Antibacterial-containing dental adhesives' effects on oral pathogens and on *Streptococcus mutans* biofilm: Current perspectives

CAROLINA BOSSO ANDRÉ, DDS, PHD, DANIEL C.N. CHAN, DDS, MSC, PHD & MARCELO GIANNINI, DDS, PHD

ABSTRACT: Purpose: To describe the literature findings regarding commercially available antibacterial-containing dental adhesives and the futures perspectives of this field. **Results:** High-risk caries patients could yield benefits from restorative materials containing antibacterial properties in order to reduce the recurrent caries formation. Dental adhesives with antibacterial agents may reduce restoration replacement, as recurrent caries is still one of the major reasons for replacing a resin restoration. Literature results of three commercially available adhesives: Gluma 2Bond, Clearfil SE Protect and Peak Universal Bond, containing glutaraldehyde, MDPB and chlorhexidine, respectively indicates that Clearfil SE Protect seems to have better results against oral pathogens and on *Streptococcus mutans* biofilm. Besides the promising findings, clinical studies are still necessary in order to validate the clinical efficacy when exposed to a more complex environment and the long-term effect of either commercially available materials, experimental antibacterial monomers or antibacterial incorporations. As a suggestion of this article and according to the current scientific trends in this specific field, future directions should focus on restorative materials with therapeutic components targeting the virulence factors of cariogenic biofilm with minimal toxicity and side effects, and long-term action. (*Am J Dent* 2018;31:(Sp Is B):37B-41B).

CLINICAL SIGNIFICANCE: Antibacterial-containing dental adhesives may have therapeutic effects, working as an additional source to reduce recurrent caries development in patients with high-risk of caries, and consequently the reduction in restoration replacements.

Dr. Marcelo Giannini, Department of Restorative Dentistry, Operative Dentistry Division, Piracicaba Dental School, State University of Campinas, Av. Limeira, 901, Bairro Areião, Zip Code: 13414-903, São Paulo, Brazil. E-E: giannini@fop.unicamp.br

Introduction

Practitioners have spent a lot of time replacing or performing resin restorations due to recurrent caries formation, tooth fractures, restoration fractures, loss of marginal integrity or lack of marginal sealing and non-carious cervical lesions, such as erosion, abrasion and abfraction.¹⁻⁶ To restore small and middle-size cavities, resin-based composites have been used due to their outstanding esthetic appeal⁷ and excellent adhesive strength to dentin and enamel in combination with bonding agents. Several dental adhesive systems are commercially available for clinical use and are classified according to their application mode.⁸

Etch-and-rinse adhesives can be applied in two or three steps and their main characteristic is the application of an adhesive after phosphoric acid etching in wet demineralized dentin. Threestep etch-and-rinse adhesives use a primer, which is generally an aqueous solution containing HEMA (2-hydroxyethyl methacrylate), while two-step etch-and-rinse adhesives present a combination of primer and bonding resin in a single bottle, which contains organic solvents, such as alcohol or acetone.⁹

Self-etch adhesives are applied in one or two steps and the main compositional characteristic is the presence of functional monomers, which are responsible to etch and infiltrate into mineralized tooth structures. Two-step self-etch adhesives use an acidic primer followed by a bonding or hydrophobic resin. Single-step or all-in-one self-etching systems are user-friendly bonding agents; however, many studies have criticized this category of adhesives regarding clinical durability.¹⁰

Besides resin monomers, chemical initiators and organic solvents, dental adhesives may contain filler, fluoride, desensi-

tizing or antimicrobial agents.⁸ Many compounds and substances, such as triclosan, dimethylaminododecyl methacrylate (DMADDM), silver nanoparticles, doxycycline-encapsulated halloysite nanotube, zinc methacrylate, methacryloxylethyl cetyl dimethyl ammonium chloride (DMAE-CB) have been incorporated into dental bonding agents in order to promote antibacterial activity.¹¹⁻¹⁵

Antibacterial properties in adhesive systems or composites are considered a viable option to reduce the bacterial colonization around dental restorations, prevent recurrent caries by suppressing biofilm formation and acid production, and thereby reduce restoration replacement.¹⁶⁻¹⁸ Although extensive research on antibacterial agents incorporated into dental adhesives or antibacterial monomer syntheses is available, just a few commercial adhesives contains antimicrobial agents, such as Clearfil SE Protect^a (methacryloyloxydodecylpyridinium bromide, MDPB), Gluma 2Bond^b (glutaraldehyde) and Peak Universal Bond^c (chlorhexidine).^{19,20}

