

FIVFDTLTLADDRLEAFYDNPNALRDYFRDIGRMVLAAEGRKANDSHADCYQYFCVPEYGTANGRLHFHAVHFMRTLPTGSVDPNFGRRVRNRRQL NSLQNTWPYGYSMPIAVRYTQDAFSRSGWLWPVDAKGEPLKATSYMAVGFYVAKYVN

>phageNC3

FFVFDTLTLADDRLQAFNENPNALRDYFRTVGRAVLRAEGRSVKDSYNDCYRYLCVPEFGGQHGRLHWHVVHMVRTLPLGSHDPNFGRKVRNYRQI NSFRGMWPYGFTQPIAVRYQHDAYSRKGWLWPVDKSGKAMQSKPYQAVAWYVTKYVA

>ERBP1

YCIFNTLTVNESSIEKVFEKGSRIFSDYVRSLDRGVGIAIHKNWRQAVTKRKEGNEFHTYFAVVERGTKNGRLHIHVIHMMKELPNGCVDPNAGRA IPNRREVTYLKRYWKYGYSAPIAVRFNTNDAFGKKYWRFPVKEVAKNRFESLECKDAGSIIGYIGKYMT

>P2

VGMFITLTAPSKYHPTRQVGKGESKTVQLNHGWNDEAFNPKDAQRYLCHIWSLMRTAFKDNDLQVYGLRVVEPHHDGTPHWHMMLFCNPRQRNQII EIMRRYALKEDGDERGAARNRFQAKHLNQGGAAGYIAKYIS

>Sphage_RE2010

CAVFYTITCPSRFHSTLNNGRPNPTWTNATVRQSSDYLVGMFAAFRKAMHKAGLRWYGVRVAEPHHDGTVHWHLLCFMRKKDRRAITALLRKFAIR EDREELGNNTGPRFKSELINPRKGTPTSYIAKYIS

>phiE122

RGVMFTLTCPSRFHAVTTTDSWVRPNPRYDDVDPRAAQAYLRKVWQRTRAELKREGIVYFGMRVAEPNHDGTPHWHGLVFADKIERFCSVMRKHGL RDSGDEPGAQRHRVRFEMIDRAKGSAVGYVAKYIS

>phi_Lf

AWYFLTLTYRDGSDSSPRDVSELFKRMRGHFNRLKSGRARWNRESFRYVWVGELTQRFRPHYHVMLWVPQGMFFGKVDQRGWWPHGSSQIEKARNC VGYLAKYAS

>SVTS2

NLSFLTLTYAVNEKDVKKCKNDLKLFFNNINRWWNNPIRSKNHKGILKYMYTYEYQKRGAVHFHIILNQKIPNSVVQQYWKHGINKNIKVRAGSNE DVVKYLAKYIV

>Rhizob R404

RPAMLTLTYREVGQWNPKHISDLLQRIRVWVRRRGHGLRYVWVAELQQRGALHYHLLLWLPRGLTLPKPDKQGWWTHGSTRIEWARKPAGYLAKYA

>RSIBR1

VTHMITLTTRECITDLDWFLGLWDAFRRAMARYSQFHYIAVPELQKRGAWHMHVAVSGRVALNLARRVWLKVVGGRGKGYCHIRNPQGAHFGKQWK LDALASYVAKYIG

>GkshoV Hs

SNYFVTLTYRPDALPYTKDGKPTLRPKDLTNFFKRLRKHKKGNEKIRYFACGEYGEKKGRPHYHVALFNLKLDDLKPLGPSQGYMLYKSKTLQNIW GLGFVVIGELTYKSASYISRYVM

>GkshoV_Bird

 $\label{eq:construction} ENYFVTLTYDNDNVPLSQMHMNTLKKRDFQLFMKRLRKRGNDGIRFFACGEYGSTTMRPHYHAILFNLHLDDLEKLYEKDGMVYYTSQFLQSVWKKGFVIITSMTWETCAYVARYVC$

>GkshoV Marine

SSSFITLTYDNKHLPPNNSLDYTHWQKFIRSLKKRNNGKSIRYFGVGEYGENFGRPHFHAILFGHTFNDLIPMHSNISKSQQLLSAWPRGFVSVGD VTPESISYVCGYVQ

>HRPV1

SGVMVTLTTDPKRYDSMLDGLMDAWQNLHETLNYLEGTRLDFIRALEFGGSGLPHLHVCVFGVPYIDHRWLKHYWSHAEIVHIHGMNKRGNDSWIM TSGTHAGKSVAGYLGKYLS

>HRPV2

NAVFCTLTTDPKKFDSLYDAVMSINENFHRLMSYLRSVTGRPRETLDYIKVLEFTSAGYPHLHVLFFDVPWLVDKRELSAKWKQGQIVDLYPLVHR DDDDWVEEQTRSDDVYQSKTAGSYVGKYIS

>H_rubripr

NAVLVTLTTDPKRQDSLLDGIDSINENLNRLLSYFDSVTGRPRDRPDYIKALEFTEKGYPHLHVLFFDVPWLCDKSEVAAKWAQGEIVDVYPLTYR DDEDWVRERTRDDGHEKESTAGAYLGKYLS

>pPhasyl

NVGFLTLTFRDHVTDPKEAQRRFNSLKTNILAKRYRAYIRVMEPMKSGRIHYHLLVALHSDIRTGFDFPAVYRQDYSSANKAIRSEWSFWRKTAPK YGFGRTELMPVRSNSEGIGRYVGKYIS

>pGL3

RLSFITLTLPPAVAEDLSGRWAHVVDLMKRRLPTEIIACTEVQEKVALHLHIVMVGRHSRGSPRQLEKMWSECCETAVRNVIEPNERVTSRVTNSR TESESNGNGNATGNTSSNANSNGNANGNIHTEVNWNAAVNVQRIKKSASAYMGKYLS

>pHT926

KPVFMTLTFAENVTDVDLANKAFKQFIRKLNGHVYGRGRVGLKYVTVIEFQKRGAVHYHCVFFNLPFIDSGVIASLWGQGFIKVNSMKKRDGTNCD NVGAYVTKYMQ

>pSA1

 $\label{eq:prvfatltapelgipldpatydasdlwryftiylrresrvsfkvaeyqkrgavhfhavirfdgagdqpartlhwgtqldvqpigafghgeeiteqavasyvakytt$

>pUnnamed2

KSTFLTLTFKENIQDIERANREFTLFIKRLKRYLKNQQLKYIATWELQQRGAIHYHLVLFSVPYIDNKKLGELWANGFIKINKIKETVKNEAVGVY ITKYFV

>pHGN1

HTAMVTLTASTTEEDGGPRPLVDHLRDLLSSWSAVYDALRHTLEDREFEYLAIIEPTPAGYAHIHLGVFVKGPVVAEQFQDVLDAHVKNSEGAGRE AHRAVVEDDEDEAAVSIRRSARPDREDGIENLGAYLAAYMA

>pGRB1

 $\label{eq:htgmvtltasstddegrlrplehfedllesweavrralarvlegreweylailephesgyvhihlgvfvrgpvvaeqfepvldahlrncptaged ahqvfdengdedavrvrrsshpsrsggvenlgaylaayma$

>pZMX201

 $\label{eq:htmltftassrpngpppdhldellaswdalttaldrvlgdrryarlgilephnngylhihvavfidgkveqedfapvirshvnnceyatedahdptsedtisirhagdpkrdsdvigelaiylaeylg$

>pHF2

TTAMLTLTASHRNEKGGWRCPADHMRDIMDGYDAARKQLHQVLSGRKWEYARVWEPHADGYGHLHIAVFVEDDLRADDFEPVMRSHVENCGPAGSK AHDPAGDSVSVRDDVENLGSYISEYIG

>pHK2

ATAMLTFTASSVPNGERLPPVEHTDALHDSYDGVRDTLRNTLDADEWGYWLQAEPHNACYSHLHVGVYFDAAVVGPEFERVIDKHVEECEYASFSA HDYRNTDYLNDSISLNAGVENMGSYLAAYMG

>pNB101

TMVMVTLSASSENAKGGRRCPADHMRDIARGWNSARKALHRVLRRFEWEYAKVWEPHQSGYGHMHVAVAVDDPIEGETFRPVVRSHVENVEPAGSA AHGLNAVGMGDTVSVNREVENLGSYISEYIG

>pTP2

DAVFLTLTTDPSRFSNLYEANRQFSHSFNRFMSRLRGYFARRGQHLEYIAVYEFTKSGLLHAHVIIFGVRYVISRWWSQGRVVYIYRLRNVDGRWV WARRPRDVRAGEGAEDYLKKYLR

>pML

PITMITLTTYQDSQYSVKKHKVDHEQALEMLVDGFRKLRELITRICEGHTPDYFWILEPHESGYPHMHLCYLEEFTEGEQEHIKSIWGAGEQVDFS FRKPEDTVRSIRNYLMKYMS

>Helen_A_aeg

PTMFLTLSASETQWPLLLKQLHKLTLVNDDAVTCCLYFNKLVDVLMGILSSPRYVVDFFKRIEFQHRGSPHAHIMLWLANDPNETVSELIRKVCSI SAIHLSETISHTFTCYKRNEKRCRFNIPYWPMNEERTLYEYYLDVLRSSIQRPTIFLKRSMNEMWTNPFNPWIAEKLRSNMDLQFILDVYSCACYL AGYVN

>Helen D rer

PTFFCTFSAAEMRWPEIVTVIKAQEILRSNPVTVMRMFEKRVDALMAHLLLSPEVEDFFYRVEFQARGSPHIHLLAWVKDAPDPEEDNFIDRYVSC KLPDPNVDPELHKIVTNHSKSCKKGKVVCRFGFPKLPMPKTMITMDDYLNYAEGLTTGSAVLLKRDPKETWVNGYNPDLLRAWNANMDIQYILDAY SCIMYMLSYVS

>Helen_D_kik

PTFFITFSAAESKWNELLVTLSRLRLIRSDPVTCSRYFDFRFRQLIKLFKSSETLVHYYWRIEFQHRGSPHSHGMYWFSGAPKLEGPEFIDRFITT TGDDPELQEVIKHSSSCLREGQEFCRFQMPYPPMPETMVLFEEYKFAIRSSLKKPQVFLRRKFSDRLVNAYNRDILGLHRANMDIQFILDAFACCS YIINYIN

>Helen_N_vec

ATLFCSFSSAETQWMHLLRILGQLRLIQSDPVTCARHFDYQVNQFLTNFLFSSKISDWFYRVEYQQRGSPHIHMLMWLEDAPQFQIDSFIDKIITC QKPVDNADLLVLVRHSHTCRKNTSSKCRFNYPQPPMKQTMIIKQNYLLAVSSSINTPTVFLKRNPNELRINNYNPDCLSAWRANMDIQFVLDVYAC AVYIVNYIS

>Helen M cir

PTLFITLSAAESKWTELLAMLKKIWLVQSDPVTCASYFDYRFRELKKTRTAPCNVQEYFFRTEFQHRGSPHIHMLIWLEDAPRILPDSFVDGIITC EKEWDGSPATWDDIIKHTATCKRKDQIVCRFNIPFLPMDVTRVLVDAYIYSIRSTLKTTKVFLRRTPNQVLTNSYNRKILSMFRSNMDLQFIVDGY ACCSYVADYIN

>Helen_C_gig

PTWFCSFSAAETKWIPLLKTLGKLRLIKSDPVTCSRYFDYRFQRFLHGVLLHKEVVDYFFRVEFQQRGSPHVHMLLWVKNAPNVSSDSFVDRYVSC SKSGADPVLVRHAKTCMKKNKPICRFNFPIPPMPKTVTLFETYTLAIRSSLTQSKLFLKRQPYEIRINSYNCTLLKSWLANMDIQFILDPYACATY IVSYIS

>Hel2 F oxy

PGAFITFSPADLHWRSLYQHMPQYRLLRQNPHIAAFHFYRRYTLFRDIVLSKKSITDYWDRYEWQGRGSPHNHGLYWMDNCPGADMEDTWGFHVTA INPEPSRTLRLSQIVEAANVANPERECRFDFPRALRELAAVIGRSYYVFEAARNDSLMNNFNPAIILGWLANIDISPCTSLAVITYAAKYCS

>Hel A tha

PDLFITFTCNPKWPHITRYCDKRLNPKDRLDIIARIFKIKLDSLMNDLTVKKKTVASMYTVEFQKRGLPHAHILLFMHAKSKLPTSDDIDKLISAE IPDKEKEPELYEVINVKSPCMVDGECSKLYPKKHQDITKVGSDGYPIYRRKIDDYVEKGGIKCDNRYVMPYNKKFSLRYNAHINVEWCNQNDSIK YLFKYIN

>Hel c35

PDLFITFTCNPKWIEITQLLLPGQTSSDRHDITARIFRQKIRSLMNFIVKQRDTRCWMYSIEWQKRGLPHAHILIWLVERIQPDQIDDIICAEIPD YEVDPDLHDVVNPQSPCMVDGKCSKRYPRKLTAETVTGNDGYPLYRRSPDDKVKRMDFVVDNSWIVPYSPLISKSFKTHCNVEYCNSVKSIKYIC KYVT

>Hel M luc

PDLFITMTCNPKWADITNNLQRWQKVENRPDLVARVFNIKLNALLNDICKFHKVIAKIHVIEFQKRGLPHAHILLILDSESKLRSEDDIDRIVKAE IPDEDQCPRLFQIVNPNSPCMENGKCSKGYPKEFQNATIGNIDGYPKYKRRSGSTMSIGNKVVDNTWIVPYNPYLCLKYNCHINVEVCASIKSVKY LFKYIY

>Hel A nid

PSLFITFTANPAWDEVTRELRPGETWEDRPDIVSRVFNILRAEMVDELCKKKVAPGRFFTIEYQKRGLPHMHLVLFLEERERFLDAAHIDEMVSAE LPDPREDLELYKLVNSRAPCCDKNMIYCTKRFPKAEQYETQPIEEGYPLYRRADPRGAYNDMVRIDNTWVVPYNPYLLKRFRSHINVEVCRGVDV IKYITKYIY

>Hel C ele

PDIFLTFTCNPAWTEISENLGPRQSASDRPDLIARVFKLKVVDALFDDLLNRDHVAAYISVFEWQKRGLPHVHMLLTMAENSKPRTSEDIDKIVQA EIPNPDNEPELHRIVNPHSPCMVDGHCSKRYPKDFHPSTTLNVDGYPGYRRRDDGRYVEYGTQHLDNRRVVPYNKWLLLRYNAHMNVEICGFIEAV KYLFKYVY

>Hel_A_gam

PDLFITVTCNPKWPEITQCLLPRQQAPDRPDVIVRVFRLKLKAILNDLTMGIEVARIHVIEFQKRGLPHAHILVILAEEDKPQTPADYDKIVSAEL PNPATSSQLFETVNPAAPCMKDGTCEKGFPKSFCEQTRSMDNGYPQYRRNNGRSVTVKGIELDNRYVVPYNPWFTHKYNCHINVEVCTSISSVKY LYKYVY

9.2 Material suplementar do Capítulo 2

Supplementary Material

Supplementary Figure S1. Conserved domains from sequences containing Pif1 domains. (A) Human Pif1 domain. (B) Best candidate for the genomic Pif1 sequence in the fungal species *Rhizophagus clarus*. (C) Example of a *Helitron* transposase sequence with the Rep and Hel (Pif1) domains. (D) Example of a second candidate genomic Pif1 gene from *R. clarus* structurally similar with the human Pif1 domain. (E) Coding sequence upstream from the ORF in (D) containing a Rep domain, indicating that (D) is part of a broken RepHel ORF starting in (E). Image from the Conserved Domain Database (CDD) search tool (Lu et al. 2020).

	ification			
DEAD-like_helica	se_N and SF1_C_RecD domain-con N and SF1_C_BecD domain-containing	taining protein (domain architecture ID	13718164)	
	Concern Marcol			10 . 10
Query seq.	No a state balling and it		AT hading .	esta A
Specific hits		PIF1		PLUM
Superfamilies	DEAD-116	helicase_N superfamily		DOG-12he_belics
List of doma	n hits	Description		hatara
PIF1	plano5970 PtF1-like helicase; This of 18922	amly includes homologs of the PIF1 helicas	e, which inhbits	4-302
-				
Conserved	domains on [gi 139440214	[2 dbj G8885025]		
hypothetical pr	cein RoHRI 01160019 (Rhizoph	agus darus]		
DEXSc_Pif1_like	and SF1_C_RecD domain-containing	ig protein (domain architecture ID 13030	209)	
DEXS: P#1 like a	nd SF1_C_RecD domain-containing pro	tein		
Query seq.	1 1 1 1 T 1 1 1	a site a second se		
Specific hits		DEXSC_PIFI_like		
Superfamilies		Of the believe & control	Red	÷
		and the particular of the same		
List of doma	in hits			
Name	Accession	Descript	ion	Inter
 DEXSc_PI1_III RecD super fail 	e od18037 DEAD-box helicase iliy p33920 ATP-dependent exi	domain of PIT: PITI and other members of t DNAse (exonuclease V), alpha subunit, heli	tris tamily are RecD-like case superfamily I [Replication,	213-213-
nypomental pr	Iden Konkt_01100033 [Knizoje	agus carus)		
Barry set.	end SF1_C_RecD domain-containin domains Heltron_Jike_N, DEAD-like_h	g protein (domain architecture ID 13835- ricase, N, and SP1_C,RecD	199) 	
Heitron_like_N protein containing Query seq. Specific hits Superfamilies	And SFI, C, RecD domain-containing domains Heltron, Jike, N, DEAD-Hike, N 214 Aldren, Jike, N Hiltron, Jike, N	g protein (domain antifications ID 13855- incase_N, and SFL_C_RecD	1993) PIF1	<u></u> r
Heitron_lka_N protein containing Query seq. Specific hits Superfamilies	and SFL_C_RecD forestainin-containin domains Haltron, Jike N, DEAD-Ske, N Million, Jike N, DEAD-Ske, N Million, Jike N, DEAD-Ske, N Million, Jike N, Status, S	g pretein (Jonan architecture ID 1305 Hone, N. and SFL, C. RecD Spa evening KP bindes of the Bindes and SFL (Second	1993) PTE 1	<u></u>
Heitren jike N protein containing Query seq. Specific hits Superfamilies	In hits	p probain (domain ant because ID 11055 Hose, N, and SPL_C, RecD	tan tan PTF 1 B Ked inne, 8 area faults	
Helitren_like_N protein containing Specific hits Superfamilies List of dome Name PP1	Accession Accession	a problem (dismute exclusionary of the PF) - Red minime (N, end SFL_C, Red)	199)	inter Inter
Helitron_Jika_N protein containing Query seq. Specific hits Superfamilies List of doms Name P PF1 Helitron_Data_N P RecD super la	In hits Accession Figure 2017	a problem (dismant exchances to 1.1855- microse, N. end SPL_C, Red)	INT)	
Helitren_Jike_N protein containing Specific hits Superfanilies Name Pifi = Heliton_Dau_N = RocD superfan	In Dits Accessing photometry like (Color Star, N Accessing photometry like (Color Star, N Accessing photo	a problem (dismute rechtericher Di 1955- nicises, N. end SFL_C, RecD ************************************	1993) TET 1 Martin Martin Ambrido In Nethron S. Hochty Case Andri Infolto In Nethron S. Hochty Case Angerlandy Perphatication,	1000 - 1000 455-11 972-310
Helitren, Jike, N protein containing Geory seq. Seperific hits Superfanilies List of dome P Pit Name P Pit Name P Pit Name P Pit Name	In Dits	Prefer (Insert instance to 1385) The second secon	1977) PIFI	100
Helitren, Jike, N protein containing Overy see, Specific hits Soperfaillies Name P/Fi N Name P/Fi N Name P/Fi N Name P/Fi N Name P/Fi N Name P/Fi N Name P/Fi N NA NA NA NA NA NA NA NA NA NA NA NA N	In the Read Annual Contract of the Second Se	Professional Annual Inclusions In 2015 Section 2	1997) The second secon	цер (рад. Войст (1952)
Heitren, Jan, JA preten containing Borry see, Specific hits Specific hit	In Nite Accession of the second seco	Perform (Instance Instance In 1995) Perform (Instance Instance Ins	ntri PEF Seat Seat Seat Seat Seat Seat Seat Seat	190
Heitrea, Jaa, J. Protein containing George seel. Seecific hite Seperfamilies List of domme Protein Case Conserved hypothetical pr Protein Class	An office and the second secon	protect (another inclusions to 13055 the set of 1, 2, 2000) the set of 1, 2000 the set	1977	ter ye dela 975 to
Heitren, Jan, J. Protein containing Serry see. Securities in the securities of the securities List of domin Protein containing and the security Protectical per Protectical per Protection Class Decontainum and the security Decontainum and the security of the security Decontainum and the security of the security Protectical person of the security of the security of the security Protectical person of the security of	And State St	Perform (Annual Inclusion In 2015) Perform (Annual Inclusion In 2015) Perform (Annual Inclusion Inclusion) Perform (Annual Inclusion)		, 192 , 192 Inter 1 6 192 975 ft
Heitren, Jan, J. Protein containing Serry see, Seecific hits Seecific hits Seecific hits Seecific hits Prot Name Heitren, Bie, M. Heitren, Bie, M. Protein Class Protein Class DDAN-like, Meloc	In the second se	preter (and a set of the set of t	norr 21 December 2010 Sector	- 40° - 40°
Heirren, Jan, J. Portal containing Derry see, Specific hits Specific hits Specific hits Heirren State Conserved hypothetical pr Procein Class DEAD-like, Julie DEAD-like, Julie DEAD-like, Julie	In http://www.initestance.com/	protection (contrast to the contrast to t		- 197 - 197 - 197
Heitrea, Jaa, J. protein containing Specific hits Specific hits	In Site Constant indiana, Sac X, Colonia M, Marian Indiana, Sac X, Colonia M, Marian I, Sac X, Colonia M, Marian I, Sac X, Colonia M, Mariana M, Sac X, Sac	Protection and and an endbandow to 13855 Marked, and 974, C. Nex.D Marked, and 974, C. Nex, Nex.D Marked, and 974, C. Nex, Nex.D Marked	1977 1971	
Helitera, Jan, J. Georg see, Specific hiles Specific hiles List of domini Protein Conserved hypothetical and Protein Clark Conserved hypothetical and Protein Clark DEAD-like helice Specific hiles Specific hiles	In the second se	preter (and a product or 100000000000000000000000000000000000	1977	in transformer
Norma See, See, See, See, See, See, See, See	MALE AND ADDRESS OF THE ADDRESS OF T	protect (name inclusion to 13855	Market Andrew Constraints of the second seco	ter in the second secon
Nerry see, Specific obtaining Specific hits Specific hits Specific hits Specific hits Problem Class Conserved Protoin Class Conserved Protoin Class Color-line Juliez, Hits Specific hit	ADD Second	pertent (and a collection to 1305 the second secon	series and a series of the ser	in the second se
Burry set. Seery set. Seering containing Seery set. Seering containing Conserved hypothetical or Protoin Case Secret, State, Jule Secret, State, Ju	In hits In	protect (name inclusion to 1385		
Halinea Jaa, J. Servery sea. Systelific hills Sager failles List of doma Part Resources Conserved hypothetical pr Protocharger have Server the hills Server the hill	In http: In htt	protection (content on the content on the 13055		Letter and the second s
Name Series Containing Series Containing Series Containing Series Containing Series Series Containing Series Series Containing Series Series Containing Series Seri	Market Alexandro established and an and a second and a se	protection contraction to 13055 the second	The second seco	197
Network of the second s	In the second se	protect (name inclusion to 13855	Hand State and S	r or or or of the second secon
Burry see, Seesific hits Seesific h	In the second se	perter (normal inclusion in 13855 inclusion in the second	entry 297 - 1 207 - 2 207 -	197 - 197 66 o c 66 o c 97 5 f 97 5 f 10 10 10 10 10 10 10 10 10 10 10 10 10
Berry see. Specific hits Specific h	In hits In	protect (name inclusion to 1305		ter traja mentanta ter traja mentanta ter traja
Advised Collaboration Alex Jack March Collaboration Alex Collaboration Ale	In this Consists on Carlos and C	protection constrained to 13455 protection	the second	
List of come Secret and	Market And American Control of Co	perturbative in 2 12055		
Survey see. Second Colomo Second Co	Market And State Contract of Contract of Contract on Contract	protect (name inclusion to 13855	the second	t under of main indef