The most well-known adhesive with antimicrobial activity is Clearfil SE Protect, a two-step self-etch system that contains MDPB in the primer solution. MDPB is a polymerizable quaternary ammonium methacrylate that copolymerizes with other adhesive monomers and disrupts the bacterial cell membrane when bacterium is in direct contact with the adhesive layer (by contact of the negatively charged bacteria with positively charged quaternary ammonium).^{21,22} Antibacterial monomers that copolymerize with other adhesive monomers may provide long-term antibacterial activity.²³ After the development of MDPB, several other monomers with quaternary ammonium have been synthesized and incorporated into dental materials as antibacterial agents.²⁴⁻²⁶ Despite the increased development and evaluation of experimental antibacterial monomers, containing or based on substances with broad antimicrobial action, as antibacterial agents, the focus of this article is to discuss commercially available dental adhesives and their future perspectives.

Regarding the commercially available dental adhesives containing antibacterial agents, using a direct contact method the Clearfil SE Protect was tested against four facultative bacteria and four strict anaerobic microorganisms and had a bactericidal effect against Fusobacterium nucleatum after 10 minutes, against Streptococcus mutans, Porphyromonas gingivalis, Prevotella intermedia and Prevotella nigrescens after 30 minutes and against Staphylococcus aureus and Lactobacillus casei after 24 hours.¹⁹ Another study²⁰ showed antimicrobial effects against oral pathogens by inhibition halo method and the decrease of viability of S. mutans biofilm grown on top of the adhesive layer, compared to Clearfil SE Bond. The same adhesive was tested in simulated Class I restorations and a significant reduction in formation of biofilm of S. mutans was also achieved, when compared to an adhesive without antibacterial agent.²⁷ In situ studies^{28,29} indicate that Clearfil SE Protect is capable of controlling the caries progression in enamel at the restoration interface under conditions of high cariogenic challenge, compared to an adhesive with fluoride in its composition. Likewise, an in vivo study³⁰ showed a reduction in caries formation around brackets after 30 days compared to conventional methods. In addition, it was reported³¹ that *E. faecalis* and *S. mutans* were not able to adapt to MDPB, which may suggest a lower risk of producing drug resistance.

A two-step etch-and-rinse adhesive, Gluma 2Bond, contains 5% glutaraldehyde, which is a desensitizing and strong antibacterial agent.³²⁻³⁴ This adhesive showed bactericidal contact activity against Staphylococcus aureus, Enterococcus faecalis, Lactobacillus casei, Streptococcus mutans, Prevotella nigrescens and Fusobacterium nucleatum after 24 hours and against Porphyromonas gingivalis and Prevotella intermedia after 1 hour.¹⁹ The qualitative analysis of S. mutans biofilm using scanning electron microscopy showed a decrease of colonies when using Gluma 2Bond compared to a similar adhesive without glutaraldehyde; a result that was confirmed by colony counting.²⁰ Another study³⁵ also investigated dental adhesives containing glutaraldehyde (Gluma Primer^b and Syntac Classic System^c) and glutaraldehyde present in Gluma Primer^b and Syntac Adhesive^c appears to be effective against infected dentin. An in vivo study 36 also showed the dentin disinfecting capacity of a glutaraldehyde-containing adhesive compared to an adhesive without antibacterial agent. Glutaraldehyde-containing bonding agents have been criticized due to toxicity and mutagenic potential of this type of aldehyde. These effects were already described.^{37,38}

Peak Universal Bond contains 0.2% chlorhexidine di(acetate), which is a cationic polybiguanide, bisphenol component containing chlorine that reacts with the negatively charged microbial cell surface, destroying its membrane. Chlorhexidine has a wide spectrum of action against grampositive and gram-negative organisms, facultative, anaerobes, aerobes and fungi.^{39,41} This two-step etch-and-rinse adhesive demonstrated bactericidal contact activity only for strict anaero-

bic microorganisms (Porphyromonas gingivalis, Prevotella intermedia, Prevotella nigrescens and Fusobacterium nucleatum after 24 hours).¹⁹ No effect against Streptococcus mutans biofilm was observed for this adhesive, compared to the same adhesive without chlorhexidine. However, these adhesives (with and without chlorhexidine) presented a reduction in biofilm of S. mutans similar to Clearfil SE Protect, which implies that other components, such as adhesive monomers and solvents may have antibacterial activity.²⁰ These results suggest that chlorhexidine may stay trapped in the polymer chain, without the release properties.^{19,20} In another study,⁴² Peak Universal Bond presented a lower S. mutans biofilm formation compared to the same adhesive version without chlorhexidine; however the specimen preparation was different and the incubation time was lower. In addition, this non-light-cured adhesive presented an inhibition halo against some bacteria, suggesting that it may work as a cavity disinfectant.²⁰ Also, for Peak Universal Bond, an inhibition halo for S. mutans was identified when it was not light-cured.43

The complex interactions between the specific oral bacteria, salivary constituents, dietary carbohydrate, and tooth surface modulates the transition from a condition of health to a diseased state by the establishment of cariogenic biofilms and consequently surface cavitation by acid dissolution, resulting in dental caries.⁴⁴ Regarding the role of the aforementioned bacteria at the pathogenesis of caries disease, S. mutans is considered the main pathogen involved in caries formation.⁴⁵ S. *mutans* is not always the most predominant at the initial colonizing community, however the primary role of S. mutans resides with its ability to assemble an insoluble polymeric matrix, forming the core of the matrix-scaffold in cariogenic biofilms.⁴⁶ Besides the extracellular polysaccharides production, the virulence of S. mutans is also associated to the production of weak acids from sugars, to adapt to large fluctuations in pH, oxygen tension and nutrient availability.^{47,48}

Other microorganisms present in the complex oral microbiota also play an important role in caries disease development and progression.²⁰ *Lactobacillus casei* is an acidogenic and acid tolerant bacteria that can grow and survive in an acidic environment;^{49,50} *Staphylococcus aureus* is found in individuals with aggressive periodontitis⁵¹ and *Enterococcus faecalis* is associated with chronic periodontitis and frequently is the only species that persists in endodontically treated teeth.^{20,52,53}

Strict anaerobic bacteria are more related to periodontal disease and can be found in cariogenic biofilm around the gingival margin.⁵⁴ Due to further accumulation of biofilm, the number of obligatory anaerobic bacteria increase, changing the antimicrobial biofilm composition from streptococcus-dominated to *Actinomyces* spp. that is involved in root caries, and *P. gingivalis* involved and periodontal disease.^{19,54,55} *P. intermedia* is also a periodontal pathogen found in patients with early periodontitis, advanced periodontitis, and acute necrotizing ulcerative gingivitis.^{56,57} *P. nigrescens* also plays a role in the pathogenesis of periodontal disease, gingivitis and some odontogenic infections.^{58,59} *F. nucleatum* is frequently associated with periodontal diseases and is commonly found in human dental plaque with a crucial role in plaque development.^{19,60-62}

Clearfil SE Protect, Gluma 2Bond and Peak Universal Bond

present dentin bond strengths around 40 MPa and did not differ among them when the specimens were analyzed after artificial saliva storage for 1 year. These adhesives form a hybrid layer and resin tags, which represent the bonding mechanism of contemporary bonding agents. Thus, the presence of antimicrobial components in the composition of adhesives seems to not interfere in the bond strength and bonding mechanism.^{19,20}

Bonding agents containing antibacterial compounds are indicated for patients with very poor oral health, due to the high probability of recurrent caries development. Elderly patients with greater incidence of root caries and patients who have limitations to promote their own oral hygiene may also benefit when restorations are performed with materials containing antibacterial agents. Although some advantages have been extensively reported in the dental literature, there are concerns regarding the side effects produced by antibacterial agents and little clinical evidence that supports the in vitro findings has been reported.^{24,33} Also, the antibacterial activity in multispecies biofilm may be lower compared to results with planktonic bacteria,⁶³ considering that the bacteria are protected by a diffusion barrier, the extracellular matrix.⁶⁴ Another concern regarding in vitro tests remains on the interaction between the adhesive layer and the saliva pellicle. Some publications suggested that the saliva pellicle could attenuate antibacterial properties of underlying surfaces.^{65,66} However, the antibacterial effect of Gluma 2Bond and Clearfil Protect Bond was expressed in the biofilm of S. mutans, even covered with clarified saliva.²⁰

One of the major side effects related to substances with broad antimicrobial spectra into restorative materials is the oral health resident bacterial interference and the promotion of bacterial resistance, producing undesirable outcomes on oral health.⁶⁷ In order to reduce these side effects, the incorporation of natural products are been proposed, as a result of the lower probability of producing bacterial resistance. Natural products are considered a potential alternative approach to the current chemotherapeutic strategies, owing to the fact that natural products are a safer technology, biologically and environmentally, when compared to compounds synthetized by chemical or physical methods.^{68,69}