Supplementary Figure S2. Phylogeny of Pif1-like sequences. Same phylogeny as in Figure 3 (main text), displaying branch support values and taxa names (see Table S1 below). Specifications of the procedures used for phylogenetic inference are described in the Materials and Methods.



Group	Abbreviation*	Taxon/Host name	Accession
Helentron			
	Helen A alb	Aedes albopictus	XP_029715674.1
	Helen D rer	Danio rerio	XP_021330385.1
	Helen L ser	Lucilia sericata	XP_037823532.1
	Helen N vec	Nematostella vectensis	XP_032223796.1
	Helen M cir	Mucor circinelloides 1006PhL	EPB86818.1
	Helen C gig	Crassostrea gigas	XP_019922950.2
	Helen C qui	Culex quinquefasciatus	EDS39572.1
	Helen G occ	Galendromus occidentalis	XP_028966621.1
	Helen L roh	Labeo rohita	RXN14713.1
	Helen A cal	Astatotilapia calliptera	XP_026026780.1
	Helen M sac	Melanaphis sacchari	XP_025191627.1
	Helen A mil	Acropora millepora	XP_029180665.1
	Helen D gig	Dendronephthya gigantea	XP_028394532.1
	Helen L ana	Lingula anatina	XP_013378814.1
	Helen C cuc	Choanephora cucurbitarum	OBZ82310.1
	Helen S pur	Strongylocentrotus purpuratus	XP_011671010.2
	Helen O fav	Orbicella faveolata	XP_020609775.1
	Helen A dig	Acropora digitifera	XP_015779364.1
		Bemisia tabaci	LIED01008227.1
		Perkinsus marinus ATCC 50983	XP_002772304.1
lelitron2			
	Hel2 F oxy	Fusarium oxysporum	AKC01507.1
	Hel2 P lil	Purpureocillium lilacinum	OAQ59778.1
	Hel2 P chl	Pochonia chlamydosporia 170	XP_018136201.1
	Hel2 F mon	Fonsecaea monophora	XP_022510545.1
	Hel2 M ani	Metarhizium anisopliae	KFG84029.1
		Ectocarpus sp. CCAP 1310 34	CAB1116976.1
		Papaver somniferum	RZC87713.1
		Symbiodinium microadriaticum	CAE7237458.1
Helitron			
	Hel A tha	Arabidopsis thaliana	AAD15468.1
	Hel X lae	Xenopus laevis	XP_041421549.1
	Hel C ele	Caenorhabditis elegans	NP_493834.1
	Hel A ara	Anopheles arabiensis	XP_040164812.1
	Hel B nap	Brassica napus	XP_022553550.1
	Hel B vul	Beta vulgaris subsp. vulgaris	XP_010692805.1
	Hel C sup	Chilo suppressalis	RVE40746.1
	Hel M dem	Microplitis demolitor	XP_008549021.2
	Hel F can	Folsomia candida	XP_021953640.1
	Hel E jap	Eumeta japonica	GBP49736.1
	Hel C pur	Claviceps purpurea 20.1	CCE31728.1
	Hel R del	Rhizopus delemar RA 99-880	EIE75949.1
	Hel A can	Ancylostoma caninum	RCN43056.1
	Hel N ame	Necator americanus	XP_013304266.1
	Hel P inf	Phytophthora infestans T30-4	XP_002905633.1
	Hel H ann	Helianthus annuus	XP_022020320.1
		Brassica rapa	XP_033143622.1
		Capsella rubella	XP_006279329.2
		Ananas comosus var. bracteatus	CAD1820584.1
		Erythranthe guttata	XP_012840144.1
		Helianthus annuus	XP_022031972.1
		Oryza sativa Japonica Group	ABA95557.1
		Setaria italica	RCV07316.1
		Panicum virgatum	XP_039834415.1
		Papaver somniferum	XP 026386115.1
		Aquilegia coeruleg	PIA60703.1
		Panicum viraatum	XP 039793773 1
		Fragrostis curvula	TV/127820 1
		Aegilons tauschii suben, strangulata	YD 020107274 1
		Aegilops tauschil subsp. strangulata	XP_020197274.1
		i nalictrum thalictroides	KAF5187279.1
		Papaver somniferum	XP_026391420.1
		Ceratodon purpureus	KAG0566608.1
		Phytophthora rubi	KAE9276432.1
		Plasmodiophora brassicae	CEO98944.1
		Chondrus crispus	XP_005716008.1

		Porphyra umbilicalis	OSX80228.1
		Streblomastix strix	KAA6365738.1
Bacteria			
	C collierbacteria	Candidatus Collierbacteria bacterium	KKT34677.1
	Rickettsiales	Rickettsiales bacterium	MBO87943.1
	cd WWE3	Candidate division WWE3 bacterium	OGC46700.1
	Prevotella sp	Prevotella sp.	EID32542.1
	Curtobacterium sp	Curtobacterium sp.	PZF21459.1
	A illinoisensis	Alkanindiges illinoisensis	TEU24735.1
	Clavibacter sp	Clavibacter sp.	RIJ49579.1
	C Uhrbacteria	Candidatus Uhrbacteria bacterium	PIQ67211.1
	C Pacebacteria	Candidatus Pacebacteria bacterium	PIR60552.1
	Leucobacter sp	Leucobacter sp.	RRD35472.1
	B ovatus	Bacteroides ovatus	CDB60500.1
	Hyphomonas sp	Hyphomonas sp. Mor2	WP_070961327.1
	C Zambryskibacteria	Candidatus Zambryskibacteria bacterium	OHB16576.1
	Flavobacteriaceae	Flavobacteriaceae bacterium	QCX39293.1
	Parabacteroides	Parabacteroides	WP_075965667.1
	C Falkowbacteria	Candidatus Falkowbacteria bacterium	PKM88561.1
	Aalborg AAW1	Candidate division SR1 bacterium Aalborg AAW-1	AKH32407.1
	P faecalis	Pseudoclavibacter faecalis	WP_019619849.1
	C Moranbacteria	Candidatus Moranbacteria bacterium	PID52462.1
	Robiginitomaculum sp	Robiginitomaculum sp.	PHS28547.1
		Candidatus Levybacteria bacterium	MBP6882245.1
		Candidatus Buchananbacteria bacterium	MBD3359246.1
		Candidatus Magasanikbacteria bacterium RIFOXYA2	OGH84178.1
		Chloroflexi bacterium	MBI2830749.1
		Psychrobacter sp. FDAARGOS 221	WP_096064617.1
		Enhydrobacter sp. H5	ONG38169.1
		Bifidobacterium merycicum	WP_033523136.1
		Rickettsiales bacterium	MBL6664806.1
		Proteobacteria bacterium	NBR95534.1
		Sphingobacteriia bacterium	MBN8828841.1
		Candidatus Gastranaerophilales bacterium	MBQ7287145.1
		Cyanobacteria bacterium SIG32	MBE7709962.1
		Mycoplasma sp.	MBQ6280177.1
		Acidobacteria bacterium	NMD11668.1
		Brevinematales bacterium	NPV00061.1
		Spirochaetes bacterium	HHG53312.1
		Candidatus Collierbacteria bacterium CG10	PIR99148.1
		Thermogngerobaculia bacterium	MBP7674859.1
		Henriciella sp.	NOY15510.1
		Spirochaetales bacterium	MBT3274541.1
		SAR86 cluster bacterium	MBI 6903300 1
		Clostridia bacterium	MBR5312660 1
		Spirochaetaceae bacterium	MB07366747 1
Archaea		Sprochuetaceae Bacteriani	WDQ/300/4/.1
Archite	uncult archaeon	uncultured archaeon	VVB99669.1
	C Micrarchaeota	Candidatus Micrarchaeota archaeon	01026558.1
	C Aenigmarchaeota	Candidatus Aeniamarchaeota archaeon	OIN88664 1
	archaeon CG07	archaeon CG07	PILI63205 1
	C Pacearchaeota	Candidatus Pacearchaeota archaeon	06122063.1
	M mazei	Methanosarcina mazei	TAH75514 1
	Thermonlasmata	Thermonlasmata archaeon	RIE60972 1
	mernoplasmata	Nitrosarchaeum sp	MR\$3022031 1
		Mathanosarsinalas arshanon	NKO29702 1
		uncultured archaeon	VV/P7/200 1
		Candidatus Weesearchaeota archaeon	MDIE066474 1
		Candidatus Wetsenemethylaphilasaga archaoon	MDD2/10/92 1
		Nanoarchaeota archaeon	MPU/060076 1
Fukarvota		Hundarendeota arenaeon	11004003370.1
Lukaryola	S cerevisiae	Saccharomyces cerevisiae	NP 013650 1
	H opuntiae1	Hanseniaspora opuntiae	OEJ88177 1
	H opuntiae?	Hanseniaspora opuntiae	OF 188178 1
	T nhaffii	Tetranisisnora nhaffii	XP 003684282 1
	Cviswapathii	Candida viswanathii	RCK63232 1
	C viswaridumi	Duricularia arisea	VD 02007774E 1
	P griseal	Purioularia arisan	XP_0309///45.1
	P grisea2	Pyricularia grisea	AP_030984166.1
	H sapiens	nomo sapiens	INP_079325.2

D discoideum A subglobosum A castellanii E dispar P fungivorum C fasciculata H album T socialis T socialis2 M conductrix Helicosporidium sp C sorokiniana M polymorpha M polymorpha2 Blastocystis sp Blastocystis sp2 P multistriata P multistriata2 C roenbergensis N gaditana P oligandrum P oligandrum2 B saltans L braziliensis T grayi L seymouri Phytomonas sp Perkinsela sp A deanei S culicis G muris

Dictyostelium discoideum Acytostelium subglobosum Acanthamoeba castellanii Entamoeba dispar Planoprotostelium fungivorum Cavenderia fasciculata Heterostelium album Tetrabaena socialis Tetrabaena socialis Micractinium conductrix Helicosporidium sp. Chlorella sorokiniana Marchantia polymorpha Marchantia polymorpha Blastocystis sp. Blastocystis sp. Pseudo-nitzschia multistriata Pseudo-nitzschia multistriata Cafeteria roenbergensis Nannochloropsis gaditana Pythium oligandrum Pythium oligandrum Bodo saltans Leishmania braziliensis Trypanosoma grayi Leptomonas seymouri Phytomonas sp. Perkinsela sp. Angomonas deanei Strigomonas culicis Giardia muris Polysphondylium violaceum Tieghemostelium lacteum Acytostelium subglobosum LB1 Trichogramma pretiosum Nicrophorus vespilloides Danio rerio Crassostrea gigas Dendronephthya aiaantea Actinia tenebrosa Amphimedon queenslandica Caenorhabditis elegans Salpingoeca rosetta Quercus suber Pyrus x bretschneideri Hydra vulgaris Perkinsus olseni Rhododendron griersonianum Prunus persica Nicotiana tabacum Manihot esculenta Lupinus anaustifolius Arachis hypogaea Rhodamnia argentea Papaver somniferum Kingdonia uniflora Thalictrum thalictroides Nelumbo nucifera Colocasia esculenta Cinnamomum micranthum f. kanehirae Spinacia oleracea Marchantia paleacea Ceratodon purpureus Physcomitrium patens Selaginella moellendorffii Selaginella moellendorffii Entamoeba invadens IP1 Symbiodinium microadriaticum Trypanosoma brucei equiperdum

XP_642006.1 XP_012757294.1 XP_004352499.1 XP_001738818.1 PRP79697.1 XP_004367121.1 XP_020433530.1 PNH12573.1 PNH01360.1 PSC73053.1 KDD76138.1 PRW33669.1 OAE29545.1 OAE19993.1 OA012860.1 OAO14610.1 VEU33680.1 VEU34803.1 KAA0155673.1 EWM28750.1 TMW55246.1 TMW67775.1 CUE63209.1 XP_001562602.1 XP_009312949.1 KPI83497.1 CCW70641.1 KNH07790.1 EPY29325.1 EPY23385.1 TNJ28558.1 KAF2073656.1 KYQ93685.1 XP_012754920.1 XP_014228054.1 XP_017781591.1 NP_942102.1 XP_034314500.1 XP 028415325.1 XP_031556309.1 XP_003388034.1 NP_001293174.1 XP_004991536.1 XP_023909855.1 XP_009351018.1 XP_002163633.2 KAF4753487.1 KAG5544865.1 XP 020415763.1 XP 016507676.1 XP 021598660.1 XP 019426349.1 XP_029145904.1 XP_030518540.1 XP_026396572.1 KAF6167112.1 KAF5202456.1 XP_010275116.1 MQL92731.1 RWR91934.1 XP_021855182.1 KAG6555887.1 KAG0621209.1 XP_024357988.1 XP_002987435.1 XP 024538624.1 XP_004258641.1 CAE7678393.1 RHW71036.1

		Trypanosoma rangeli	XP_029239885.1
		Marchantia paleacea	KAG6541057.1
		Ceratodon purpureus	KAG0609116.1
		Aphanomyces astaci	XP 009828150.1
		Naealeria fowleri	KAF0979914 1
		Chondrus crispus	YP 005717394 1
		Cronilario crispus	NP_003717394.1
		Gracilariopsis choraa	PXF40737.1
		Giardia intestinalis ATCC 50581	EET02286.1
		Leishmania martiniquensis	KAG5479457.1
		Bodo saltans	CUF06097.1
		Porphyra umbilicalis	OSX74557.1
		Porphyra umbilicalis	OSX70336.1
TraA (plasmids)			
india (plusinius)	Methanothrix sn	Methanothrix sp	TEH49976 1
	Dhastarium	Desulfabasterasaga bastorium	PDI72509 1
	Disacterium	Desulforderer deede bacterium	NFT/ 5556.1
	Dicetonica	Desuljosarcina cetonica	WP_054694575.1
	Sphingobium sp	Sphingobium sp. B2	WP_145206887.1
	Sphingomonas sp	Sphingomonas sp. AAP5	WP_133192514.1
	Phenylobacterium sp	Phenylobacterium sp. CCH9-H3	WP_068876894.1
	S macrogoltabida	Sphingopyxis macrogoltabida	WP_054590692.1
	A tumefaciens	Agrobacterium tumefaciens	AYM81042.1
	Mesorhizohium sn	Mesorhizohium sp. B4-1-1	WP 140901472 1
	A ovcontrieve	Acticcacquilis avcontricus	W/D 012470070 4
1	A excentricus	Asticuluuis excentricus	WP_013478970.1
ukaryotic viruses			
		Emiliania huxleyi virus 99B1	CAZ69470.1
		Marseillevirus LCMAC101	QBK85639.1
		Marseillevirus LCMAC102	QBK86258.1
		Marseillevirus LCMAC103	QBK87070.1
		Sicyoidochytrium minutum DNA virus	BCU09408.1
		Organic Lake phycodnavirus 1	ADX05998 1
		Organic Lake phycodiawirus 2	ADX053330.1
		Organic Lake phycodnavirus 2	ADX00411.1
		Chrysochromulina ericina virus	YP_009173733.1
		Phaeocystis globosa virus	YP_008052747.1
		uncultured Mediterranean phage	ANS04235.1
		Virus NIOZUU159	QPI16828.1
		Invertebrate iridescent virus 22	YP 009010863.1
		Invertebrate iridescent virus Kaz2018	ONH08436 1
		Armadillidium uulgare irideseent virus	VD 000046911 1
		Armadilidium vulgare indescent virus	1P_009046811.1
		Hydra MELD virus	DAC81588.1
		Mimivirus AB566017	ARR75030.1
		Mollivirus kamchatka	QHN71346.1
		Mollivirus sibericum	YP_009165351.1
		Erinnyis ello granulovirus	ARX71979.1
		Clostera anastomosis granulovirus B	YP 009506054 1
		Charistoneura fumiferana granulavirus	VD 654536 1
		Chonstoneura runnierana granulovirus	IF_054520.1
		Phthorimaea operculella granulovirus	NP_663278.1
		Cydia pomonella granulovirus	AIU36910.1
		Pieris rapae granulovirus	ADO85536.1
		Cryptophlebia leucotreta granulovirus	NP_891963.1
		Matsumuraeses phaseoli granulovirus	QOD40078.1
		Diatraea saccharalis granulovirus	YP 009182312.1
		I vmantria xvlina nucleopolyhedrovirus	YP 003517787 1
		Oravia pseudotsugata puslopolubodravirus	01/071652 1
		Orgyna pseudotsugata nuclopolyneurovirus	QW0/1053.1
		Orgyia leucostigma nucleopolyhedrovirus	YP_001651017.1
		Epinotia aporema granulovirus	YP_006908627.1
		Agrotis segetum granulovirus	YP_009513161.1
		Spodoptera litura granulovirus	YP_001257069.1
		Spodoptera frugiperda granulovirus	AXS01146.1
		Mocis latipes granulovirus	YP 009249960.1
		Xestia chigrum granulovirus	NP 059294 1
		Mamostra configurata pueleanalukedraujeus D	ONU00674.1
		iviamestra configurata nucleopolyhedrovirus B	QNH90674.1
		Plutella xylostella granulovirus	QKV50030.1
		Sucra jujuba nucleopolyhedrovirus	YP_009186748.1
		Hyposidra talaca NPV	YP_010086327.1
		Lambdina fiscellaria nucleopolyhedrovirus	YP_009133285.1
		Peridroma alphabaculovirus	YP 009049868 1
		Charistoneura hiennis entomonovuirus	VP_002004207.1
		Trichonhusia ai associate 2-	VD 000004527.1
		i richopiusia ni ascovirus 2č	YP_803305.1
		Heliothis virescens ascovirus 3h	AYD68236.1

	Pandoravirus salinus	YP_008437119.1
	Acanthamoeba castellanii medusavirus	BBI30459.1
	Sylvanvirus sp.	AYV86632.1
Prokaryotic viruses		
	Acinetobacter phage vB AbaM ME3	YP_009595951.1
	Podoviridae sp.	DAJ82417.1
	Prokaryotic dsDNA virus sp.	QDP67633.1
	Microbacterium phage PauloDiaboli	QIG57888.1
	Myoviridae sp.	DAU76505.1
	Bacteriophage sp.	AFB75491.1
	Siphoviridae sp. ctAUQ2	DAD87486.1
	Siphoviridae sp.	DAO03073.1
	Podoviridae sp.	DAQ71114.1
	Podoviridae sp. ctfN46	DAJ22427.1
	Bacteriophage sp.	DAL07837.1
	Bacteriophage sp.	DAY30538.1
	uncultured Caudovirales phage	CAB4198187.1
	Myoviridae sp.	DAM57115.1
	Escherichia phage FV3	YP_007006388.1
	Escherichia phage LL12	AXC42890.1
	Erwinia phage pEp SNUABM 01	YP_009851551.1
	Erwinia phage Hena1	YP_009854417.1
	Escherichia phage 4MG	YP_008857219.1
	Salmonella phage GEC vB MG	QPI14547.1
	Raoultella phage Ro1	YP 009835918.1
	Acinetobacter phage ABPH49	AXN57909.1
	Cronobacter phage CR8	YP_009042324.1
	Klebsiella phage vB KaeM KaOmega	QEG12160.1
	Escherichia phage UPEC06	QUL77343.1
	Pseudomonas phage pf16	YP 009595586.1
	Prokaryotic dsDNA virus sp.	QDP60500.1
	Prokaryotic dsDNA virus sp.	QDP64781.1
	Vibrio phage 1.164	AUR91792.1
	Vibrio phage 1.124	AUR89562.1
	Bacteriophage sp.	DAE75004.1
	Siphoviridae sp. ctgK313	DAF60248.1
	Bacteriophage sp.	DAP73423.1
	Siphoviridae sp. ctgBH20	DAE16492.1
	Ackermannviridae sp.	DAG97916.1
	Myoviridae sp.	DAX71650.1
	Klebsiella phage AmPh EK29	QFR57062.1
	Enterobacter phage mvPSH1140	YP 010093920.1
	Edwardsiella phage PEi20	YP 009190175.1
	Shigella phage SP18	YP_003934641.1
	Myoviridae sp. ctCo31	DAF95488.1
	Vibrio phage VH7D	VP_009006117.1

*Sequences with abbreviated names were selected in the first round of the analysis. The ones without abbreviation were selected afterwards, without filtering taxa with *Helitrons* in their genomes (see text).