Propolis is a natural product composed of a resinous substance collected by *Apis mellifera* bees from various plant sources. It is considered a nontoxic natural product with a complex chemical composition and exhibits a wide range of biological activities, including antimicrobial, anti-inflammatory, anesthetic, and cytostatic properties.^{70,71} Two components were isolated from a Brazilian propolis, apigenin and tt-farnesol, and may represent an important alternative to current antibacterial agents, seeing that they can reduce the expression of virulence of *S. mutans* without necessarily suppressing the resident oral microbiota.⁶⁸

Apigenin (4',5,7-Trihydroxyflavone) is a potent inhibitor of water-insoluble glucan synthesis (inhibitor of glucosyltrans-ferases B and C), while tt-farnesol (trans,trans-3,7,11-trimethyl-2,6,10-dodecatrien-1-ol) changes the permeability and fluidity of the cell membrane by its lipophilic properties, affecting its glycolytic activity, production-secretion of glucosyltransferases and acidurance.^{72,73} They can be used separately or together, and seem to be more effective in the presence of fluoride.⁶⁸

One study⁶⁷ incorporated these components into comercial bonding agents that contain fluoride (Patent: BR 10 2014 024497 5): Clearfil S3 Bond Plus, a single-step self-etch adhesive and Optibond S,^e a two-step etch-and-rinse adhesive. The results were promising and may represent a novel alternative to decrease the cariogenicity of the biofilm around dental restorations, without suppressing the target microorganism. The addition of apigenin or appigenin and tt-farnesol to Clearfil S3 Bond Plus were more efficient regarding the reduction of virulence of *S. mutans* compared to Optibond S and they did not interfere on the adhesion mechanism of both adhesives.⁶⁷ Clearfil S3 Bond Plus containing apigenin reduced the amount of insoluble and intracellular polysaccharides of *S. mutans* biofilm grown for 5 days on top of the adhesive layer covered with clarified saliva.⁶⁷

The new approach of incorporating anti-caries agents that are less likely to induce bacterial resistance into restorative materials could yield benefits in terms of enhanced durability of composite restorations, mainly in areas where biofilms accumulate, such as the interproximal and cervical regions of the teeth, by targeting the main virulence factors of *S. mutans* biofilm, namely the insoluble polysaccharides and intracellular polysaccharides.⁶⁷ The reduction of both polysaccharides could affect the *S. mutans* ability to colonize the tooth surface and become the dominant bacteria and expressing it's virulence.⁷⁴ Although this approach is considered promising, further studies are necessary to clarify the effect on multispecies biofilm, on long-term action, and in vivo conditions (animal studies or long-term clinical trials).

Following the same trend of incorporating natural products that have antibacterial properties into dental materials, chitosan and Epigallocatechin-3-gallate (EGCG) were also investigated when added to dental adhesives. The antibacterial activity of chitosan remains on the interaction between the positively charged chitosan and the negatively charged bacteria cell surface, causing the cell wall rupture.^{75,76} When added to dental adhesives the antibacterial effect has been reported.76,77 Conversely one study⁷⁸ showed the absence of antibacterial activity of chitosan into a dental adhesive. EGCG, a flavonoid produced by Camellia sinensis plant (green tea), may be capable of suppressing gtf B, C, and D gene expression, disrupting S. mutans biofilm formation. This compound was able to express antibacterial activity when incorporated into dental adhesives in some concentrations.⁷⁹ In addition, the increased research in natural products brings new alternative formulations to oral health care, including antibacterial, antifungal, and anti-caries properties, still poorly explored in the dental biomaterials field.

Conclusions and future perspectives

Dental adhesive systems containing antibacterial or anticaries agents show remarkable results against oral pathogens in in vitro studies. MDPB containing adhesives had greater results and is extensively explored in the dental literature. These antibacterial findings suggest a favorable indication of antibacterial dental adhesives for patients with high caries risk. Incorporation of natural products into restorative materials that can act on the *S. mutans* virulence factors can be considered a new approach in order to reduce recurrent caries formation, without killing the target organism. Besides the promising findings, clinical studies are still necessary in order to validate the clinical efficacy when exposed to a more complex environment and the long-term effect of either commercially available materials, experimental antibacterial monomers or antibacterial incorporations. Future directions in research should focus on restorative materials with therapeutic components targeting the virulence factors of cariogenic biofilm with minimal toxicity, side effects, and with long-term action.