Species	Accession	Classification	Hit from Blastp
Oryza sativa	ABB47755	RepHel protein	-
Arabidopsis thaliana	CAB91581	RepHel protein	-
	NP_190738	79% cover, 59.50% identity to RepHel	RIA05759.1
	CAB63155	67% cover, 49.65% identity to RepHel	XP_018453621.1

Supplementary Table S3

Accession	Identity	Cover	E-value	Classification	Hit from 2 nd Blastp
*XP_007819664.1	41.68%	98%	7.00E-106	No significant identity to RepHel	
*XP_007816514.1	29.64%	92%	9.00E-49	No significant identity to RepHel	
*XP_007826535.2	28.97%	92%	3.00E-26	RepHel	-
*XP_007816691.2	26.38%	88%	6.00E-26	RepHel	
*XP_007825309.2	28.26%	87%	6.00E-20	RepHel	
*XP_007825293.2	28.42%	37%	3.00E-17	No significant identity to RepHel	
*XP_007816587.2	26.48%	87%	1.00E-16	RepHel	
*XP_007817134.1	29.06%	46%	5.00E-16	98% cover, 97.90% identity to RepHel (Best hit)	EXU95784.1
*XP_011411820.1	31.74%	38%	2.00E-12	95% cover, 74.31% identity to RepHel (second best hit)	KJK85320.1
*XP_007816591.2	29.35%	42%	4.00E-11	RepHel (cryptic)	
*XP_007826337.2	31.29%	31%	7.00E-11	RepHel	
*XP_007816573.2	30.07%	30%	3.00E-10	RepHel	
*XP_007816551.2	30.07%	30%	5.00E-10	RepHel	
*XP_007826745.1	29.79%	33%	2.00E-09	100% cover, 76.55% identity to RepHel (Best hit)	EXU95911.1
*XP_007817473.2	30.54%	35%	1.00E-08	97% cover, 78.37% identity to RepHel (second best hit)	XP_007816587.2
*XP_007817117.2	22.46%	66%	3.00E-08	RepHel	-
*XP_007826647.1	22.75%	47%	3.00E-08	100% cover, 85.49% identity to RepHel (second best hit)	KJK73666.1
*XP_007825291.2	31.62%	40%	2.00E-07	No significant identity to RepHel	
*XP_007817091.1	26.67%	66%	3.00E-07	RepHel (cryptic)	
*XP_011411726.1	26.70%	44%	4.00E-05	68% cover, 96.19% identity to RepHel (Best hit)	EXU95304.1
*XP_007826148.1	40.00%	16%	1.00E-04	78% cover, 89.06% identity to RepHel	XP_007816591.2
*XP_007817793.2	30.66%	30%	2.00E-04	46% cover, 62.30% identity to RepHel (second best hit)	EXU94892.1
*XP_007826598.2	43.18%	10%	0.02	Not Pif1 helicase	-
**XP_007826758.1	35.00%	8%	5.00E-05	89% cover, 76.07% identity to RepHel (Best hit)	KID81362.1
**XP_007816555.1	31.96%	10%	4.00E-04	RepHel	

*Result of Blastp search using the human Pif1 domain (accession 6HPH_A) as a query against Metarhizium robertsii ARSEF 23.

**Result of Blastp search using the yeast Pif1 (accession NP_013650.1) as a query against *Metarhizium robertsii* ARSEF 23 (only hits that did not overlap with the ones from the search using the human Pif1 domain).

Accession*	Identity*	Cover*	E-value*	Classification	Hit from 2 nd Blastp
*AAG52281.1	27.23%	91%	2.00E-27	RepHel	-
*AAM15154.1	26.46%	93%	4.00E-25	RepHel	
*AAD25596.1	27.19%	93%	7.00E-24	RepHel	
*BAB02793.1	25.39%	90%	3.00E-21	RepHel	
*CAB91581.1	24.72%	90%	3.00E-21	RepHel	
*AAD32757.1	25.92%	93%	1.00E-20	RepHel	
*AAG51081.1	25.73%	94%	4.00E-20	RepHel	
*BAB01023.1	26.91%	79%	6.00E-20	RepHel	
*AAD15468.1	24.36%	94%	2.00E-19	RepHel	
*AAG52315.1	25.73%	90%	2.00E-19	RepHel (cryptic)	
*BAB11364.1	27.73%	86%	4.00E-19	RepHel	
*CAB81576.1	24.71%	89%	2.00E-16	RepHel (cryptic)	
*AAC28215.1	30.93%	41%	5.00E-16	93% cover, 69.83% identity to RepHel (best hit)	CAB91581.1
*AAC62789.1	24.79%	82%	8.00E-16	80% cover, 78.30% identity to RepHel (best hit)	AAD15468.1
*OAP18984.1	37.69%	29%	1.00E-12	RepHel	-
*AAD25621.1	37.50%	28%	1.00E-12	RepHel	-
*BAB02227.1	36.72%	28%	4.00E-12	RepHel (cryptic)	-
*AAD20107.1	24.52%	85%	1.00E-11	RepHel	
*CAA0384207.1	33.57%	31%	6.00E-11	99% cover, 75.32% identity to cryptic RepHel (second best hit)	XP_010421223.1
*AAD15325.1	25.57%	79%	9.00E-10	RepHel	-
*AAG51717.1	30.06%	38%	7.00E-09	RepHel	-
*OAP08664.1	28.14%	57%	1.00E-07	RepHel (cryptic)	-
*NP_190738.1	30.93%	40%	2.00E-07	85% cover 44.56% identity to RepHel (best hit outside Brassicaceae)	XP_030934889.1
*CAA0385759.1	30.93%	40%	2.00E-07	85% cover 44.56% identity to RepHel (best hit outside Brassicaceae)	XP_030934889.1
*VYS60096.1	30.93%	40%	2.00E-07	85% cover 44.56% identity to RepHel (best hit outside Brassicaceae)	XP_030934889.1
*AAF06079.1	32.79%	27%	1.00E-06	RepHel (cryptic)	-
** Same hits	-	-	-	-	

* Result of Blastp search using the human Pif1 domain (accession 6HPH_A) as a query against Arabidopsis thaliana.

** Result of Blastp search using the yeast Pif1 (accession NP_013650.1) as a query against Arabidopsis thaliana (only hits that did not overlap with the ones from the search using the human Pif1 domain).

Accession	Identity	Cover	E-value	Classification	Hit from 2 nd Blastp
*AAK54302.1	27.39%	93%	5.00E-26	RepHel	
*ABF97674.1	26.19%	93%	2.00E-25	RepHel	-
*XP_025876548.1	26.17%	93%	3.00E-25	RepHel (cryptic)	-
*AAP52492.2	27.25%	91%	8.00E-25	RepHel	
*AAM92800.1	27.25%	91%	8.00E-25	RepHel	
*XP_015613561.1	27.25%	91%	8.00E-25	RepHel	
*AAN09850.1	27.25%	91%	1.00E-24	RepHel	
*AAP52578.2	27.25%	91%	1.00E-24	RepHel	
*XP_015613597.1	27.25%	91%	1.00E-24	RepHel	
*XP_025879680.1	26.79%	92%	1.00E-24	RepHel (cryptic)	
*BAF26194.2	27.25%	91%	1.00E-24	RepHel	-
*BAH91204.1	26.37%	92%	1.00E-22	RepHel	
*EEC77075.1	25.93%	91%	2.00E-22	RepHel (cryptic)	
*XP_015624412.1	25.44%	92%	3.00E-22	RepHel (cryptic)	
*BAD81603.1	25.89%	88%	5.00E-22	RepHel	
*BAF04484.1	25.89%	88%	6.00E-22	RepHel	
*BAH93748.1	27.51%	93%	3.00E-21	RepHel	-
*XP_025879790.1	25.96%	93%	7.00E-21	RepHel (cryptic)	
*AAO34493.1	26.45%	93%	2.00E-20	RepHel	
*BAF08763.2	24.59%	89%	2.00E-19	RepHel	
*AAX95750.1	25.32%	93%	3.00E-19	RepHel	-
*XP_025879706.1	25.24%	93%	2.00E-18	RepHel (cryptic)	
*XP_015621010.1	26.68%	84%	3.00E-18	RepHel (cryptic)	
*BAH92578.1	25.11%	93%	3.00E-18	RepHel	-
*CAD40309.2	25.11%	93%	3.00E-18	RepHel	-
*XP_025878111.1	26.52%	69%	5.00E-18	100% cover, 59.24% identity to RepHel (best hit)	BAD81603.1
*AAP54489.2	29.12%	62%	2.00E-17	100% cover, 90.36% identity to RepHel (best hit)	BAF04484.1
*BAC55632.1	26.88%	69%	4.00E-17	RepHel	-
*AAM93454.1	26.52%	66%	2.00E-16	100% cover, 100% identity to RepHel (best hit)	AAM92800.1
*BAH93891.1	36.31%	36%	5.00E-16	RepHel	-
*ABA99439.1	24.08%	81%	6.00E-16	99% cover, 70.98% identity to cryptic RepHel (best hit)	EEC77075.1
*ABA95256.2	25.32%	81%	7.00E-16	RepHel	-
*XP_025878227.1	38.46%	32%	2.00E-15	99% cover, 81.39% identity to cryptic RepHel (best hit)	XP_015637912.1
*XP_015620800.1	39.31%	74%	2.00E-15	RepHel (cryptic)	-
*CAE76063.1	34.93%	32%	2.00E-15	RepHel (cryptic)	-
*CAE76056.1	34.93%	32%	3.00E-15	RepHel	-
*XP_015637912.1	38.19%	68%	8.00E-15	RepHel (cryptic)	
*BAC84865.1	37.50%	68%	1.00E-14	RepHel	
*BAF22399.2	37.50%	68%	2.00E-14	RepHel	-
*BAD01692.1	37.50%	68%	2.00E-14	RepHel	-
*CAH66128.1	37.50%	68%	2.00E-14	RepHel	•
*AAX95983.1	37.50%	68%	2.00E-14	RepHel	
*AAU44208.1	35.66%	67%	3.00E-14	RepHel	-
*ABA94634.1	35.66%	67%	3.00E-14	RepHel	-

*ABA94947.1	35.66%	67%	3.00E-14	RepHel	-
*BBD82308.1	35.66%	67%	3.00E-14	RepHel	-
*ABA95236.1	35.66%	67%	3.00E-14	RepHel	-
*AAT85173.1	34.93%	74%	3.00E-14	RepHel (cryptic)	
*AAV44035.1	34.97%	67%	7.00E-14	RepHel	
*AAK13103.1	26.43%	71%	9.00E-14	RepHel	-
*BAS88751.1	24.79%	71%	1.00E-13	RepHel (cryptic)	-
*XP_015627019.1	23.96%	69%	1.00E-13	RepHel (cryptic)	-
*BAF14458.1	24.79%	71%	1.00E-13	RepHel (cryptic)	-
*XP_025880731.1	24.58%	72%	2.00E-13	RepHel	-
*ABA94881.2	37.06%	68%	2.00E-13	RepHel	-
*AAK54292.1	33.95%	35%	2.00E-13	RepHel	-
*ABB47755.2	33.95%	35%	2.00E-13	RepHel	-
*KAB8095338.1	24.79%	71%	2.00E-13	RepHel (cryptic)	-
*EEC77085.1	24.79%	71%	3.00E-13	RepHel	-
*BAH94916.1	33.95%	35%	3.00E-13	RepHel (cryptic)	-
*CAD40616.1	24.79%	71%	4.00E-13	RepHel	
*BAD68127.1	31.21%	32%	3.00E-12	RepHel	
*EEC82986.1	32.81%	29%	1.00E-11	RepHel	-
*BAF04591.1	34.42%	32%	8.00E-11	RepHel (cryptic)	-
*ABA93595.1	37.80%	28%	1.00E-10	RepHel (cryptic)	-
*BAF29741.2	35.38%	65%	1.00E-09	RepHel (cryptic)	-
*BAG93269.1	27.44%	48%	2.00E-09	RepHel (cryptic)	-
*BAG93269.1 *XP_025880729.1	27.44% 24.93%	48% 67%	2.00E-09 2.00E-09	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit)	- BAH91022.1
*BAG93269.1 *XP_025880729.1 *BAD68018.1	27.44% 24.93% 27.44%	48% 67% 48%	2.00E-09 2.00E-09 2.00E-09	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit)	- BAH91022.1 BAG93269.1
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1	27.44% 24.93% 27.44% 35.38%	48% 67% 48% 65%	2.00E-09 2.00E-09 2.00E-09 2.00E-09	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic)	- BAH91022.1 BAG93269.1 -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2	27.44% 24.93% 27.44% 35.38% 31.88%	48% 67% 48% 65% 35%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2	27.44% 24.93% 27.44% 35.38% 31.88% 50.88%	48% 67% 48% 65% 35% 13%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel RepHel (cryptic)	- BAH91022.1 BAG93269.1 - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86%	48% 67% 48% 65% 35% 13% 25%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel RepHel	- BAH91022.1 BAG93269.1 - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94%	48% 67% 48% 65% 35% 13% 25% 46%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel RepHel (cryptic)	- BAH91022.1 BAG93269.1 - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03%	48% 67% 48% 65% 35% 13% 25% 46% 35%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 31.43%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-07 3.00E-07	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel RepHel (cryptic) RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 31.43% 28.03%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35%	2.00E-09 2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-07 3.00E-07 3.00E-07	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 31.43% 28.03% 28.03%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 35%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-07 3.00E-07 3.00E-07 5.00E-07	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - -
*BAG93259.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *BAH91022.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 31.43% 28.03% 28.03% 28.03% 24.44%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 35% 67%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 5.00E-07 3.00E-07	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH94330.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 28.03% 28.03% 24.44% 25.77%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 35% 67% 52%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 5.00E-07 3.00E-07 3.00E-06 4.00E-06	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 67% 52% 34%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 4.00E-06 6.00E-06	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 52% 34% 31%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 4.00E-06 6.00E-06 0.016	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 35% 67% 52% 34% 31% 56%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 4.00E-06 6.00E-06 0.016 0.02	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel (cryptic) RepHel RepHel (cryptic)	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AT85232.1 *XP_015650422.1 *BAH91022.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1 *AAQ56555.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11% 41.51%	48% 67% 48% 65% 35% 25% 46% 35% 35% 35% 67% 52% 34% 31% 56% 9%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 6.00E-06 6.00E-06 0.016 0.02 0.036	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *AAT85232.1 *AT85232.1 *AF2015650422.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1 *AAQ56555.1 *AAL75753.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11% 41.51% 27.42%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 67% 52% 34% 31% 56% 9% 24%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 0.016 0.02 0.036 0.038	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1 *AAQ56555.1 *AAQ56555.1 *AAL75753.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11% 41.51% 27.42% 26.47%	48% 67% 48% 65% 35% 13% 25% 46% 35% 23% 35% 67% 52% 34% 31% 56% 9% 24%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 6.00E-06 0.016 0.02 0.036 0.038 2.00E-06	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *EEE67922.1 *BAH91022.1 *BAH91022.1 *BAH94330.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1 *AAQ56555.1 *AAQ56555.1 *AAQ56555.1 *AAQ56575.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11% 41.51% 23.11% 41.51% 27.42%	48% 67% 48% 65% 35% 35% 46% 35% 23% 35% 67% 52% 34% 31% 56% 9% 24% 33%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 4.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 6.00E-06 0.038 2.00E-06 3.00E-05	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -
*BAG93269.1 *XP_025880729.1 *BAD68018.1 *ABA98117.1 *BAF08718.2 *ABA99343.2 *BAF24192.1 *ABA95557.1 *BAH94086.1 *AAT85232.1 *XP_015650422.1 *BAH94030.1 *XP_025880332.1 *ABA96519.2 *BAF05239.1 *AAQ56555.1 *AAQ56555.1 *AAQ56555.1 *AAQ5677503.1 *XP_025877503.1 *XP_025877503.1 *ABA97607.1 *KAB8082674.1	27.44% 24.93% 27.44% 35.38% 31.88% 50.88% 31.86% 25.94% 28.03% 28.03% 28.03% 24.44% 25.77% 27.50% 28.17% 23.11% 41.51% 27.42% 26.47% 24.17% 27.42%	48% 67% 48% 65% 35% 35% 46% 35% 23% 35% 67% 52% 34% 31% 56% 9% 24% 33% 17%	2.00E-09 2.00E-09 2.00E-09 1.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-08 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-07 3.00E-06 6.00E-06 0.016 0.02 0.036 0.038 2.00E-06 3.00E-05 1.00E-04	RepHel (cryptic) 96% cover, 97.98% identity to RepHel (best hit) 100% cover, 100% identity to RepHel (best hit) RepHel (cryptic) RepHel (cryptic) RepHel (cryptic) RepHel	- BAH91022.1 BAG93269.1 - - - - - - - - - - - - - - - - - - -

* Result of Blastp search using the human Pif1 domain (accession 6HPH_A) as a query against Oryza sativa.

** Result of Blastp search using the yeast Pif1 (accession NP_013650.1) as a query against Oryza sativa (only hits that did not overlap with the ones from the search using the human Pif1 domain).

Supplementary Table S6

Group*		Species	Accession	Identity	Cover	E-value	Classification	Hit from 2 nd Blastp
Brassicales	•	Brassica napus	XP_022547407.1	26.84%	93%	8.00E-33	99% cover, 88.42% identity to RepHel	KAF8111651.1
	*	Camelina sativa	XP_010436751.1	27.27%	94%	1.00E-31	97% cover, 58.80% identity to RepHel	RID40682.1
	*	Brassica napus	XP_022551638.1	26.88%	93%	1.00E-30	RepHel	-
	٠	Brassica rapa	XP_033148559.1	26.88%	93%	1.00E-30	RepHel	-
	*	Brassica napus	XP_013725746.1	26.88%	93%	1.00E-30	RepHel	-
	*	Brassica napus	XP_013719709.1	26.88%	93%	1.00E-30	RepHel	-
	*	Raphanus sativus	XP_018453621.1	26.67%	93%	5.00E-30	RepHel	-
	*	Brassica rapa	XP_033143195.1	27.90%	94%	9.00E-30	RepHel	
	*	Arabidopsis thaliana x	KAG7586339.1	26.92%	94%	9.00E-30	RepHel	-
	٠	Eutrema salsugineum	XP_024013997.1	27.10%	93%	1.00E-29	RepHel	
	**	Brassica napus	CAF2097984.1	26.52%	40%	3.00E-12	RepHel	
	**	Microthlaspi erraticum	CAA7047626.1	26.77%	42%	1.00E-11	RepHel	
	**	Microthlaspi erraticum	CAA7039386.1	26.24%	44%	1.00E-11	RepHel (cryptic)	
	**	Microthlaspi erraticum	CAA7015018.1	26.77%	35%	3.00E-11	RepHel (cryptic)	
	**	Brassica napus	XP_022544095.1	25.07%	35%	6.00E-11	RepHel	
	**	Raphanus sativus	XP_018460436.1	26.18%	33%	8.00E-11	RepHel (cryptic)	
	**	Brassica napus	XP_013694041.1	25.76%	33%	1.00E-10	RepHel	
	**	Brassica rapa	RID62868.1	25.93%	40%	1.00E-10	RepHel	
	**	Brassica napus	XP_022564371.1	26.10%	33%	1.00E-10	RepHel	
	**	Brassica napus	XP_013694540.1	27.87%	33%	1.00E-10	100% cover 95.47% identity to RepHel	XP_022548462.1
Commelinids	*	Zea mays	ONM60906.1	29.15%	92%	4.00E-28	RepHel	
	*	Zea mays	ONM39160.1	28.05%	92%	2.00E-27	RepHel	
	٠	Sorghum bicolor	XP_002446095.2	27.16%	93%	1.00E-26	RepHel	
	*	Musa acuminata	ABF70031.1	26.94%	93%	2.00E-26	RepHel	-
	*	Zea mays	AQK52428.1	28.33%	92%	2.00E-26	RepHel	
	٠	Zea mays	AQK60686.1	27.53%	92%	2.00E-26	RepHel	
	*	Zea mays	PWZ05004.1	26.82%	95%	3.00E-26	RepHel	
	*	Sorghum bicolor	XP_021314672.1	26.94%	93%	3.00E-26	RepHel	
	٠	Zea mays	PWZ25377.1	28.33%	92%	3.00E-26	RepHel	-
	*	Zea mays	ONM39853.1	27.53%	92%	5.00E-26	RepHel	
	**	Zea mays	AQK64577.1	26.61%	35%	1.00E-13	RepHel	-
	**	Oryza sativa Japonica	XP_025876548.1	26.65%	33%	8.00E-14	RepHel (cryptic)	-
	**	Zea mays	AQK84207.1	26.61%	35%	2.00E-13	RepHel	
	**	Zea mays	PWZ13396.1	27.27%	35%	2.00E-13	RepHel	-
	**	Zea mays	PWZ06906.1	25.84%	35%	2.00E-13	RepHel	
	**	Zea mays	AQK97791.1	26.33%	35%	3.00E-13	RepHel	-
	**	Zea mays	ONM55810.1	26.61%	35%	4.00E-13	RepHel	
	**	Zea mays	PWZ04632.1	26.99%	35%	5.00E-13	RepHel	-
	**	Zea mays	PWZ11828.1	26.61%	35%	8.00E-13	RepHel	-
)	**	Oryza sativa Japonica Group	AAK13103.1	27.24%	33%	8.00E-13	RepHel	

Malvids	*	Theobroma cacao	EOX92974.1	43.82%	98%	2.00E-102	No significant identity to RepHel	-
	*	Theobroma cacao	XP_017972716.1	43.59%	98%	1.00E-100	No significant identity to RepHel	-
		Durio zibethinus	XP_022774647.1	43.09%	98%	1.00E-99	No significant identity to RepHel	
	*	Herrania umbratica	XP_021274232.1	42.99%	98%	2.00E-99	No significant identity to RepHel	
	*	Corchorus olitorius	OMO60853.1	42.12%	97%	7.00E-94	No significant identity to RepHel	-
	٠	Punica granatum	XP_031384248.1	40.79%	97%	1.00E-92	No significant identity to RepHel	-
	*	Rhodamnia argentea	XP_030518540.1	41.07%	98%	2.00E-92	No significant identity to RepHel	
	*	Eucalyptus grandis	XP_010035891.2	40.75%	98%	3.00E-92	No significant identity to RepHel	-
	٠	Rhodamnia argentea	XP_030518284.1	40.93%	98%	9.00E-92	No significant identity to RepHel	
	*	Punica granatum	PKI33626.1	40.56%	97%	1.00E-91	No significant identity to RepHel	
	**	Corchorus capsularis	OM061479.1	40.06%	52%	7.00E-66	No significant identity to RepHel	-
Liliopsida	*	Colocasia esculenta	MQL92731.1	41.90%	96%	1.00E-89	No significant identity to RepHel	-
	٠	Asparagus officinalis	ONK72744.1	36.41%	92%	1.00E-74	No significant identity to RepHel	
	*	Asparagus officinalis	XP_020262994.1	37.87%	83%	2.00E-73	No significant identity to RepHel	
	*	Zostera marina	KMZ75646.1	34.50%	70%	2.00E-44	No significant identity to RepHel	
	*	Zostera marina	KMZ70362.1	35.32%	59%	4.00E-39	No significant identity to RepHel	
	*	Zostera marina	KMZ65819.1	36.67%	55%	1.00E-36	No significant identity to RepHel	
	*	Zostera marina	KMZ67804.1	43.62%	33%	3.00E-32	No significant identity to RepHel	
	٠	Zostera marina	KMZ56065.1	36.00%	39%	2.00E-30	No significant identity to RepHel	
	•	Zostera marina	KMZ65715.1	32.45%	60%	1.00E-29	No significant identity to RepHel	-
	*	Zostera marina	KMZ68271.1	43.80%	31%	4.00E-28	No significant identity to RepHel	-
	**	Same hits				-	-	

* Results of Blastp searches using the human Pif1 domain (accession 6HPH_A) as a query against the corresponding group. The 10 best hits from each search are shown.

** Results of Blastp searches using the yeast Pif1 (accession NP_013650.1) as a query against the corresponding group. The 10 best hits from each search are shown (only hits that did not overlap with the ones from searches using the human Pif1 domain).

Group	Species*	Accession*	Identity*	Cover*	E-value*	Classification	Hit from 2nd Blastp
Brassicales	Brassica oleracea	XP_013639271.1	28.21%	84%	7.00E-22	RepHel	
	Raphanus sativus	XP_018465781.1	26.99%	90%	9.00E-21	RepHel (cryptic)	-
	Brassica rapa	XP_033147243.1	26.82%	84%	3.00E-20	RepHel	
	Brassica oleracea	XP_013629542.1	26.72%	90%	2.00E-19	RepHel (cryptic)	
	Eutrema salsugineum	XP_024006484.1	25.64%	88%	7.00E-19	RepHel (cryptic)	-
	Eutrema salsugineum	XP_024007971.1	28.40%	81%	3.00E-19	100% cover, 95.18% identity to RepHel (best hit)	XP_024004792.1
	Eutrema salsugineum	XP_024014429.1	25.46%	88%	8.00E-19	RepHel (cryptic)	-
	Capsella rubella	EOA12259.1	26.56%	82%	5.00E-19	RepHel (cryptic)	-
	Capsella rubella	XP_023633617.1	26.69%	84%	2.00E-18	RepHel	-
	Capsella rubella	EOA12327.1	27.17%	84%	2.00E-18	RepHel (cryptic)	-
Commelinids	Oryza sativa	XP_025876548.1	27.31%	84%	2.00E-26	RepHel (cryptic)	-
	Oryza sativa	AAK54302.1	26.81%	82%	7.00E-25	RepHel	-
	Oryza sativa	BAH94916.1	26.38%	82%	7.00E-25	RepHel (cryptic)	-
	Sorghum bicolor	OQU91688.1	25.93%	80%	2.00E-24	RepHel	-
	Zea mays	AQK95425.1	28.15%	84%	3.00E-24	RepHel (cryptic)	-
	Triticum dicoccoides	XP_037419736.1	24.84%	82%	3.00E-24	RepHel	-
	Triticum dicoccoides	XP_037474121.1	26.05%	83%	3.00E-24	RepHel	-
	Triticum urartu	EMS67201.1	24.84%	82%	4.00E-24	RepHel	-

Supplementary Table S7

Sorghum bicolor	XP_021305262.1	27.43%	80%	2.00E-24	100% cover, 99.19% identity to cryptic RepHel	XP_002444425.2
Sorghum bicolor	XP_002452524.1	28.15%	80%	3.00E-24	(best hit) 99% cover 94.29% identity to cryptic RepHel	XP_002444425.2
					(best hit)	

* Results of Blastp using the best hits from searches in malvids (EOX92974.1) and Liliopsida (MQL92731.1) (see Table S6) as queries against Brassicales and commelinids, respectively. The 10 best hits from each search are shown.

Supplementary Data S1. Consensus sequences of the Hel and Rep domains from Helentron (including Helitron2) and Helitron variants.

>Helentron_Hel_consensus

INKEQREFFYHVLHLIKTSPEPFYLFLSGGAGVGKSHLIKALYQALLKYLNSLPGFRGPKVLLAAPTGKAAFNISGTTLHSLLKLPISQSPYKPLSASRLNTLRCKLRDLKLLIIDEI SMVGSRMFNWINNRLRDIKGSDEPFGGISIIAVGDLFQLPPVGDKPIFKDPENYILARNLWWEFFKMFELTEIMRQRDDKAFAEALNRLREGQLTDEDIKLLKQRVVTEKNRPSDALH LFATNDEVNEYLNREVLDRLKGEKIQIKAIDVVIGARTKADTRKTGGLAKLLQLAVGARYMLTRNLDVEDGLVNGAGTVK

>Helitron Hel consensus

ZINEEQRAÄDTILAAVSDGSGGLFFLDGPGGTGKTFLYKTLLAAIRSQGKIVLCVASSGIAALLLPGGRTAHSRFKLPLNLNETSVCGIKKQSKLARLLKEAKLIIWDEAPMAHKHA LEAVDRLLKDIMNNDQPFGGKVVLLGGDFRQILPVVPRGTRADIVNACLKSSYLWPNFKTLKLTKNMRVTSGEDQEFSEWLLKIGDGNLNVDGEGLIEIPEDFLIIEEIIEEIYPDII

>Helentron_Rep_consensus PTLFCTFSAAETKWPHLLKILGKLVDNKYTEDELENLDWDEKCRLIQSDPVTCARYFDKRVDALLTTLLLSPAQPFGKVVDYFYRVEFQQRGSPHIHMLLWLEDAPKFGVDSDEEVIE FIDKIITCQKPDLNELKDLVNRQTHRHTHTCKKKNKKSCRFNIPQPPMPKTMILYPLEDDDSERKELKEKWKKIKDLLNDKEGSFDTFEEFLAKLNLSEEDYLLAVRSSLKRPTVFLK RQPNELRINNYNPDILKAWRANMDIQFILDVYACAMYIVSYIS

>Helitron_Rep_consensus PDLFITFTCNPKWPEITENLLPGQTAEDRPDIVARVFKLKLKSLLNDLTKKHVLGKVRAWIYVIEFQKRGLPHAHILLILKEEDKPRTPEDIDKIISAEIPDKETDPELYEIVTSNMI HGPCGAANPSSPCMVDGKCSKRFPKKFQEETVINVDGYPLYRRRNNGRGVEKGGIELDNRWVVPYNPYLSLKYNAHINVEVCNSIKSIKYLFKYVY

Supplementary Data S2. Final alignment of Pif1-like amino acid sequences used in the phylogenetic and NMDS analyses.

>Helen_A_alb

-----A----P-G-----------L-D-C----------MS-----

>Helen_C_qui

ATNAEQRA-LILHVIHLM-HCYEEHEPLQVFLTGPAGSGKTFVLRALMETI-NRYSQ
THNSRD-NAYVASASTGKAASAIGGTTLHHA-YHITMSR-QAAKMNFETLFETL
QMYRNEMQNIKFHIIDEVSMVGAHTLNT-AHIRLQDVYM
ARAR
CGA-VVWQSL-MFHELKRVMRQ-ADK
QFSDILTKIGNGLKL-TADEDETKLI-ESRF
DLSK-EDT
AVRLFHRNIDVTSYNNEALR-NIDG
T
KKMS
AGLQYTTKFCPGMPYMVTTNVNVE-DGIVNGAIGDLMYV

>Helen G occ

----KLNAEQRE-LILEV-----IHR----L-HDP------NSEA-----IQIFLTGPAGCGKTYTLKALMET---Y-N--R--YAQ--

>LIED01008227.1 Bemisia tabaci

>Helen L ser

>Helen M sac

>Helen M cir

>Helen C cuc

_____QPFGG______RHVILFGDFLQLPPVK_____GQ_____GQ_____ --AH----E-T--

>Helen D rer

_____ _____ -MG----K-K-----DNL-----G----LDTI-----QVAVGVRIMVIRNLD-----V---E-D----G----LVNGCFGKIGNI

>Helen_L_roh

>Helen A cal

>Helen A mil

>Helen N vec

>Helen A dig

>Helen D gig

>Helen_L_ana

-----SQ-----------MNL-----G----ITNGAPNVVKLV

>Helen O fav

-----SLNEKQRQ-FFYHV-----LHS----I-KTRD------DP-----DP----LRLFLSGGAGVGKSTVTNALYEA---L-I-R--YLNSIA---------MGL-----YSLV-----SLATMAKYDLTTNID-----V----T--D----G-----LTNGAECMIENI

>Helen C aia

>Helen S pur

-GGL-----PHVL----ELKVGSRVMLTRNMD-----V----T--D----G----LVNGALGTVVDF

>XP 002772304.1 Perkinsus marinus ATCC 50983

>Hel2 F oxy

_____ -----АР-----DD-----К-А-------GNL-----AKQI-----PICIGARLMLTSNLW-----Q----P-V----G----LCNGARGTVYDI

>Hel2 M ani

>Hel2_P_chl

>Hel2 P lil

>Hel2 F mon

>CAB1116976.1_Ectocarpus_sp._CCAP_1310/34

>RZC87713.1_Papaver_somniferum

>Blastocystis sp

>Blastocystis sp2

----N-TG--------К-----К -----CQA-----PAVL----PLKVGAQVMLLKNLS-----V---E--M----G----LVNGSRGVVDSF

>D discoideum

-CPA-----K----G-----LVNGSQVVLLRKIE-----K----G-D-----G----LVNGSRGVVVDF

>KAF2073656.1 Polysphondylium violaceum

_____QPFGG-___IQLVLVGDFFQLPPVY-_____GN-____GN-_____QPFGG-____QPFGG-_____YAFES-_____YAFES-_____KAWKK--_SI-D-ICLELTTVMR---Q-R--__D-L---------

>KY093685.1 Tieghemostelium lacteum

---T----LVNGSRGVVVDF

>XP_012754920.1_Acytostelium_subglobosum_LB1

>A subglobosum

>H album

>C fasciculata

----K-N--

>XP_014228054.1_Trichogramma_pretiosum

>XP_017781591.1_Nicrophorus_vespilloides

>NP_942102.1_Danio_rerio

>H_sapiens

>XP_034314500.1_Crassostrea_gigas

>XP_028415325.1_Dendronephthya_gigantea

-----EFINILNNIRVG--R-C-P----D-----EV-----EV----------VE--RL-SR--SK------ENKI------

-----D-SE---------G-----G-----------CPA-----KAKL----ELKEGAQVLLTKNLD-----V---G-Q----G----LVNGARGVVKSF

>XP 031556309.1 Actinia tenebrosa

----R----R------

>XP_003388034.1_Amphimedon_queenslandica

>NP 001293174.1 Caenorhabditis elegans

-TLA-----QKKL-----VLKVGAQVMLIKNID-----V----I--K-----G----LCNGSRGFVEKF

>XP 004991536.1_Salpingoeca_rosetta

-----I.-A--

>M conductrix

-----CPA-----RRTL----ELKVGAQVTLIKNIS-----Q----R--Q-----G----LVNGARGVVEKF

>Helicosporidium sp

PLSTFORR-ALFAVASGRSLFFTGCAGTGKSHLLRAVLDSL-PA
HGTHVTGTTGLAASALGGCTLASW-AGTGRLD-HGAFAELSFAELL
-RGERARRWLAVRTLVVDEVSMLDGRWFDA-LERLAREIRRRD
RPWGGVQLVLSGDFHQLPPVSRDGRDGRDG
STWGRVIK-EQLTMTQVFRQGEDL
DFVHLLADVRRGV-C-TGEGEGVRAL-RLRC
RSLENHGPGEEEERKQDQAGVERKQDQAGVERKEDRAGATKTEDQ-IDVIFKNDPAQPPFSLA-SAPFSLA-SAAAA
I-VSP
R-V-FGDDD
VR
G-ACPAESRLELKLGAOVILVRTVCAAAA

>XP_023909855.1_Quercus_suber

-----D-V-V-F----------CMA-----PERI-----VLKKGAQVMLIKNVD-----D----S----LVNGSPGRVLGF

>P griseal

-TLSNEORH-VKDLV------RS-----QSVFFTGPAGTGKSVLMRAIIED---L-K--KWK------CS--

>XP_009351018.1_Pyrus_x_bretschneideri

--LLSHEQRH-ILQLV-----E-----E----------EG-----HSIFYTGSAGTGKSVLLREIIKT---L-R--R--KYS------

>S cerevisiae

--CLSKEQES-IIKLA-----E-----E----------NG----HNIFYTGSAGTGKSILLREMIKV---L-K--G--IYG---------FLA-----PKEL----HLKVGAQVMMVKNLD-----A----A-----A-----LVNGSLGKVIEF

>T phaffii

-----FMA-----PKVL----PLKVGAQVMMIKNVD-----S----T----LVNGSLGKIVAF

>C_viswanathii

----ILSKEOEY-ILKRV-------HG----VSLFYTGSAGTGKSVLLRSIIKS---L-R--E--KYD--------M-----

>XP_002163633.2_Hydra_vulgaris

-----GFVSLLNRLRIG---Y-L-T----P-----LD-----LD--------IE--VL-KH--CK-----GTAF-----

-----P-DD----------G-----G-----------SRY-----FKVL----NLKVGAQVMLLNNLS-----V---S-N----G----LVNGARGVVTKF

>KAF4753487.1 Perkinsus olseni

>C roenbergensis

>KAG5544865.1_Rhododendron_griersonianum

>XP_020415763.1_Prunus_persica

----E-OL----E-O--

>XP 016507676.1_Nicotiana_tabacum

-----K-N--

>XP 021598660.1 Manihot esculenta

>XP_019426349.1_Lupinus_angustifolius
>XP_029145904.1_Arachis_hypogaea

>XP_030518540.1_Rhodamnia_argentea

>XP_026396572.1_Papaver_somniferum

>KAF6167112.1_Kingdonia_uniflora

>KAF5202456.1_Thalictrum_thalictroides

>XP_010275116.1_Nelumbo_nucifera

_____D____ -----GIA-----PDEV-----ELCVGARVMLTKNIA-----L---S-D----G----LVNGATGTITGF

>MQL92731.1_Colocasia_esculenta

-GIA-----PEVL----ELCVGARVMLIKNTD----P---A--A----G----LVNGSTGVVTGF

>RWR91934.1_Cinnamomum_micranthum_f._kanehirae

>XP 021855182.1_Spinacia_oleracea

>KAG6555887.1_Marchantia_paleacea

>KAG0621209.1_Ceratodon_purpureus

>XP_024357988.1_Physcomitrium_patens

-GMA-----PTKL-----QLCVGARVMLLQNLN-----V----K--G-----R----LVNGATGTIIGF

>M polymorpha

-----D-N--

>XP_002987435.1_Selaginella_moellendorffii

>XP_024538624.1_Selaginella_moellendorffii

>E_dispar

>XP_004258641.1_Entamoeba_invadens_IP1

>A_castellanii

>N gaditana

-----G---------P-PP---------CLA-----ESRL-----RLRTGAQVMLLKNVN-----A---N-L----G----LVNGAKGRVTSF

>P multistriata

-----SLTEEQRK-AAEWI-----FGN----A-GEDHD------EE----DSAP----RNVFVTGSAGTGKSHLLKYIVHA--L-Q--S--RES-------

>CAZ69470.1 Emiliania huxlevi virus 99B1

----SLTVCOOD-VLNKT-----L-----L-----------

>CAE7678393.1 Symbiodinium microadriaticum

>RHW71036.1 Trypanosoma brucei equiperdum

FE-D-----STL-----PTDL-----ALKVGTRVMVLQNIS-----L----R--L----G-----LVNGSVGEVVGF

>XP 029239885.1_Trypanosoma_rangeli

--P-------SDL-----PAVV-----SVRVGCRVMLLKNLD-----V---S--V----G----LVNGSVGTVENF

>T gravi

>L braziliensis

P-D-----GAL-----AKVV-----QLRKGCRVMLIKNFD-----S----R--L----GAL-----LVNGSTGTVTDF

>L_seymouri

----ALSDEORY-AYRLA---VH------EH----RNVFITGGAGTGKSHLLRAIIKD---M-P--C---

P-A-----NNL-----KEVV-----RLRKGCRVMVIKNFD-----A----O--T----K----LVNGSTGTVTGF

>B_saltans

----TLSAEQQF-ILDLV---------HQ----RSVFLTGGGGTGKSFLLREIIDQ---L-D--K---------VK----D-Q-----CPA-----G----LVNGSIGVVTTF

>P fungivorum

--PMSDEQAE-IYAAV-----M-----M----------SG----NNLFFTGSAGTGKSFLLKKIWAG---L-D--K--L----

 Image: Non-Normal Non-No ------I.I.------

N-P-----LTI-----E----HINGSRGIITSF

>T socialis

-----CIA-----AQQL-----SLKEGAQVMLLKNLD-----P---A--G-----G----LVNGSRGVVTGF

>T_socialis2

----MLDAIQQE-VTDKV-------RG----ESVFFTGSAGTGKTFLLNTILOC---L-K--E--KWG------- I.------

>M polymorpha2

----PFSPEOOR-VIKLV--------EG----KNIFFTGAGGTGKTYVLKYIIAS---L-K--K----N-----

N-----D--K-D-----YFVKLLQNIRTG---L-N-P-----D-----D---------SV--------EE--IV-LK--CS-

-----LQDH-----GFWH-A-----CIA-----DQVL-----RLKTGAQVMLIRNIKRP-----GSQ----K--L----S----LVNGSRGIIVGW >KAG6541057.1 Marchantia paleacea

----С-ЕН----

-----R-S--

----EE--IV-LK--CS----LQDH-----GFWH-A-----CIA-----DQVL-----RLKTGAQVMLTRNIKRP-----GSQ---K--L---S----LVNGSRGIIVGW

>KAG0609116.1 Ceratodon_purpureus

>P oligandrum2

-----CPA-----PPTL----SLKKHARVMLIKTIN-----P----A--S-----G----LVNGCRGVITGF

>XP 009828150.1_Aphanomyces_astaci

----K-G--

>P multistriata2

----N-----S-------CLA------ERKL-----QLKIGAQVMLIRNLS-----Q----N--S-----G-----LVNGSRGTIVGF

>MBS3922931.1_Nitrosarchaeum_sp.