- a. Kuraray Noritake Dental Inc., Tainai City, Niigata, Japan.
- b. Heraeus Kulzer GmbH, Hanau, Hessen, Germany.
- c. Ivoclar Vivadent, Schaan. Liechtenstein.
- d. Ultradent Products Inc., South Jordan, UT, USA.
- e. Kerr Corp., Orange, CA, USA.

Acknowledgements: The authors thank the financial support from Brazilian Financial Agencies: FAPESP: 2010/13599-0, 2011/17841-2 and 2014/17543-0; CNPq: 140698/2013-2. Funding sources were not involved in study design; in the collection, analysis and interpretation of data; in the writing of the report; or in the decision to submit the article for publication.

Disclosure statement: The authors declared no conflict of interest.

Dr. André is a postdoctoral researcher, Dr. Giannini is Associate Professor, Department of Restorative Dentistry, Piracicaba Dental School, State University of Campinas, São Paulo, Brazil. Dr. Chan is Professor, Department of Restorative Dentistry, School of Dentistry, University of Washington, Seattle, Washington, USA.

References

- Gordan VV, Garvan CW, Blaser PK, Mondragon E, Mjör IA. A long-term evaluation of alternative treatments to replacement of resin-based composite restorations: Results of a seven-year study. J Am Dent Assoc 2009;140:1476-1484.
- Demarco FF, Corrêa MB, Cenci MS, Moraes RR, Opdam NJ. Longevity of posterior composite restorations: Not only a matter of materials. *Dent Mater* 2012;28:87-101.
- Opdam NJ, van de Sande FH, Bronkhorst E, Cenci MS, Bottenberg P, Pallesen U, Gaengler P, Lindberg A, Huysmans MC, van Dijken JW. Longevity of posterior composite restorations: A systematic review and meta-analysis. J Dent Res 2014;93:943-949.
- Makishi P, Thitthaweerat S, Sadr A, Shimada Y, Martins AL, Tagami J, Giannini M. Assessment of current adhesives in class I cavity: Nondestructive imaging using optical coherence tomography and microtensile bond strength. *Dent Mater* 2015;31:e190-e200.
- Nedeljkovic I, Teughels W, De Munck J, Van Meerbeek B, Van Landuyt KL. Is secondary caries with composites a material-based problem? *Dent Mater* 2015;31:e247-e277.
- Pena CE, Rodrigues JA, Ely C, Giannini M, Reis AF. Two-year randomized clinical trial of self-etching adhesives and selective enamel etching. *Oper Dent* 2016;41:249-257.
- Ferracane JL. Resin-based composite performance: Are there some things we can't predict? *Dent Mater* 2013;29:51-58.
- Van Landuyt KL, Snauwaert J, De Munck J, Peumans M, Yoshida Y, Poitevin A, Coutinho E, Suzuki K, Lambrechts P, Van Meerbeek B. Systematic review of the chemical composition of contemporary dental adhesives. *Biomaterials* 2007;28:3757-3785.
- Pashley DH, Tay FR, Breschi L, Tjaderhane L, Carvalho RM, Carrilho M, Tezvergil-Mutluay A. State of the art etch-and-rinse adhesives. *Dent Mater* 2011;27:1-16.
- Giannini M, Makishi P, Ayres AP, Vermelho PM, Fronza BM, Nikaido T, Tagami J. Self-etch adhesive systems: A literature review. *Braz Dent J* 2015;26:3-10.
- Chai Z, Li F, Fang M, Wang Y, Ma S, Xiao Y, Huang L, Chen J. The bonding property and cytotoxicity of a dental adhesive incorporating a new antibacterial monomer. *J Oral Rehabil* 2011;38:849-856.
- Henn S, Nedel F, de Carvalho RV, Lund RG, Cenci MS, Pereira-Cenci T, Demarco FF, Piva E. Characterization of an antimicrobial dental resin adhesive containing zinc methacrylate. J Mater Sci Mater Med 2011; 22:1797-1802.
- Feitosa SA, Palasuk J, Kamocki K, Geraldeli S, Gregory RL, Platt JA, Windsor LJ, Bottino MC. Doxycycline-encapsulated nanotube-modified dentin adhesives. J Dent Res 2014;93:1270-1276.