>C Zambryskibacteria

>C Falkowbacteria

>MBP6882245.1_Candidatus_Levybacteria_bacterium

>NKQ38702.1_Methanosarcinales_archaeon

>MBD3359246.1_Candidatus_Buchananbacteria_bacterium

>OGH84178.1_Candidatus_Magasanikbacteria_bacterium_RIFOXYA2

>C Moranbacteria

-----QIUQLGDIRNN--T-V-N----E-----GT-----GT-----QIWKE----MD--V-KICYLSEQFR----H-C------D--N-----------QIUQLGDIRNN--T-V-N-----E-------GT----------------------VE--KL-QE--TG-------TDF------DS---

-----s-----

>MBI2830749.1 Chloroflexi bacterium

-----K-----K------

>Flavobacteriaceae

>WP_096064617.1_Psychrobacter_sp._FDAARGOS_221

-VRT-----SDEL----TLKIGAKVMFIKNNT-----E-L-----G----VSNGTMGELVGF

>ONG38169.1_Enhydrobacter_sp._H5

>Aalborg AAW1

-----K-S-------MLA-----PEVL-----YLKVGAQVLFVKNNP-----V--K----G-----YYNGTTGEVVGF

>WP_033523136.1_Bifidobacterium_merycicum

>YP_009595951.1_Acinetobacter_phage_vB_AbaM_ME3

>DAJ82417.1 Podoviridae sp.

>Rickettsiales

>MBL6664806.1_Rickettsiales_bacterium

>NBR95534.1_Proteobacteria_bacterium

>MBN8828841.1_Sphingobacteriia_bacterium

>MBQ7287145.1_Candidatus_Gastranaerophilales_bacterium

-D-N---------CSA-----EKSI-----SLKIGARVMLLVNLD-----F---D-K----G----LINGSCGNVKEI

>MBE7709962.1 Cyanobacteria bacterium SIG32

>QBK85639.1_Marseillevirus_LCMAC101

>QBK86258.1 Marseillevirus LCMAC102

-----EAI-----PKAI-----ALKVGAQVMLKCNLD-----V----K--G-----G----LVNGSRGVILKI

>OBK87070.1 Marseillevirus LCMAC103

-----V-R-T-Y----------I.D------

>BCU09408.1_Sicyoidochytrium_minutum_DNA_virus

>ADX05998.1_Organic_Lake_phycodnavirus_1

_____QK-____K-N--

>ADX06411.1_Organic_Lake_phycodnavirus_2

>YP_009173733.1_Chrysochromulina_ericina_virus

>YP_008052747.1_Phaeocystis_globosa_virus

>ANS04235.1_uncultured_Mediterranean_phage

>QPI16828.1_Virus_NIOZUU159

>YP_009010863.1_Invertebrate_iridescent_virus_22

>QNH08436.1_Invertebrate_iridescent_virus_Kaz2018

>YP 009046811.1 Armadillidium vulgare iridescent virus

>KAF0979914.1_Naegleria_fowleri

>H_opuntiae1

-----CMF------FKTL-----DLKVGSQVMLVKNNF-----P--E-----G-----VINGTKGVVVGF

>XP_005717394.1_Chondrus_crispus

>PXF40737.1_Gracilariopsis_chorda

>G muris

>EET02286.1_Giardia_intestinalis_ATCC_50581

----NLSFEQKL-LFNAA---V--I--------RR----KSLFFSGSAGTGKSHLLRAIIKG---L-S--R--LED---

-----O-H-----LLDP------PPVI----SISIGAQVIITKNID-----V----V----Q-R------G-----LCNGROCVVKDI

>A deanei -EWTSEORR-ATOLF----SG----RNVFVTGAAGTGKTQWLLHLIRQ---VIP--N--S-------Q----

-----G----G----G----G----CINGSLGVVMDF

>KAG5479457.1_Leishmania_martiniquensis

-SWTREQQR-AMQLV-----R-----R-----------AG-----HNVFVSGAAGTGKTEWLLHVLQHV--LPRTRQ--RQGLKSGAHPGAEEG-

-----D-E-----VSL-----PPVL-----TLKVGAQVVLLASLP-----N----E-P----S----LANGNLGVVVGF

>CUF06097.1 Bodo saltans

---VLDASQQA-AVEAA-----G-----G----------RG----ENLFVTGGAGTGKTLVVKRIVDS---L-R--A--A---____K__ -----I.N-D------A---------GRL-----AQTI-----PLKIGTKAMLLTNLN-----V----R--A-----G----LVNGAVGVVTGF

>Phytomonas sp

-----ASLFVGGKAGTGKSFLLREIVHK---M-R--L--R---------RLSAEQAQ-TLSLA-----L----

-----KSHIMLLGVGKGDSAGCFSSVEAQK----CKYTKQKFWSDVILL--HFTNREGFSIRFGHGRGDLGSRRWARGKPRE----ITYSEVQSIVH-EICQAKTLDSER---FFAYV--LPFAYCYSPNIHSVAVRTFGHNRKEATMQLKGFLASASERM-----------DLV-----TQSK-----KLKVGCRVMLLRNLN------

>S culicis

>DAC81588.1_Hydra_MELD_virus

-----FIFTC----Q-K-----DD--E----

_____TP_____TP_____K-T_____

>H_opuntiae2

>MBQ6280177.1_Mycoplasma_sp.

>P grisea2

V-D-

>QDP67633.1_Prokaryotic_dsDNA_virus_sp.

L_______KN=_____KN=______SK=____SK=__DNVFLTGAPGTGKSWLVDRYVEW=-L_L_E=_N-_____QDEI=____I -----G-EEPVITASTGIAALNI=____NG=___KTLHSW=GGL=_R=N-__DH=P=-I=-D=__ER=__D-__ER=___QDEI=___I -K=__G-__Y=___SYENY=_IS=TCTLIIDEVSM=_VSAALLEN=INILAKRIR=____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____GR=_____EFTDILQNIRGG=_FL=T=___EFTDILQNIRGG=_FL=T=____FQ=_____PQ=_____EDWDE=___AD=F-TVCYLHENKR=___QS=____IKDA=___SI=____ _____ -----VE-----D-P------K---------K-N--

>C Uhrbacteria

----KISKEFKK-ALAIM---ED--------TK----EHLFLTGNAGTGKSTLLQYFRKH---T-A---------S-R--

>C Aenigmarchaeota

----EINDKFKE-SLGLM-------TS-----KNIFITGKAGTGKSTLLNYFRSL---T-D------------LPT-----REISL-----KLKVNSQIMLVNNDP-----N--G----R----WVNGTVGKIIGI

>C_Pacearchaeota

-YGPPP
KPEGGVOMAFIGDLYOLPPVVKGE-KGVKGVF
DD
LPTWVNGTVGKIIEI
>NMD11668.1 Acidobacteria bacterium
EINDQFRQ-ALHWMEETA
KKIAVLAPTGVAALNVKGQTIHSF-CGFK-PDI-TLAKVRAKVR
INAKKDPDRAALLRKLDAVVIDEISMVRADLLDC-VEKFLRLNGPPK
-PRAGV-E-KTLRPFGGLQLILIGDLYQLPPVVAGV-E-KTLRPFGGF
TLHDSFRL-EFVELEKIYRQ-TYFFSAHCLLRDSFRL-EFVELEKIYRQ-TD
AGFIALLNAVRNRS-A-GPEDEDEDLEKL-HSRYDPEF
/PPEPED
YRVTLTSTNDLAAARNREKLA-LLPGRRR
EEEEE
3-SLPTDEHLEIKAGAQVMLLNNDAA-GRWVNGSIGRIAGV
>NPV00061.1_Brevinematales_bacterium
EINPEFAK-AMDFMENENENGKHHVFLTGKAGTGKSTLLSYFCENT-G
LNHVILAPTGVAALNVGGQTIHSF-FGFR-PNI-TKDQIKDQIK
-SDWN
TGN-E-EQIETFGGIKMIFIGDLF0LPPVVTGN-E-EQITGN-E-EQIFF
KDYV-RYIELTKIYRO-NDR
REIDILNVRNNN-I-TYRDRD
SEDE-F
(
GDD
F
>cd WWE3
SGSGSGSG
QLSNEFKN-AINLIETSGSGSGKNIFITGNAGTGKSTLLTYFTKVT-D
QLSNEFKN-AINLIETSGSGSGSGSGSGSG
QLSNEFKN-AINLIETSGSGSG

>archaeon CG07

>VVB74890.1_uncultured_archaeon

-----DFIAILDSIRTG---E-F-D-----E-----KT-----KT-----------NPNF-----NA--------D-----E-S-----

>MBI5066474.1_Candidatus_Woesearchaeota_archaeon

>HHG53312.1_Spirochaetes_bacterium

>PIR99148.1_Candidatus_Collierbacteria_bacterium_CG10

 >>PIR99148.1_Candidatus_colleral_bacteria_ba

>C_Micrarchaeota

_____ _____ -----G-----E-L-----

>Prevotella sp

----LONPELOK-ALOII---OF---------TH----NSLFLTGKAGTGKSTFLRYISST---T-K---------S-S--

>B ovatus

----PONHEOOL-AYELV-----AN-------TN----SSFFLTGRAGTGKTTFLHNVQKL---A-G-----------FPV-----DLEL-----RLKVGAQVMFTRNDQ-----Q--K----R----WANGTLGKVTKL

>M mazei

>Thermoplasmata

>MBP7674859.1_Thermoanaerobaculia_bacterium

>C collierbacteria

>C Pacebacteria

TLSQEQQE-VFNKLETETTNTNGHFFITGKAGTGKSLLLQYFRTYS-Q
KKLVVLAPTGVAALNVGGQTIHSL-LRLPFSA-ITLDSFRRLRV
-DTKKKLLQS-LDCIVIDEISMVRVDIMEA-IDYILKKARNSSS
TSGEPFGGVQMIMFGDLYQLPPVVTSGELQQYFF
DDTYGGAYCFNANSWRAAKP-EIITLSKIFRQ-SDA
TFIDLLNSLRDGN-P-NSIADFDF
PP
ADDDD
GKKKK
EEE
FPTDKVLQLKKGAQIMMLKNDRDKRWVNGSLGTIHSI

>Curtobacterium_sp

ELSDEORA-VFEYIEHEH
KOVVICAPTGVAALNVGGOTIHSL-FRLP-IGL-IADAELRAE
-GPDTRKLLNTIDTLVIDEVSMVNADLLDG-MDRSLRKARG
GDADE-RAY
TDHAEL-NIIELATVHRO-R
AFAAMLTAVRHGR-V-TA
P
AKKKKKK
G
PPADEALELKPGAHVMFLRNDADQRWVNGTLGIVTAI

>Clavibacter sp

-----R-T--

>P faecalis

_____ -----GA-----R-T--

>Leucobacter_sp

-----ET-----ET----------TR----EHLFITGRAGTGKSTLLNHLAQN---S-S----------V----E-G------

>QIG57888.1_Microbacterium_phage_PauloDiaboli

>Parabacteroides

----------SE-----LE-----D-G -----S-A---

>DAU76505.1_Myoviridae_sp.

----IINSAMKE-AIDLV-------LN-------TN----TNVYLTGRAGTGKTTLLRYILGV---C-K-------N-A--

>AFB75491.1 Bacteriophage sp.

-----QD--------DE----NNVFVTGKAGSGKTTFLKYLIEK---S-G----- >DAD87486.1_Siphoviridae_sp._ctAUQ2

>DA003073.1_Siphoviridae_sp.

>DAQ71114.1_Podoviridae_sp.

>DAJ22427.1_Podoviridae_sp._ctfN46

-----APC-----EDKL-----VVKVGAKVLITRNGC-----G-----G-----YVNGSTGIITSI

>DAL07837.1_Bacteriophage_sp.

DKNVEQGR-ALKKIFTFTFTFTTRTRENLFITGRAGSGKSTFMRRIVKFL-G
KCVIVAPTGVAALNAGGQTIHSF-FSIKNDP-YIPSIERGMLSNKV-D-V
PF
TADFEPFGGVRLIMFGDLSQLPPVVTADDFF
DKYSGF-SVITFENVFRQ-KFFFSSFALRASGF-SVITFENVFRQ-KDP
QLLSVLEDIRCGV-I-TDESES
DN_DN

>DAY30538.1 Bacteriophage sp.

-----APV-----EKTL----FLKEGSRVMITRNGG------E----YFNGSLGTVLSI

>Hyphomonas_sp

TIYAK-PAEWVSRGSRGAGAQGNLFLTGRAGTGKTTLLRRFVEQA-GSRG
DSAIVLAPTGVAAMNAGGQTLHSF-FKLPPRL-IQDVKQDVK
R
RGD-E-DPIRFFGGVRMILSGDLHQLPPVVRGD-E-DPIRFI
KERAEF-ALLALKHVFRQ-ED-P-PPAFKEAEF-ALLALKHVFRQ-EDP

-----SDRD-----AV--------EA-----S-----E-Т-----

>NQY15510.1_Henriciella_sp.

>Robiginitomaculum_sp

-----EK-----EK-----------SR-----DNVYLTGRAGTGKTTLLKAFVAR---N-A-------

>MBT3274541.1_Spirochaetales_bacterium

_____ -----О-Е--

>A_illinoisensis

_____ _____ -----P------P------Q-G-

>MBL6903300.1_SAR86_cluster_bacterium

-----SFDEIKDQ-VIHLL-----DND----------EQ----EFIYLTGAAGTGKTTLLEVIKAD---L-D---------N-D--

>MBR3410882.1_Candidatus_Methanomethylophilaceae_archaeon

-----YPA------PEEL-----VLKKGAKVMFVRNDD-----P--H-----G-----YVNGTFGIVESV

>MBR5312660.1_Clostridia_bacterium

-FPV-----E--R-----WVNGTLGIVNNL

>MBU4069976.1_Nanoarchaeota_archaeon

-----TN----TNILITGPGGTGKSTILKKFKEK---T-K------TNILITGPGGTGKSTILKKFKEK---T-K----------VDKFDKEG-LFTFL----ER-----

--CNA-----G---EEKL-----ELKIGARIMVLINDA-----G---EN--K-----R-----YFNGSLGTVKEL

>MBQ7366747.1_Spirochaetaceae_bacterium

-----FLT------PETL-----ELCIDATVMVTKNTS------S-----S-----G-----LINGNMGRVVSF

>CAB4198187.1_uncultured_Caudovirales_phage

-----CFDDPN--K-----A-----YFNGSIGLFRGF

>DAM57115.1 Myoviridae sp.

TYEIGTDA-ALAAVL-GL-GCGCGENVYISGPGGTGKTHLLQDIQSLL-G
ESCMVVAPTGVAALNAGGVTAHRA-FDLSAGV-TVPEDFTEDFTEI
-RSKTAKPLKSKALRTLVIDEVSMVRADKFVE-MDKKLQHLRKKT
STQE-REDEPFGGLQVIMFGDFYQAQPVISTQE-RED
YKYWDTDLCFYTQSWKDLNL-KCVALVEQFRQ-ESI
RFATMLNCVREGR-R-TGDVDVVKEL-NSRC
AEE-R-M-YE-R-M-YE-R-M-Y
KK
LPVEDLMCLKVGMKVMIVANDLN-PNHKVPCYVNGSRGTILKF

>YP 007006388.1 Escherichia phage FV3

QYDVGTDA-ALVAIMM
DSCITVAPTGVAALNVNGATAHRT-FDLAAGV-SMESDWTSDWT
-RAKTAKPLKSKAFTILIIDEISMIRADKFIE-MDRKLRFLRKKNNDD
SSME-KEAKPFGGIQVLLFGDFYQAPPVVSSME-KEAKEA
FNFYHTD
RFATMLNCVREGR-R-IKEVEV
PP
APPPPE-K-T-YAIKAPPPP
G
F
LPVEQEMRLKIGMKVMITSNDVDPTHKVPYYVNGTRATVVKF

>AXC42890.1_Escherichia_phage_LL12

QYDVGTDA-ALIAIM
DSCITVAPTGVAALNVNGATAHRT-FDLAAGV-SMESDWTNG
-RAKTAKPLKSKAFTILIIDEISMIRADKFIE-MDRKLRFLRKKNDD-
SSM-E-KEAKPFGGIQVILFGDFYQAPPVVSSM-E-KEAKEA
FNFLD-L-L-HNIALVDQFRQ-ESISIESWKELD-L-HNIALVDQFRQ-ESI

-----S-T--

>YP_009851551.1_Erwinia_phage_pEp_SNUABM_01

-----RPG-----PDEL----ELKEGLKVMITANQM-----CQPNED--P----A-----YVNGSIGFIKKM

>YP_009854417.1_Erwinia_phage_Hena1

_____ -----N----P-Q------------KPG------PDEL-----ALKVGLKVMITANQI------SKPHED--P----A-----YVNGSIGFIRKM

>YP_008857219.1_Escherichia_phage_4MG

-----KPV-----PEEL----YLKEGAKVMITVNDP-----K-GFEE--P-----E-----YVNGSRGEVIEL

>QPI14547.1_Salmonella_phage_GEC_vB_MG

_____ -----P-----L-D------ITITSTNAAADKVNKKRFE-EV----P--G------M-----M-------P-TL-Y-------A----------KPV-----PEEI----YLKEGAKVMITVNDP-----K-GFDE--P----E----YVNGSRGVIIEL

>YP_009835918.1_Raoultella_phage_Rol

-----DFGVGTEE-ALGAI-----I----------EG----KNVFITGPGGSGKSHLIKTIQSL---Y-S-----------A-Q-------RPV-----AESL----HLKVGTRVMITVNDQ-----NPDEDG-P----K----FVNGTRGIIKAL

>AXN57909.1_Acinetobacter_phage_ABPH49

-----SYEIGSES-AIKSI-----M-----------SG----KNTFITGPGGSGKSQIIHTVQDM---L-G----------RPV-----AEVL-----ELKEGLKVMITANDQ-----A---VP--S-----R----YVNGTVGIVRRM

-NEK	N- /
TEFLCSTNKEADAVNKHNYD-DVMGEEEEEEEE	
LPVPVLSLRVGVRVLICANAEDGSYYNGMTGYVEKM >QDP60500.1_Prokaryotic_dsDNA_virus_sp.	
	iD-
TEHLCCYNKDADYINQLYYS-KMEGEEEEEEE	-E
>QDP64781.1_Prokaryotic_dsDNA_virus_sp. NNSVEQDL-ALKYIL	< K-
VPPP	-D
>AUR91792.1_Vibrio_phage_1.164 DLKLKQQE-ALGLMKGGATIHRT-FKLPIANVFLTGKAGTGKSFVTDLFTAWA-EEQ	>

--AQ---RS----

----R--V---

------RPV-----DevL----NLKEGLKVMIVVNDN-----DQKKKE-P---D-----DVNGTVGIIKKI

-DDTIFLAPTGIAALNI-----KG----ATIHRT-FKL---Q--L----GY-L---D---P--

>QEG12160.1_Klebsiella_phage_vB_KaeM_KaOmega

>YP 009595586.1 Pseudomonas phage pf16

----OLNKKOTY-AFEOI-----M------

>AUR89562.1_Vibrio_phage_1.124

>DAE75004.1_Bacteriophage_sp.

>DAF60248.1_Siphoviridae_sp._ctqK313

>DAP73423.1 Bacteriophage_sp.

_____ _____ ----P-F-_____ -----BAV-----D------S--G-----N----YVNGTIGIIQKI

>DAE16492.1_Siphoviridae_sp._ctqBH20

-----KLNKKQRY-ALDTM-----L----------SG----SNVFLTGDAGTGKTTVIQTFIQE---A-E--E--M----------N-I--

>DAG97916.1 Ackermannviridae sp.

-----ELTEEFKK-AYDLL-----EH-------TK----EFVFLTGDAGSGKTTFLKWWLSN---T-S----------CPV-----REIT-----RVRPGCRIMCRNNDK-----D-E----R----WVNGTIAKFVKK

>DAX71650.1_Myoviridae_sp.

>ARR75030.1_Mimivirus_AB566017

>QHN71346.1_Mollivirus_kamchatka

>YP_009165351.1_Mollivirus_sibericum

>C sorokiniana

>ARX71979.1_Erinnyis_ello_granulovirus

TLNEKQQK-LFDYLTQTK
D-KIVFVAAYTHLAARNINGKTCHSL-FRFDFEL-NLLRRRR
AG
KHKHKH
DIWYYF-ELYELTENMRQ-SEP
EFIANLNMLRVGD-VVNAEKCKCLSYF-NRFVVNAE
TQNI-QDC
CDDDD
T
T

>YP_009506054.1_Clostera_anastomosis_granulovirus_B

-____TLNTRQQK-LFDYL-____TQT-__K-____SF-___SF-___SFVFISGSAGTGKSALLVALREH--W-L-A-E-____ ---__D-KIVFVTAYTHLAARNI---__NG--__KTCHSL-FRF--D-F-__DL-N--L--L--R--___R-____ -----A--___Q--I--GV--PHYVIIDEISM--VPEKMLDG-IDSRLRQNS------G--__K--___S-____K--___S-____S -____A--____DFGG--__VNVVVFGDLYQIPVD--____KH--____KH--_____KH--____S-____NUVVFGDLYQIPVD--_____KH--_____S-____NUVNFGDLYQIPVD--_____KH--_____S-____NUVNFGDLYQIPVD--_____S-____DIWHS--___FELYELTENMR---Q-S--____PPYKS--_____DIWHS--____FELYELTENMR---Q-S--_____

-----UDAA-----VDAA-----

>YP_654526.1_Choristoneura_fumiferana_granulovirus

>NP_663278.1_Phthorimaea_operculella_granulovirus

>AIU36910.1_Cydia_pomonella_granulovirus

>AD085536.1_Pieris_rapae_granulovirus

>NP_891963.1_Cryptophlebia_leucotreta_granulovirus

---TLNKEQKY-LFDKV-----ADT----H-----------W-R--NF----SPIFVTGSAGTGKSALLMTLRNY---W-R--N--Q-----SPIFVTGSAGTGKSALLMTLRNY---W-R--N--Q-------------Y-T.-----EQ-----E-K--

>QOD40078.1_Matsumuraeses_phaseoli_granulovirus

-LIF-----KKDL-----KVCVGTRVMVTHTTT------H----FCNGDTGVIERI

>YP_009182312.1_Diatraea_saccharalis_granulovirus

D-KNVCVTAFTHLAARNIFGKTCHSV-FGFDF	KM-NLDNN
	DIVIERNS
	N
WVIVFGDDIQDQFVG	
	LD VE DV UV
FIMÖNTNPIKAGN-I	MKPIMKPI
	S1-A
YKG	KDD-E-V-M
ESVESV	RTHQH-T
TVF	GQK-T
IIFKPIIRLFPGARVMITHTTD	WFCNGDAGIVERI
>YP 003517787.1 Lymantria xylina nucleopolyhedrovirus	
LLNAKQQY-IFDYFTQRD	SFAPVFVSGSAGTGKSALLMALHEFW-RRRR
KTCHSL-FGFDF	NL-NAKCC
TTPPLPV-KPRCVIIDEISMIPAKMLDG-II	DRKLQQTTHD-
VNVIVFGDLYQLPPVN	KTKT
KPVYAA	DAWNAF-RLYELTENMRQ-SES
VFIDNLNLLRVGD-FKCKCKCKCKC	LKYF-NSLKLKYF-NSLK
	QL-K
SSSA	QSR-V-V-V
NAV	REHMERD-A
QIF	EqE-K
LIFKPQLTLCAGARVMITHTTA	EFCNGDLGTVESV
NOVO71652 1 Orania providetovente puelopelukodnovinue	
>Qw0/1653.1_Orgy1a_pseudotsugata_nuclopolynedrovirus	
KLNIKQQL-LFDFLTQAT	EFRPLFVSGCAGTGKSALLRALRNFW-TRQ
N-ETVYVAAYTNLAARNVDGKTCHSL-FGFDF	KL-NVKRPF
SLKVPHCLILDEISMIPGQMLDK-II	DEILKRACDE-
VNLVVFGDLYQLPPVD	KNKN
KPVYEA	KVWPQF-TLYELTENMRQ-SEA
KCKCKC	LKTP
	PTV-ENOL-N
NTCLVSTHNESNSINVNCYN-AITI	OVE-T-V-V
RR	
	E-NE-NE-N
	NFCNGDEGIVESV
MIT KOND KDCFGIKIMVINIIN	N FCNODEGIVESV
>YP_001651017.1_Orgyia_leucostigma_nucleopolyhedrovirus	
OLNAKOOS-IFNYLTEKD	TFEPIFVSGCAGTGKSALLKALRKFW-FKE
=====K=KTCVVVAAYTNLAARNV=======OG======KTCHSA=FGF===D==F====	KL-NIRRR
KTVVVAAYTNLAARNVQGKTCHSA-FGFD-F KTCHSA-FGFD-F	KL-NIRR
K-KTVVVAAYINLAAKNVQGKICHSA-FGFDD-FC PSS-KPDYVIIDEISMIPAQMLDK-II 	KL-NIRR
K-KTVVVAAYINLAARNVQGKTCHSA-FGFDF IPL-SS-KPDVVIDEISMIPAQMLDK-II VGVVVFGDLYQLPPVD	KL-NIR
K-KITVVAAYINLAAKNVQGKICHSA-FGFDF KICHSA-FGFDF 	KL-NIRRG
K-KITVVAAYINLAARNVQGKICHSA-FGFDF KICHSA-FGFDF 	KL-NIRG
KTCHSA-FGFDF KTCHSA-FGFDF 	KL-NIRRGATS- DTKLKYSSF
K-KITVVAAYINLAAKNVQGKICHSA-FGFDF 	KL-NIR
K-KITVVAAYINLAAKNVQGKICHSA-FGFDF 	KL-NIRGATS- DTKLKYSSF
K-KITUVVAAYINLAAKNVQGKICHSA-FGFDF 	KL-NIRRGATS- DTKLKYSSF-KLFELTENMRQ-SEA VDFF-ST-LT
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	KL-NIRG
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	KL-NIRGATS- DTKLKYSSF-KLFELTENMRQ-S
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	
KTCHSA-FGFDF 	KL-NIRGATS- DTKLKYSSF-KLFELTENMRQ-SEA
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF KICHSA-FGFDF 	
KICHSA-FGFDF- 	
KICHSA-FGFDF 	
K-KITUVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	
KICHSA-FGFDF 	
K-KITVVVAAYINLAAKNV	
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNV	
K-KITVVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITUVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVVVAAYINLAAKNV	
K-KITVVVAAYINLAARNV	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNV	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
KICHSA-FGFDF 	
K-KITUVVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVVVAAYINLAAKNV	
K-KITVUVAAYINLAAKNV	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF DF	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	
K-KIVVVAAYINLAARNVQGKICHSA-FGFDF 	
K-KITVUVAAYINLAAKNVQGKICHSA-FGFDF 	

-----QK--FF-DD--KV-----LQKS-----

>AXS01146.1_Spodoptera_frugiperda_granulovirus

>YP_009249960.1_Mocis_latipes_granulovirus

>NP_059294.1_Xestia_cnigrum_granulovirus

---GO------

>QNH90674.1_Mamestra_configurata_nucleopolyhedrovirus_B

>QKV50030.1_Plutella_xylostella_granulovirus

---TLNEQQQK-IYNYL----TSV----D-----------CF----EPIFVSGSAGTGKSALLVTLTKA---W-T--M--K--------

>YP_009186748.1_Sucra_jujuba_nucleopolyhedrovirus

-GVI-----PEKI-----TLAIGSRILVTSNCV-----N--S----H----CINGDIGVIVDF

>YP_010086327.1_Hyposidra_talaca_NPV

-SMI-----PDNL----TLAQGSRIMVIANCK-----E--S----K----CINGDLGVVEEC

>YP_009133285.1_Lambdina_fiscellaria_nucleopolyhedrovirus

>YP 009049868.1 Peridroma alphabaculovirus

-QVV-----D-S----E----CVNGDLGVVEKF

>YP_008004327.1_Choristoneura_biennis_entomopoxvirus

-KIF----E--NI----YICKGTKIMITANCV----E--N----E--N-----E--N-----KNSDMGYIDNI

>YP 803305.1 Trichoplusia ni ascovirus 2c

MMTPCQLR-AYNILIENM-NKNPLDPTRLPIFISGGGGTGKSYVLKKFKDYV-VNVNV
N-KKIAVVATQAIAATLIDGKTIHSV-FNIRGGN-AQTPDPDDGKTIHSV-FNIRG
QRCTLTSFPYDVLIIDEISMLNGELLDL-IENTLVTVKRSS
KSKSKSKS
P-RLISLVTNVRH-KGDD
EFSNIMARVRIGD-R-SALKTILKTI
PEI-EHLRFEE-GE-G
ITIVATNRQVQKINNAATK-KFSDNNNN
KTIHN
-IMY
RIVPKSIDIFIGAIVMITANDINGN-GRCN-GDTCKIVNI

>AYD68236.1 Heliothis virescens ascovirus 3h

EVE-----RIV-----PKEL-----SVFPGATLMFTANGL-----S----G---G----P----WCNGDICKVVSL

>P_oligandrum

-----FA--TL-QQ--RT-----SIDP---

>YP_008437119.1_Pandoravirus_salinus

>BBI30459.1_Acanthamoeba_castellanii_medusavirus

>OSX74557.1_Porphyra_umbilicalis

>OSX70336.1_Porphyra_umbilicalis

>AYV86632.1_Sylvanvirus_sp.

----HLSOSORI-AAYLI-----IH-------GR----HNMLLTGGAGVGKSLLADFTSKA---F-K--K--M-----

>CAE7237458.1_Symbiodinium_microadriaticum

----PETKHQRH-AMEHI-----IQE----V-LSRP----------NTK----DGSNPERLHMLLHGPGGCGKSVVIRAAAHM---L-R--Q--G--------GGL-----G-----EALV-----RVAVGVRVMLRHNID-----V---Q--D----G-----LVNGACGFVEQV

>Perkinsela_sp

R-QVVLVTASTGIAAQSMNGRTFQHF-FGIRGDCGDC
SFREGLAKLRYGQ-L-TSEMYSLI-RSRA
DELS-D-
AES-L-A-FPKVLTEP KLSTWTSSVAFTCSEKIPPALRSTGNLRSALNYV-LSGKLFARGAPSDALVLEVLS- ETADVATYFTHANTFWFRFFAADAEDKAATLRRNLIRAIEYIGGDIVHGQGDAILKKVSPAVSPAID
>QFR57062.1_Klebsiella_phage_AmPh_EK29
MLNKGQKK-AFDYIISRI-KAGKG
PIIKVATDIRNGKWIYDHQRDDHGVHGFTSTTALKD-FM-MKYFTTALKD-FM-MKYF
EIVKDP-EDMF-E
DLKITGEVIVMQEPLIKELEFEGKRFND-LKFNNGQYVRIVSA
>YP_010093920.1_Enterobacter_phage_myPSH1140
UNEDQKD-TFNRVVERI-KAGRGRGGHITINGPAGTGKTTMTKFIINYL-IST
E
21F_003130173.1_Edwardsielid_Dhage_PEL20
GULGVULAPTHQAKKVLAKMSGMEANTIHRV-LKIN-PMT-YEQDQDQDQD
DVVKTP-EDLF_E TRMLAFTNKSVEKLNNIIRR-KLY
T = E
VFNNGEMVRIKDC
VFINGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GUMGIVLAAPTHQAKKVLSKLSGQRASGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKVLSKLSGOTANTIHSI-LKIN-PTT-Y-EDQNIFNOGEMVRIKDC
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKVLSKLSGQRANTIHSI-LKIN-PTT-YEDQNI
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKVLSKLSGQTANTIHSI-LKIN-PTT-YEDQNIPHPDMSKCNVLVCDEASMYDGSLFKI-ICNSVPQNI
VPTINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 DLNTGQKE-AFDYITEAI-QRSGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPFINEEVIVMQEPFIKELEFDGKKFSE-I FNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKULSTEA-L-I-QRRSGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPTINEEVIVMQEPFIKELEFDGKKFSE-I PNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKULSTEAI-QRRSGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPFINEEVIVMQEPFIKELEFDGKKFSE-I FNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKVLSTEAI-QRSGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 CUNTGQKE-AFDYITEAI-QRSGECITLNGPAGTGKTTLTKFVIDHL-VRN
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 GVMGIVLAAPTHQAKKVLSKLSGQTANTIHSI-LKIN-PTT-YEDQN1FNF
VPFINEEVIVMQEPFIKELEPDGKKFSE-IFNNGEMVRIKDC >YP_003934611.1_Shigella_phage_SP18
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 CW
VPFINEEVIVMQEPFIKELEFDGKKFSE-IVPNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 CT
VPFINEEVIVMQEPPIKELEFDGKKFSE-IVFNNGEMVRIKDC >YP_003934641.1_Shigella_phage_SP18 DLNTGQKF-AFDYITEAI-QRRSGECITLNGPAGTGKTLTKFVIDHL-VRN

-----PLIQVATEVRQG---EWLRT---NWSK----ELRQGVLHVPN-----------VNK-ML-DT--YL----------SKINTP-EDL----------L-D-----_____IP-----N-E------F---------PVC-----VLVTQMPVMQSNGKY-----PVC-----V----V----IDNGEIVKILDV

>Hel A tha

>Hel_B_nap

>XP_033143622.1_Brassica_rapa

>XP_006279329.2_Capsella_rubella

-----GLP-----KHEL----TLKKGAPIMLLRNID-----P---K--G----G----LCNGTRLIVTOM

>Hel B vul

----CLTCEORS-VYDEI---MMA----V-SRG-------QG-----GVFFVYGYGGTGKTYVWKTLCAA---I-R--S--K---------IYSTE--_____IR-____I -----QLP----T-S----Q----LCNGTRLVIKHL

>CAD1820584.1_Ananas_comosus_var._bracteatus

-SLTDEQKG-VYETI-----ISV----V-SKN---------EG-----GVFFLYGYGGTGKTFIWRTLSAA---I-R--S--K----------GVP------NHML-----KLKQGAPVMLLRNID-----K----S--S----G----LCNGTRLVITQL

>XP_012840144.1_Erythranthe_guttata

-----SITDEORK-VYDVI-----MDA----V-TND-----SG----SGMFFLYGHGGTGKTFLWKTLSAA---V-R--S--K-----

>Hel H ann

-----QFAKWLLDIGEG---N-V-GG----PN----DGEASIEIPSD----LLIT-----DTSDPIST-LI-DF--VY----P-----SIL----E---------GLP------NHRL-----ALKVGVPVMLLRNID-----Q----Q--N-----G-----LCNGTRLQVKKM

>XP 022031972.1 Helianthus annuus

-----GMP------NHKL-----VLKVGVPIMLLRNLD-----Q----K--N----G----LCNGTRLQVVKL

>ABA95557.1_Oryza_sativa_Japonica_Group

-----GIP------NHEL-----KLKVGLPVMLLRNIN-----Q----T-A-----G-----LCNGTRMTITQL

>RCV07316.1_Setaria_italica

>XP 039834415.1 Panicum virgatum

>XP_026386115.1_Papaver_somniferum

-----DFGKWVLDVGDG---K-I-PIS--ETK----DDSTWIQIPDD----LLVKC-----D-NGDYINT-IV-ES--TY----P-----SLL----E----

>PIA60703.1_Aquilegia_coerulea

------GVP------NHLL-----ELKVGIPVMLVRNIN------P----S-R-----CNGTRLVVTSL

>XP_039793773.1_Panicum_virgatum

>TVU37829.1_Eragrostis_curvula

-----GLP------PHEL-----KIKVNCPLILLRNLD------P---H--N-----G-----LCNGTRLVVRGF ----LT-----

>XP_020197274.1_Aegilops_tauschii_subsp._strangulata

>KAF5187279.1_Thalictrum_thalictroides

-----KLNNEQKH-AFDMI-----MDA----V-HHK-------TS----SVFFIDGPAGTGKTFLYRSLLAA---I-R--H--E---------LYOLE-------IS------

>XP_026391420.1_Papaver_somniferum

-----KLNEDQSR-AYKTI-----MEA----I-ERK-------ES-----KVFFIDGPGGTGKTYLCRAILAT---V-R--K--N----------GLP------SHIL-----KLKIGAPIMLLRNVD-----A----K--N-----G-----LCNGTRLIIKEF

>KAG0566608.1_Ceratodon_purpureus

>Hel A ara

>Hel P inf

>KAE9276432.1_Phytophthora_rubi

-----GIP------PHKL-----TLKEGAPIMMMRNLN-----P----P----D--L----G-----LCNGTRLRVVKL

>Hel C sup

>Hel M dem

>CE098944.1_Plasmodiophora_brassicae

>Hel_X_lae

>Hel_F_can

>Hel_E_jap

>Hel_C_ele

>Hel A can

>Hel N ame
>Hel_C_pur

-----OLNODOET-AFKAV-----TEA----V-RDDP------ST-----AHFYLOGPGGTGKTFLYETLACH---Y-R--S--E------

>Hel R del

>XP_005716008.1_Chondrus_crispus

-----ALP-----DHKL-----KLKKGFIVMLLRNLD-----P----A--T-----G-----HVNGARYVIENM

>OSX80228.1 Porphyra umbilicalis

-----SVP-----THAM-----VLKVGMTVMLLRNLA-----A----Q--N----G-----DCNGTRYIVTRL

>KAA6365738.1 Streblomastix strix

>Methanothrix sp

>D bacterium

-----SMSNEQEA-AFRYI-----AD----ISSG---ITSKTIHKAMLDI-----------E--H----NREQF-TR--KDIIIIDEAGM--VATRQMQK-IISEART-------

>D cetonica

>Sphingobium_sp

>Sphingomonas sp

>S_macrogoltabida

>A tumefaciens

>Mesorhizobium_sp

>Phenylobacterium_sp

ALGGEQRD-ALEHITGTGTG
G-YQVRGAALSGIAAESLEAGSSIPSRTIASLEHSW
GRQGRDLLTSSDVLVIDEAGMIGSRQMDR-VLLAAER
AGAGAGAG
WQREATRELATGR-T-GAAAALERY-DAAGMVRAHETREAAREALVDGWE-
AVRRE-APGA-SA-S
QEEEEEEE
GGEE
LAL
GerTFAAGDRIMFLRNERS-LGVKNGTLGTVERI
>A_excentricus

9.3 Material suplementar do Capítulo 3

Supplementary Material

	Та	bl	le	S1
--	----	----	----	----

•			Species	# 01 sequences	location	Sustinission institution	Jubrilission Dat
Insecta							
Order Lenidoptera							
	Superfamily						
	Papilionoidea	Family					
		Nymphalidae					
			Pararge aegeria	31	Scotland/UK	Wellcome Sanger Institute/UK, Stockholm	2021-01-28, 20
			Fabriciana adippe	1	Romania	Wellcome Sanger Institute/UK	2021-04-15
			Heliconius wallacei	1	Peru	University of Cambridge/UK	2015-11-29
			Vanessa cardui	5	Scotland/UK	Wellcome Sanger Institute/UK	2021-02-13
			Dryas Iulia Danaus melanippus	1	India	Iridian Genomes/USA	2021-06-28
			Nymphalis polychloros	1	Spain	Wellcome Sanger Institute/UK	2021-02-13
		Family Riodinidae	Anodemia ares	1	1154	Elorida Museum of Natural History/USA	2021-05-03
			Emesis lacrines	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Emesis aurimna	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Emesis ocypore Emesis heterochroa	1	Peru Peru	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
		Family					
		Papilionidae	Demossius enalle		Cormonu Italu	Florido Mussum of Notural Listory (LCA, Stockholm	2021 05 02 20
			Parnassius apolio	5	Germany, Italy	FIORIDA MUSEUM OF NATURAL HISTORY/USA, STOCKHOIM	2021-05-03, 20
			Parnassius imperator	1	China	Florida Museum of Natural History/USA	2021-05-03
			Parnassius smintheus	1	Canada	Florida Museum of Natural History/USA	2021-05-03
			Zeryntnia polyxena Archon anollinus	1	Greece	Florida Museum of Natural History/USA	2021-05-03
			Protesilaus protesilaus	2	Peru	Florida Museum of Natural History/USA	2021-05-03
		Family Lycaenidae	Current a hullin				2024 05 02
			Curetis bulis Cvaniris semiaraus	1	Myanmar Romania	Florida Museum of Natural History/USA Wellcome Sanger Institute/LIK	2021-05-03
			Lysandra coridon	2	Romania	Wellcome Sanger Institute/UK	2021-02-13
			Lycaena phlaeas	1	Scotland/UK	Wellcome Sanger Institute/UK	2021-03-17
			Aricia agestis Lysandra hellaraus	1	Romania Snain	Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK	2021-01-25
			Lepidochrysops patricia	1	South Africa	Florida Museum of Natural History/USA	2021-05-03
		Courth Direction	Eumaeus atala	7	USA	University of Texas Southwestern/USA	2021-03-02
		Family Pieridae	Pieris rapae	3	Scotland/UK	Wellcome Sanger Institute/UK	2021-01-25
		Family Resperiidae	Pyrgus malvae	3	Romania	Wellcome Sanger Institute/UK	2021-07-21
			Satarupa nymphalis	1	China Costa Disa	Florida Museum of Natural History/USA	2021-05-03
			Pvrrhopvae telassa	1	Peru	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
			Pyrrhopyge sergius	1	Peru	Florida Museum of Natural History/USA	2021-05-03
			Pyrrhopyge hadassa	1	Peru	Florida Museum of Natural History/USA	2021-05-03
			Pyrrhopyge crida	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Pyrrhopyge pelota	1	Bolivia	Florida Museum of Natural History/USA	2021-05-03
			Celaenorrhinus cf. opalinus	1	Kenya	Florida Museum of Natural History/USA	2021-05-03
			Morvina fissimacula	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Ouleus salvina	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Cecropterus casica	1	USA	Florida Museum of Natural History/USA	2021-05-03
			Eburuncus unifasciata	1	Panama	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
			Oxynetra roscius	1	Brazil	Florida Museum of Natural History/USA	2021-05-03
			Duroca duroca	1	Brazil	Florida Museum of Natural History/USA	2021-05-03
			Charidia lucaria Aurina azines	1	Peru Guyana	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
			Mimia cf. chiapaensis	1	Ecuador	Florida Museum of Natural History/USA	2021-05-03
			Pythonides amaryllis	1	Costa Rica	Florida Museum of Natural History/USA	2021-05-03
			Zopyrion sandace Mimoniades ocyalus	1	Brazil	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
			Dalla cyprius	1	Peru	Florida Museum of Natural History/USA	2021-05-03
			Signeta flammeata	1	Australia	Florida Museum of Natural History/USA	2021-05-03
			Erynnis tuges Ectomis octomaculata	1 1	Costa Rica	Florida Museum of Natural History/USA	2021-01-25 2021-05-03
			Cecropterus confusis	1	USA	Florida Museum of Natural History/USA	2021-05-03
			Thymelicus sylvestris	6	England/UK	Wellcome Sanger Institute/UK	2021-07-21
			rii una pirus Timochares trifasciata	1	USA Costa Rica	Florida Museum of Natural History/USA Florida Museum of Natural History/USA	2021-05-03
			Autochton oryx	1	Ecuador	Florida Museum of Natural History/USA	2021-05-03
	Geometroidea	To well.					
		Family Geometridae					
	Superfamily Geometroidea		Campaea margaritaria	2	England/UK	Wellcome Sanger Institute/UK	2021-08-18
			nyariomena furcata Ectropis grisescens	4 9	Engiand/UK China	vvencome sanger Institute/UK Institute of Plant Physiology and Ecology/CHN	2021-08-18 2021-03-22
	Superfamily						
	Noctuoidea	Family Noctuidae					
			Amphipyra tragopoginis	2	England/UK	Wellcome Sanger Institute/UK	2021-02-13
			Griposia aprilina Atethmia contrace	1	England/UK	Wellcome Sanger Institute/UK	2021-09-30
			Mythimna ferrago	4 1	England/UK	Wellcome Sanger Institute/UK	2021-03-17
			Autographa pulchrina	1	England/UK	Wellcome Sanger Institute/UK	2021-04-14
			Autographa gamma	1	England/UK	Wellcome Sanger Institute/UK	2021-01-25
			i richopiusia ni Mamestra brassicae	1 2	USA Wales/UK	cornell University/USA Wellcome Sanger Institute/UK	2018-10-01 2021-01-25
			Sesamia nonagrioides	1	France	Paris-Saclay University/FRA	2021-04-13
		Family Notodontidae					
			Clostera curtula Ptilodon capucinus	3 4	England/UK England/UK	Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK	2021-04-14 2021-09-11
		Family Erebidae	Filome correction	1	England (11)	Wallaama Canzar Inskind- // W	2021 00 24
			Eilema sororculum	1	England/UK	wellcome Sanger Institute/UK	2021-09-24

			Spilosoma lubricipeda Euproctis similis Spilarctia lutea Schrapkia contactrigalic	3 3 11	England/UK England/UK England/UK England/UK	Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK	2021-02-13 2021-01-25 2021-09-18 2021-04-14
			Arctia plantaginis Lymantria monacha	3	Finland? England/UK	University of Cambridge/UK Wellcome Sanger Institute/UK	2021-04-14 2020-04-10 2021-01-25
	Superfamily		Lymanna aispar	5	зарап, спіпа	Lavai University/CAN	2021-05-04
	Domoșeolacă	Family Bombycidae	Demokra meni	2	lanan		2020 11 05
		Family Sphingidae	Laothoe populi Hyles vespertilio	7	England/UK Italy	Wellcome Sanger Institute/UK Max Planck Institute of Molecular Cell Biology and	2021-02-13 2020-01-29
		Family Saturniidae	Samia ricini	7	India*	Genetics/DEU	2020-06-20
	Superfamily Pyraloidea	Family Crambidae	Sama nem		maid		1010 00 10
	Superfamily		Chilo suppressalis Chrysoteuchia culmella	1 2	China England/UK	Huazhong Agricultural University/CHN Wellcome Sanger Institute/UK	2019-01-08 2021-07-06
	Gelechioidea	Family					
		Blastobasidae	Blastobasis lacticolella Blastobasis adustella	15 4	England/UK England/UK	Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK	2021-01-25 2021-05-19
	Superfamily Drepanoidea	Family Drepanidae					
	Superfamily Tortricoidea		Habrosyne pyritoides	1	England/UK	Wellcome Sanger Institute/UK	2021-05-11
Order Diptora		Family Tortricidae	Apotomis turbidana	1	England/UK	Wellcome Sanger Institute/UK	2021-01-25
order Diptera	Superfamily						
	Diopsoidea	Family Diopsidae	Teleopsis dalmanni	4	Malaysia	SUNY Geneseo/USA, University of Maryland/USA	2020-09-23, 2020- 10-30
	Superfamily Syrphoidea						
		Family Syrphidae	Cheilosia vulpina Melanostoma mellinum	1 3	England/UK England/UK	Wellcome Sanger Institute/UK Wellcome Sanger Institute/UK	2021-09-30 2021-09-11
	Superfamily Tephritoidea						
		Family Tephritidae	Bactrocera dorsalis	1	USA	Agricultural Research Service-USDA/USA	2014-12-03
	Superfamily Ephydroidea	Family					
		Drosophilidae	Drosophila biarmipes	7	India to SE Asia*	University of Pennsylvania/USA	2019-05-08
			Drosophila ficusphila Drosophila auraria	1 1	Taiwan Japan	Stanford University/USA University of California. Berkeley/USA	2021-04-28 2019-08-21
			Drosophila bifasciata Drosophila obscura	1 3	Japan Europe*, Serbia	University of California, Berkeley/USA National Institute of Genetics/JPN, Stanford University/USA	2019-11-15 2017-10-14, 2021- 04-28
			Drosophila ambigua Drosophila guanche Scantomyza montana	1 1 2	Serbia Canary Islands/ESP USA*	Stanford University/USA Centro Nacional de Análisis Genómico/ESP Stanford University/USA	2021-04-28 2018-09-20 2021-06-16
	Superfamily Oestroidea		Scaptomyza flava	1	USA	University of California, Berkeley/USA	2018-12-17
		Family Tachinidae	Tachina fera	1	England/UK	Wellcome Sanger Institute/UK	2021-02-13
Order Orthoptera			ł		• ·	* ·	
	Superfamily Grylloidea	Family Gryllidae					
	Superfamily		Teleogryllus occipitalis Gryllus bimaculatus	4 1	Japan Japan	Waseda university/JPN Tokushima University/JPN	2020-02-22 2021-02-13
	Eumastacoidea	Family Morabidae					
Order			Vandiemenella viatica	1	Australia	Uppsala University/SWE	2021-08-07
Hymenoptera	Superfamily						
	Ichneumonoidea	Family Braconidae					
		Family	Cotesia vestalis Cotesia vestalis bracovirus segment c35	1 1	South Korea China	Andong National University/KOR Zhejiang University/CHN	2015-03-18 2011-05-09
		Ichneumonidae	Mesochorus sp.	1	Costa Rica	University of Georgia/USA	2021-06-16
Order Coleoptera							
	Superfamily Tenebrionoidea						
		Family Pyrochroidae	Pyrochroa serraticornis	5	England/UK	Wellcome Sanger Institute/UK	2021-03-17
Order Neuroptera							
		Family Chrysopidae	Chrysoperla carnea	1	England/UK	Wellcome Sanger Institute/UK	2021-04-14
Order Siphonaptera			,		<u>,,</u>		
press	Superfamily Pulicoidea	Family Pulicidae					
Order		, i ancidac	Ctenocephalides felis	5	USA	West Virginia University/USA	2018-08-24
Phasmatodea		Family					
		Phasmatidae	Clitarchus hookeri	1	New Zealand	Landcare Research/NZL	2017-11-16

Class Arachnida								
	Order Araneae							
		Superfamily Araneoidea						
			Family Nephilidae					
				Trichonephila inaurata madagascariensis	1	Madagascar	Institute for Advanced Biosciences - Keio University/JPN	2021-07-22
			Family Linyphiidae					
				Oedothorax gibbosus	1	Belgium	Royal Belgian Institute of Natural Sciences/BEL	2021-07-22

*Original or known distribution of the species (geographical location of biosample not available).

Table S2. Average base differences per site between groups in the main clade containing CvBV Hel_c35.

40																		Γ	Γ																					0.0676
39																																							0.0620	0.0465
38																																						0.0557	0.0751	0.0623
37																												2-1									0.0503	0.0410	0.0634	0.0465
36																																				0.0363	0.0503	0.0318	0.0552	0.0350
32																																			0.0287	0.0403	0.0564	0.0340	0.0626	0.0399
34																																		0.0304	0.0288	0.0366	0.0520	0.0343	0.0605	0.0379
33																																	0.0293	0.0318	0.0296	0.0309	0.0446	0.0331	0.0554	0.0390
32																															10	50.0242	8 0.0225	t 0.0267	5 0.0225	t 0.0325	0.0465	9 0.0285	5 0.0544	9 0.0322
15									_										L											~	3 0.0246	50.0255	3 0.030	5 0.0314	5 0.0286	5 0.0354	10.0501	3 0.0345	7 0.0596	5 0.0415
<u>م</u>																													2	7 0.0238	5 0.0213	90.0236	0.028	30.0295	20.0265	3 0.0335	9 0.0474	2 0.0328	2 0.0567	30.0386
67																												8	5 0.024	1 0.027	1 0.026	7 0.0279	7 0.032	0.0338	5 0.031	5 0.0373	0.0539	5 0.0362	5 0.060	2 0.0408
87																											0	90.0278	30.0256	30.0361	10.018	10.0257	5 0.024	5 0.0250	2 0.0226	3 0.0325	5 0.0480	30.0386	0.056	10.047
77																										80	7 0.025	7 0.023	4 0.020	9 0.022	4 0.015	1 0.022.	3 0.020	9 0.021	4 0.017.	2 0.029	7 0.044t	3 0.024	1 0.053	0 0.031
56																									4	5 0.010	6 0.015	4 0.022	7 0.019	7 0.020	6 0.014	5 0.021	6 0.020	0 0.020	2 0.017	4 0.029.	8 0.043	6 0.024	1 0.053	0 0.030
5									L									L	L					9	4 0.011	7 0.013	1 0.023	18 0.023	4 0.020	0 0.022	8 0.014	60.021	60.019	9 0.021	6 0.017	10.029	60.044	2 0.024	7 0.053	7 0.034
74																			L				90	90.015	9 0.012	18 0.014	*4 0.028	60.019	15 0.015	50.014	7 0.015	3 0.017	9 0.021	15 0.022	2 0.019	8 0.027	5 0.042	0 0.028	00:050	70.031
52																						00	17 0.019	95 0.023	31 0.022	91 0.023	13 0.027	96 0.025	39 0.023	\$2 0.027	97 0.025	17 0.027	72 0.031	90 0.033	36 0.031	0.036	73 0.052	16 0.036	52 0.061	1 0.041
77																			_		22	0 0.013	0.014	90.015	5 0.018	0.015	5 0.024	1 0.015	20.018	00.023	4 0.015	4 0.021	7 0.027	90.029	6 0.023	18 0.03C	0 0.047	0.031	90.056	00.037
7									_			-								16	55 0.008	29 0.01	52 0.016	0.010	33 0.019	01 0.020	46 0.02	05 0.02	97 0.019	37 0.025	0.022	24 0.02	76 0.02	93 0.030	48 0.026	22 0.033	53 0.047	23 0.032	51 0.055	77 0.037
7	\vdash																		71	81 0.00	74 0.00	23 0.01	46 0.01	97 0.02(77 0.018	97 0.02(46 0.024	98 0.020	93 0.019	32 0.02	05 0.02(20 0.02	69 0.02	85 0.029	45 0.024	15 0.03	68 0.04	21 0.03	63 0.05	74 0.03
19	-											-						91	91 0.00	18 0.00	90.006	43 0.01	20 0.01	70 0.01	56 0.01	71 0.01	23 0.02	76 0.01	70 0.01	03 0.02	72 0.02	94 0.02	45 0.02	56 0.02	07 0.02	910.03	48 0.04	99 0.03	34 0.05	47 0.03
120	-		_			_											960	20 0.00	28 0.00	33 0.01	24 0.00	71 0.01	0.01	58 0.01	21 0.01	42 0.01	70 0.02	68 0.01	39 0.01	77 0.02	54 0.01	73 0.01	26 0.02	30 0.02	92 0.02	68 0.02	190.04	275 0.02	01 0.05	57 0.03
1	\vdash		_			_			_			_			_	060	0.010.00	118 0.0	121 0.0	120 0.0	110 0.0	166 0.0	121 0.0	175 0.0	139 0.0	152 0.03	296 0.02	127 0.03	154 0.03	192 0.0	167 0.0	185 0.03	238 0.02	243 0.02	210 0.03	277 0.02	126 0.04	297 0.0	530 0.05	370 0.0
19	-				_	_						8			143	126 0.0	170 0.0	197 0.0	199 0.0	201 0.0	193 0.0	241 0.0	131 0.0	160 0.0	140 0.0	146 0.0	209 0.0	243 0.0	203 0.0	211 0.0	181 0.0	214 0.0	247 0.0	251 0.0	212 0.0	294 0.0	448 0.0	279 0.0	528 0.0	317 0.0
1	\vdash					_								086	153 0.0	122 0.0	150 0.0	175 0.0	182 0.0	188 0.0	178 0.0	215 0.0	133 0.0	162 0.0	131 0.0	137 0.0	267 0.0	225 0.0	185 0.0	202 0.0	172 0.0	187 0.0	230 0.0	228 0.0	198 0.0	279 0.0	438 0.0	281 0.0	519 0.0	350 0.0
14	\vdash					_			_		-	_	073	0.0 060	157 0.0	130 0.0	148 0.0	174 0.0	180 0.0	185 0.0	174 0.0	216 0.0	134 0.0	159 0.0	131 0.0	143 0.0	275 0.0	225 0.0	183 0.0	2010.0	167 0.0	189 0.0	223 0.0	235 0.0	203 0.0	275 0.0	434 0.0	285 0.0	524 0.0	353 0.0
17											-	074	0.0 820	0.0860	1159 0.0	1130 0.0	148 0.0	174 0.0	180 0.0	187 0.0	176 0.0	1214 0.0	139 0.0	169 0.0	135 0.0	149 0.0	1284 0.0	1229 0.0	187 0.0	1207 0.0	177 0.0	194 0.0	1238 0.0	1239 0.0	1210 0.0	1280 0.0	1431 0.0	1293 0.0	1530 0.0	1355 0.0
17	╞	\vdash		\vdash	\vdash	\vdash		\vdash	╞		1900	J0770C	3053 0.C	J093 0.0	0154 0.C	0.127 0.C	0.149 0.C	0.177 0.C	0.182 0.C	0195 0.C	0.176 0.C	3217 0.C	0135 0.C	0164 0.C	0135 0.C	0142 0.C	3265 0.C	7228 0.0	7185 0.0	7208 0.0	0.177 0.C	7195 0.0	3234 O.C	3233 0.C	7205 0.C	7283 0.C	3441 0.C	3286 0.C	3526 0.C	3343 0.C
	╞	\mid			-			╞		0034	00310.0	0048 0.0	0028 0.0	00660.0	0128 0.0	0104 0.0	01190.0	0147 0.0	0153 0.0	0162 0.0	0143 0.0	0187 0.0	01110.	01380.0	0107 0.0	0115 0.0	0249 0.(0194 0.0	0154 0.1	0178 0.1	0146 0.1	0161 0.0	0205 0.(0208 0.(0177 0.0	0252 0.0	0412 0.0	0265 0.(0499 0.(0329 0.1
Ĭ					⊢	\vdash			0031	0061 0.	0057 0.	0073 0.	0058 0.	09600	0159 0.	0132 0.	0149 0.0	0176 0.0	0182 0.	0189 0.	0178 0.0	0215 0.1	0138 0.	0168 0.0	0136 0.0	0148 0.	0280 0.	0228 0.	0183 0.	0210 0.	0173 0.	0190 0.	0233 0.	0235 0.	0205 0.1	0278 0.	0438 0.	0289 0.	0524 0.	0355 0.
5	\vdash				\vdash			9900	00400.	00700.	00670.	00580.	00650.	00780.	0132 0.	01120.	01380.	0165 0.	01700.	01770.	0164 0.	0208 0.	01100.	01420.	.0123 0.	0128 0.	02230.	.0215 0.	0176 0.	0189 0.	.0160 0.	0182 0.	02160.	0225 0.	01870.	02720.	04270.	.0265 0.	0508 0.	0324 0.
×					-		0054	00720.	0045 0.	0074 0.	00710.	.00610.	00700.	00790.	01530.	01230.	0142 0.	0167 0.	01730.	0182 0.	0167 0.	02110.	01300.	01560.	.01210.	0138 0.	0268 0.	02190.	0175 0.	0201 0.	.0165 0.	01860.	02220.	02300.	01850.	02760.	04250.	.0275 0.	05170.	03410.
+	\vdash	\vdash			\vdash	.0029	00510	.0068 0.	.0041 0.	.00710.	0068 0.	.0058 0.	00660	00800	.0150 0.	0123 0.	01410	0167 0.	0173 0.	01790	01660.	0208 0.	0127 0.	01530	.0121 0.	0136 0.	0267 0.	.0214 0.	.0173 0.	.0199 0.	.0161 0.	.0183 0.	0218 0.	0228 0.	0189 0.	02730	04200.	.0279 0.	.0510 0.	.0332 0.
-					0014	00190	0.0044 0	0.0063 0	0.0034 0	0.0063 0	0 2900.0	00200	00560	0.0067 0	0.0142 0	0 0100	01260	01570	0.0161 0	0.0168 0	01530	02040	01200	01430	0.0108 0	01210	02600	02070	0.01660	01940	01510	01810	02150	02150	01840	02590	04160	02700	0.0483 0	03250
	F			0.0012	0.00210	0.0024 0	0.0043 0	0.0063 0	0.0035 0	0.0064 0	0.0062 0	00500	0.0058 0	0.0073 0	0.0148 0	01130	0.01310	01580	0.0165 0	01720	0.01580	0.01990	0.01210	01500	0.01150	0.0127 0	0.0262 0	0 6020.0	0.0165 0	01970	0.01560	01790	0.02110	0.02170	0.01800	0.0267 0	0.04160	0.02720	0.0508 0	0.03190
~	T		0003	0.0012 0	0.0019 0	0.0023 0	0.0044 0	0.0062 0	0.0035 0	0.0065 0	0.0061 0	0.0052 0	0.0060 0	0.0073 0	0.0145 0	0.0115 0	0.0133 0	0.0159 0	0.0165 0	0.0172 0	0.0159 0	0.0200	0.0121 0	0.0147 0	0.0115 0	0.0128 0	0.0261 0	0.0208 0	0.0165 0	0.0193 0	0.0154 0	0.0177 0	0.02120	0.0220 0	0.0183 0	0.0267 0	0.0417 0	0.0272 0	0.0506 0	0.0336 0
~		0.0003	0.0004 0	0.0013 0	0.0020	0.0024 0	0.0046	0.0063 0	0.0036	0.0066 0	0.0063 0	0.0053 0	0.0061 0	0.0074 0	0.0146 0	0.0116	0.0134 0	0.0160	0.0166	0.0173	0.0160	0.0202	0.0122 0	0.0148	0.0116	0.0129 0	0.0262 0	0.0209	0.0166	0.0194 0	0.0155 0	0.0178 0	0.0213 0	0.0221 0	0.0184 0	0.0268 0	0.0418 0	0.0273 0	0.0507 0	0.0337 0
	0.0002	0.0001	0.0002 0	0.0011 0	0.0018 0	0.0022 0	0.0043 C	0.0061 0	0.0034 C	0.0063 0	0.0061 0	0.0051 0	0.0059 0	0.0072 0	0.0146 C	0.0116 C	7.0134 C	7.0160 C	0.0166 C	0.0173 C	7.0160 C	0.0201 C	0.0122 0	0.0148 C	0.0116 C	0.0127 0	0.0262 0	0.0209	0.0164 C	0.0194 0	0.0156 0	0.0178	0.0211 0	0.0221 0	0.0182 C	0.0268 C	0.0416	0.0271 C	0.0507 0	0.0337 0
wirus	Γ			ia C		inis (nis (Γ			S St					Γ	Γ	Γ	-	Γ	Γ	Γ	ľ	Γ	Jalinus				5	5					ricia (galis (0		s,	
lis braco	ria	e e	lis	rgaritan	urcata	agopogi	raticorn		mphalis	uli	vritoide	odippe	tula	rbidana	ontinus	res	telassa	sergius	hadassa	kelita	crida	pelota	res	culum	nus cf op	vcausta	Imanni	imacula	DL	DUN	'tilio	ore	carnea	-	ops patr	staestri	icusphile	anippus	ides feli.	culatus
a vesta	ge aege	s malva	ia vesta	aea ma	omena j	ipyra tr	hroa ser	yx mori	rupa ny	hoe pop	rosyne p	iciana a	tera cun	tomis tu	anes br	temia a	hopyge	hopyge	hopyge	hopyge	hopyge	hopyge	sis lacrit	na soroi	enorrhir	eus holo	opsis da	vina fiss	us salvi	sis aurin	s vesper	sis ocyp	soperla	nina fera	dochryse	ankia co	ophila f	aus mela	ocephai	us bima
1.Cotesi	2. Parar	3.Pyrgu.	4.Cotes	5.Camp.	6.Hydric	7.Amph	8. Pyroci	9.Bomb	10.Sata	11.Laot	12.Habi	13.Fabr	14.Clost	15.Apot	6.Gind	17.Apoc	8.Pyrrt	Pyrrt	D.Pyrri	21.Pyrrl	2. Pyrrl	3.Pyrrh	4.Eme	5.Eilen	6.Cela	7.Katre	8. Teleu	Dom.00	30.0ule	31.Eme.	32.Hyle.	33.Eme	34.Chry.	S.Tach	36.Lepic	37.Schre	38.Dros	39.Danc	10.Cten	1.Gryli

Figure S1. Same Maximum Likelihood phylogeny as Fig. 1 (main text), displaying taxa names and branch support values. Distinct Lepidoptera superfamilies are represented by different colors and non-lepidopteran arthropods are represented in black. See Materials and Methods for details of the phylogenetic inference procedures.



Figure S2. Same Maximum Likelihood phylogeny as Fig. 2 (main text), displaying taxa names and branch support values. Colors correspond to geographical locations where the species were sampled (Table S1).



Data S1. Biopython script to only include sequences with > 70% (3705 bp) and to edit FASTA descriptions to contain only the hit accession number, the sequence match range and the species name.

>>> from Bio import SeqIO

```
>>> large_sequences = []
```

>>> for record in SeqIO.parse("blast_results.txt", "fasta"):

if len(record.seq) > 3705:

large_sequences.append(record)

>>> SeqIO.write(large_sequences, "large_seq.fasta", "fasta")

>>> clean_sequences = []

>>> for seq_record in SeqIO.parse("large_seq.fasta", "fasta"):

seq_record.id = ((seq_record.description.split()[0])+(" ")+

(seq_record.description.split()[1])+(" ")+

(seq_record.description.split()[2]))

seq_record.description = ("")

clean_sequences.append(seq_record)

>>> SeqIO.write(clean_sequences, "clean_seq.fasta", "fasta")

Data S2. List of sequences descriptions used in the analysis, with their accession number, match range and the species name.

HQ009558.1 Cotesia vestalis bracovirus segment c35 CAJHZN010000006.1_22939.28228_Pararge_aegeria CAJHZN010000001.1_159388.164677 Pararge aegeria CAJHZN010000001.1 169618.174907 Pararge aegeria CAJHZM010000138.1 366440.371729 Pararge aegeria CAJHZM010000104.1 145538.150827 Pararge aegeria CAJHZM010000080.1_261023.266312_Pararge_aegeria CAJHZM010000059.1_423418.428707_Pararge_aegeria CAJHZM010000045.1_4316124.4321413_Pararge_aegeria CAJHZM010000014.1_1104039.1109328_Pararge_aegeria CAJHZM010000011.1_2007535.2012824_Pararge_aegeria CAJVQM010000016.1_22902664.22907953_Pyrgus_malvae CAJHZM010000035.1_19926.25209_Pararge_aegeria CAJHZN010000005.1_288029.293318_Pararge_aegeria CAJHZM010000130.1_729237.734526_Pararge_aegeria CAJHZM010000130.1_840737.846026_Pararge_aegeria UIGX01030406.1 1.5288 Pararge aegeria CAJHZM010000036.1_7433354.7438643_Pararge_aegeria CAJHZM010000035.1_609216.614505_Pararge_aegeria CAJHZM010000035.1_617278.622567_Pararge_aegeria CAJVQM010000016.1 22913002.22918291 Pyrgus_malvae CAJHZM010000079.1 760808.766097 Pararge aegeria CAJHZM010000059.1_2265283.2270572_Pararge_aegeria CAJHZM010000271.1_10360.15649_Pararge_aegeria JZSA01007369.1_21189.25964_Cotesia_vestalis CAJHZM010000038.1 5963116.5968405 Pararge aegeria CAJHZM010000012.1_12234942.12240231_Pararge_aegeria CAJHZM01000002.1_2420860.2426149_Pararge_aegeria CAJHZM010000035.1_670716.676005_Pararge_aegeria CAJHZM010000041.1_658925.664220_Pararge_aegeria CAJHZM010000110.1_4418285.4423571_Pararge_aegeria CAJHZM010000130.1 1146594.1151884 Pararge aegeria CAJHZM010000102.1 657339.662628 Pararge aegeria CAJHZM010000092.1_631160.636449_Pararge_aegeria CAJHZM010000024.1_3952072.3957361_Pararge_aegeria CAJVWE010000011.1_3500217.3505507_Campaea_margaritaria CAJVWE010000087.1 1169967.1174606 Campaea margaritaria CAJVWD010000025.1 408819.414108 Hydriomena furcata CAJVWD010000070.1_510413.515617_Hydriomena_furcata CAJVWD010000025.1_377294.382499_Hydriomena_furcata CAJVWD010000012.1_3716744.3722060_Hydriomena_furcata CAJMZV010000185.1_996280.1001569_Amphipyra_tragopoginis CAJMZV010000218.1 4569479.4574767 Amphipyra_tragopoginis CAJOSM010000467.1 148944.154232 Pyrochroa serraticornis CAJOSM010000275.1_305597.310868_Pyrochroa_serraticornis CAJOSM010000934.1_7793.12723_Pyrochroa_serraticornis CAJOSL010000003.1_780653.785583_Pyrochroa_serraticornis CAJOSM010000568.1_196017.200944_Pyrochroa_serraticornis BHWX01000001.1 13148640.13153928 Bombyx mori DWHE01006833.1_1.5238_Satarupa_nymphalis CAJNAB010000311.1_202758.208045_Laothoe_populi CAJNAB010000435.1 67852.73189 Laothoe populi BHWX01000007.1 11846872.11852161_Bombyx_mori CAJNAB010000107.1_670398.675633_Laothoe_populi CAJNAB010000260.1_341534.346834_Laothoe_populi CAJRBD010001370.1_98905.104202_Habrosyne_pyritoides CAJQEY01000087.1_6085973.6091264_Fabriciana_adippe CAJNAB010000869.1_72668.77560_Laothoe_populi CAJQFY010000135.1_335220.340495_Clostera_curtula CAJQFY010000347.1 1.5279 Clostera curtula CAJQFY010001131.1 62176.67434 Clostera curtula BHWX01000022.1_752045.757343_Bombyx_mori CAJNAB010000154.1 1044455.1049578 Laothoe populi CAJNAB010001387.1 170726.175895 Laothoe populi

CAJHUU010000108.1_3275763.3280672_Apotomis_turbidana DWQB01000133.1 862.6189 Gindanes brontinus DVYD01000130.1 683.5891 Apodemia ares DWE001003961.1 926.5870 Pyrrhopyge telassa DVQA01000780.1_1045.5981_Pyrrhopyge_sergius DWOT01000734.1_76.5034_Pyrrhopyge_hadassa DVQD01000579.1_78.4419_Pyrrhopyge_kelita DWEP01000053.1_1589.6485_Pyrrhopyge_crida DWSZ010000156.1 1.5209 Emesis lacrines CAJZCT010000117.1 1734641.1739951 Eilema sororculum DWLK01000137.1_1.4747_Celaenorrhinus_cf._opalinus DWMK01000004.1_19.5148 Katreus_holocausta DWFE01002627.1_226.5042 Pyrrhopyge_pelota NLCU02019152.1 17889.23172 Teleopsis_dalmanni JACTOK010041713.1_346.5639_Teleopsis_dalmanni JACTOK010037361.1_40126.45029_Teleopsis_dalmanni JACTOK010035260.1_62.5356_Teleopsis_dalmanni DWP001000177.1_2362.6863_Morvina_fissimacula DWMC01000015.1_734.5673_Ouleus_salvina DVZB010000096.1 227.5537 Emesis aurimna WUWR01000078.1 21074.25328 Hyles vespertilio DWSV010000232.1_290.4623_Emesis_ocypore CAKAJF010000605.1 396300.401682 Griposia aprilina DWQE01036201.1 1.5321 Parnassius apollo DWTE010001035.1 1801.6867 Parnassius imperator DWHT01000605.1 4715.9222 Parnassius smintheus CAJOSY010001674.1_384948.390327_Atethmia_centrago CAJOSY010000266.1_43501.48893_Atethmia_centrago CAJOSY010000707.1_37529.42796_Atethmia_centrago CAJOSY010001674.1_347256.352311_Atethmia_centrago CAJQZP010000349.1 456251.461622 Parnassius apollo CAJQZP010001011.1_5021097.5026488_Parnassius_apollo CAJQZP010000971.1_1549.6922_Parnassius_apollo CAKAIX010000158.1_18940.23791_Cheilosia_vulpina CAJUUQ010000015.1_15920890.15926267_Mythimna_ferrago RSAL01000011.1 2782940.2788264_Chilo_suppressalis CAJNAL010000544.1 199300.204585 Spilosoma lubricipeda CAJNAL010000084.1_2628222.2633568_Spilosoma_lubricipeda CAJNAL010000037.1_461673.466985_Spilosoma_lubricipeda CAJQZP010000220.1_17370713.17376024_Parnassius_apollo DWKJ01000024.1 3286.8454 Cecropterus casica DWOG01000018.1 2139.7331 Mylon lassia DWPY01000040.1_1418.6608_Eburuncus_unifasciata DWAC01000453.1_1451.6623_Curetis_bulis DWQQ01006667.1_124.5302_Oxynetra_roscius DWES01000095.1_1328.6454_Duroca_duroca DVYP01001143.1 1.4807 Emesis heterochroa DWEU01002184.1 201.5367 Charidia lucaria DWHR010001274.1_81.5252_Protesilaus_protesilaus DWJD01000127.1_1144.5908_Aurina_azines DWKU01000031.1_2444.7269_Mimia_cf._chiapaensis DWQC01000345.1 112.4922 Pythonides amaryllis DWIN01000090.1 544.5362 Zopyrion sandace DWDV01005608.1_2783.7956_Mimoniades_ocyalus DWHJ01000209.1_1124.5942_Dalla_cyprius FAUM01003789.1_1.4923_Heliconius_wallacei JABMBV010043498.1_1630.6800_Mesochorus_sp. CAJQFV010000391.1 2439220.2444392 Autographa pulchrina DWOH01000281.1_282.5439_Signeta_flammeata DWHW01001352.1_2.5076_Zerynthia_polyxena CAJHUW010000011.1_6732089.6737217_Erynnis_tages CAJHUY010000088.1_316293.321449_Cyaniris_semiargus CAJHUY010000030.1 1176489.1181646 Cyaniris_semiargus CAJNAE010000125.1 547677.552484 Lysandra coridon CAJOSV010000025.1_2708796.2713947_Lycaena_phlaeas CAJNAE010000284.1_175958.180754_Lysandra_coridon CAJHUN010000073.1_4061456.4066594_Aricia_agestis CAJHUY010000065.1_1171637.1176785_Cyaniris_semiargus

CAJOSR010000320.1_1406546.1411686_Lysandra_bellargus CAJMZN010000024.1_1507394.1512552_Vanessa_cardui CAJMZN010000124.1 3678898.3684055 Vanessa cardui JFBF01000201.1 310245.315026 Bactrocera dorsalis BLXV01000007.1_15866984.15871803_Samia_ricini DWJG01000135.1_5347.9705_Ectomis_octomaculata BLXV01000001.1_16639069.16643890_Samia_ricini BLXV01000007.1_15019349.15023900_Samia_ricini BLXV01000007.1 13697906.13703072 Samia ricini BLXV01000004.1 17631991.17637154 Samia ricini BLXV01000003.1_13005615.13010779_Samia_ricini DWTU01020815.1_2.5164_Cecropterus_confusis PPHH01001895.1_1995326.2000041_Trichoplusia_ni CAJMZN010000221.1_54458.59610_Vanessa_cardui CAJMZN010000118.1 342983.348103 Vanessa cardui SZUW01001905.1_136961.141302_Drosophila_biarmipes SZUW01001905.1_119326.123659_Drosophila_biarmipes SZUW01001905.1_128145.132475_Drosophila_biarmipes SZUW01001905.1_123735.128069_Drosophila_biarmipes SZUW01001905.1 132551.136885 Drosophila biarmipes SZUW01001905.1_114917.119250_Drosophila_biarmipes SZUW01001905.1_141332.145691_Drosophila_biarmipes BLXV01000003.1_10750556.10755705_Samia_ricini JADLOZ010000452.1 2509044.2514246 Ectropis grisescens JADLOZ010000461.1 14720351.14725555 Ectropis_grisescens JADLOZ010000163.1 10872915.10878112 Ectropis grisescens JADLOZ010000001.1_13982744.13987947_Ectropis_grisescens CAJHUX010000250.1_682937.687759_Euproctis_similis CAJHUX010000124.1_2633936.2638758_Euproctis_similis CAJHUX010000174.1_1512495.1517317_Euproctis_similis CAJHZL010000021.1_134729.139550_Mamestra_brassicae CAJHZL010000102.1_1.4272_Mamestra_brassicae CAJHWA010000005.1_10193835.10198656_Autographa_gamma CAJHWT010000024.1_864030.868851_Pieris_rapae CAJHWT010000025.1_8267413.8272232_Pieris_rapae CAJHWT010000037.1 2774121.2778939 Pieris rapae CAJZHO010000673.1 459067.463887 Spilarctia lutea CAJZHO010000165.1_1345674.1350495_Spilarctia_lutea CAJZHO010000105.1_881927.886748_Spilarctia_lutea CAJZHO010000219.1_260539.265361_Spilarctia_lutea CAJZHO010000021.1 867128.871949 Spilarctia lutea CAJZHO010000673.1 453918.458739 Spilarctia lutea CAJZHO010000143.1_1190670.1195491_Spilarctia_lutea JADLOZ010000086.1_11071347.11076164_Ectropis_grisescens JADLOZ010000127.1_29382932.29387733_Ectropis_grisescens CAJZH0010000233.1_432824.437645_Spilarctia_lutea CAJZHO010000266.1_548611.553432_Spilarctia_lutea CAJZHO010000274.1_1798316.1803137_Spilarctia_lutea CAJZH0010000256.1_664353.669174_Spilarctia_lutea CAJZBM010000107.1_62863.67683_Ptilodon_capucinus CAJZBM010000088.1_746415.751235_Ptilodon_capucinus CAJZBM010000074.1_502408.507228_Ptilodon_capucinus CAJZBM010000044.1 1818003.1822823 Ptilodon capucinus JADLOZ010000416.1_12189676.12194702_Ectropis_grisescens JADLOZ010000492.1_6103834.6108866_Ectropis_grisescens JAHESG010000032.1_186837.191985_Dryas_iulia JAHESG010000017.1_6164655.6169460_Dryas_iulia JAHESG010000029.1_7559667.7564471_Dryas_iulia JAHESG010000004.1_15479610.15484420_Dryas_iulia JAHESG010000016.1_3595041.3599846_Dryas_iulia JAHESG010000029.1_3085886.3090704_Dryas_iulia JAHESG01000009.1_9945615.9950420_Dryas_iulia CAJMZN010000075.1_4313759.4318266_Vanessa_cardui CAJQFW010000081.1_700146.704641_Chrysoperla_carnea CAJMZR010001290.1_460177.465027_Tachina_fera DWAK01008481.1_1.4253_Lepidochrysops_patricia JAFELO010000258.1_750129.755321_Eumaeus_atala JAFELO010000541.1_77968.82990_Eumaeus_atala

JAFEL0010000139.1_388908.393951_Eumaeus_atala JAFEL0010001003.1_188367.193567_Eumaeus_atala JAFEL0010000370.1 1179360.1184501 Eumaeus atala JAFELO010000151.1_120296.125456_Eumaeus_atala JAFEL0010001549.1_174743.179937_Eumaeus_atala CAJQGC010000043.1_973037.977303_Schrankia_costaestrigalis JAECXK010000002.1_87239.91306_Drosophila_ficusphila JAAAKD010007556.1_22285728.22290784_Danaus_melanippus VNJW01002308.1 9702.14301 Drosophila auraria QVA001001492.1 48901893.48906517 Ctenocephalides felis QVA001000696.1_15497.20117_Ctenocephalides_felis QVA001001492.1_52438176.52442793_Ctenocephalides_felis QVA001002597.1_29873.34493_Ctenocephalides_felis QVA001001409.1 9024.13640 Ctenocephalides felis CAJVQL010000107.1 930262.935515 Thymelicus sylvestris CAJVQL010000166.1_2342372.2347626_Thymelicus_sylvestris CAJVQL010000077.1_2140588.2145740_Thymelicus_sylvestris CAJVQL010000077.1_2145744.2150856_Thymelicus_sylvestris CAJVQL010000007.1_627957.633205_Thymelicus_sylvestris CAJVQL010000185.1 2083593.2088691 Thymelicus sylvestris CAJNAI01000002.1_5361683.5366998_Nymphalis_polychloros CADEBC010000506.1_2189273.2194586_Arctia_plantaginis CADEBD010000226.1_11524308.11529654_Arctia_plantaginis CADEBD010000226.1_5511712.5517026_Arctia_plantaginis BMAV01001078.1 255549.259048 Trichonephila_inaurata_madagascariensis BLKR01001327.1 272646.277539 Teleogryllus occipitalis BLKR01000082.1_620997.625590_Teleogryllus_occipitalis BLKR01001662.1_236479.241317_Teleogryllus_occipitalis BLKR01000118.1_545117.549975_Teleogryllus_occipitalis WIOZ01000119.1 47745.51620 Drosophila_bifasciata BOPP01000152.1 2643530.2648840 Gryllus bimaculatus JADODW010002850.1_173738.178412_Vandiemenella_viatica JAFNEN010000473.1_151816.155317_Oedothorax_gibbosus DWOJ01002358.1_82.4857_Piruna_pirus DWHR010000359.1 2831.7604 Protesilaus protesilaus CAJHUO010000233.1 1599422.1604185 Blastobasis lacticolella CAJHUO010000267.1 2646904.2651683 Blastobasis lacticolella DWIA01002199.1_1562.6349_Archon_apollinus CAJHUO010000037.1_3811342.3816132_Blastobasis_lacticolella CAJHUO010000261.1_1929974.1934751_Blastobasis_lacticolella CAJHUO010000046.1 898348.903124 Blastobasis lacticolella CAJHUO010000427.1 199617.204394 Blastobasis lacticolella CAJHUO010000318.1_112116.116892_Blastobasis_lacticolella CAJHUO010000318.1_123543.128316_Blastobasis_lacticolella CAJHUO010000345.1_691171.695944_Blastobasis_lacticolella CAJHUO010000401.1_1220232.1225032_Blastobasis_lacticolella CAJHUO010000228.1 159363.164129 Blastobasis lacticolella CAJHUO010000331.1_164805.169582_Blastobasis_lacticolella CAJHU0010000118.1_176842.181594_Blastobasis_lacticolella CAJHUO010000212.1_73386.78146_Blastobasis_lacticolella CAJVQM010000028.1_32499509.32504298_Pyrgus_malvae CAJHUO010000203.1_1777648.1782415_Blastobasis_lacticolella CAJSMA010000017.1 23489.28276 Blastobasis adustella CAJSMA010000011.1_3727558.3732195_Blastobasis_adustella CAJSMA010000156.1_848419.853213_Blastobasis_adustella JADLOZ010000397.1_7224651.7229441_Ectropis_grisescens CAJSMA010000114.1_315849.320627_Blastobasis_adustella BDQP01000130.1 301209.305884 Drosophila obscura JAECWW010000026.1_245028.249259_Drosophila_obscura CAJUU0010000047.1_3826225.3831009_Chrysoteuchia_culmella CAJUU0010000001.1_1104634.1109407_Chrysoteuchia_culmella JAECWW010000075.1_438317.443049_Drosophila_obscura CAJHZW010001009.1 110301.115094 Lymantria monacha CAJHZW010000306.1 1864909.1869687 Lymantria monacha CAJHZW010000292.1_841817.846553_Lymantria_monacha CAJHZW010000280.1_101693.106468_Lymantria_monacha JAFEKU010001978.1_7197.11978_Lymantria_dispar JAFEKT010001198.1_1395550.1400311_Lymantria_dispar

JAFEKT010002896.1_71452.76223_Lymantria_dispar CAJHZW010001466.1_82990.87783_Lymantria_monacha CAJHZW010000168.1_160361.165147_Lymantria_monacha JAFEKT010001681.1_858485.863270_Lymantria_dispar JAFEKU010001203.1_63466.68240_Lymantria_dispar JAECWS01000095.1_10811294.10815056_Drosophila_ambigua DWOM01000141.1_313.5042_Timochares_trifasciata DWLJ01000697.1_3.4421_Autochton_oryx JAEIGR010000030.1_263589.268325_Scaptomyza_montana JAEIGR010000013.1_846018.850751_Scaptomyza_montana RKRM01002430.1_4632.9349_Scaptomyza_flava CAJZBV01000028.1_2548868.2553642_Melanostoma_mellinum CAJZBV01000038.1_234267.239006_Melanostoma_mellinum JADWQK01000026.1_7378926.7383667_Sesamia_nonagrioides OUUW0100028.1_22450.27113_Clitarchus_hookeri