- Li F, Weir MD, Fouad AF, Xu HHK. Effect of salivary pellicle on antibacterial activity of novel antibacterial dental adhesives using a dental plaque microcosm biofilm model. *Dent Mater* 2014;30:182-191.
- Melinte V, Buruiana T, Aldea H, Matiut S, Silion M, Buruiana EC. Photopolymerizable phosphate acrylates as comonomers in dental adhesives with or without triclosan monomer units. *Mater Sci Eng C Mater Biol Appl* 2014;34:176-185.
- Spencer P, Ye Q, Park J, Topp EM, Misra A, Marangos O, Wang Y, Bohaty BS, Singh V, Sene F, Eslick J, Camarda K, Katz JL. Adhesive/ dentin interface: The weak link in the composite restoration. *Ann Biomed Eng* 2010;38:1989-2003.
- Zhang K, Cheng L, Imazato S, Antonucci JM, Lin NJ, Lin-Gibson S, Bai Y, Xu HH. Effects of dual antibacterial agents MDPB and nano-silver in primer on microcosm biofilm, cytotoxicity and dentine bond properties. J Dent 2013;41:464-474.
- Imazato S, Ma S, Chen JH, Xu HH. Therapeutic polymers for dental adhesives: Loading resins with bio-active components. *Dent Mater* 2014;30:97-104.
- André CB, Gomes BP, Duque TM, Stipp RN, Chan DC, Ambrosano GM, Giannini M. Dentine bond strength and antimicrobial activity evaluation of adhesive systems. *J Dent* 2015;43:466-475.
- André CB, Gomes BPFA, Duque TM, Rosalen PL, Chan DCN, Ambrosano GMB, Giannini M. Antimicrobial activity, effects on Streptococcus mutans biofilm and interfacial bonding of adhesive systems with and without antibacterial agent. *Int J Adhes Adhes* 2017;72:123-129.
- Imazato S, McCabe JF. Influence of incorporation of antibacterial monomer on curing behavior of a dental composite. J Dent Res 1994;73:1641-1645.
- Imazato S, Russell RR, McCabe JF. Antibacterial activity of MDPB polymer incorporated in dental resin. J Dent 1995;23:177-181.
- Cheng L, Weir MD, Zhang K, Arola DD, Zhou X, Xu HH. Dental primer and adhesive containing a new antibacterial quaternary ammonium monomer dimethylaminododecyl methacrylate. *J Dent* 2013;41:345-355.
- 24. Cocco AR, Rosa WL, Silva AF, Lund RG, Piva E. A systematic review about antibacterial monomers used in dental adhesive systems: Current status and further prospects. *Dent Mater* 2015;31:1345-1362.
- Liang X, Huang Q, Liu F, He J, Lin Z. Synthesis of novel antibacterial monomers (UDMQA) and their potential application in dental resin. *J Appl Polym Sci* 2013;129:3373-3381.
- Zhou H, Liu H, Weir MD, Reynolds MA, Zhang K, Xu HH. Threedimensional biofilm properties on dental bonding agent with varying quaternary ammonium charge densities. *J Dent* 2016;53:73-81.
- Brambilla E, Ionescu A, Fadini L, Mazzoni A, Imazato S, Pashley D, Breschi L, Gagliani M. Influence of MDPB-containing primer on Streptococcus mutans biofilm formation in simulated class I restorations. J Adhes Dent 2013;15:431-438.
- Pinto CF, Paes Leme AF, Ambrosano GM, Giannini M. Effect of a fluoride- and bromide-containing adhesive system on enamel around composite restorations under high cariogenic challenge in situ. J Adhes Dent 2009;11:293-297.
- Pinto CF, Berger SB, Cavalli V, Da Cruz SE, Goncalves RB, Ambrosano GM, Giannini M. In situ antimicrobial activity and inhibition of secondary caries of self-etching adhesives containing an antibacterial agent and/or fluoride. *Am J Dent* 2015;28:167-173.
- Uysal T, Amasyali M, Ozcan S, Koyuturk AE, Sagdic D. Effect of antibacterial monomer-containing adhesive on enamel demineralization around orthodontic brackets: An in-vivo study. *Am J Orthod Dentofacial Orthop* 2011;139:650-656.
- Kitagawa H, Izutani N, Kitagawa R, Maezono H, Yamaguchi M, Imazato S. Evolution of resistance to cationic biocides in Streptococcus mutans and Enterococcus faecalis. *J Dent* 2016;47:18-22.
- Schupbach P, Lutz F, Finger WJ. Closing of dentinal tubules by Gluma desensitizer. Eur J Oral Sci 1997;105:414-421.
- Imazato S. Antibacterial properties of resin composites and dentin bonding systems. *Dent Mater* 2003;19:449-457.
- Ergucu Z, Hiller KA, Schmalz G. Influence of dentin on the effectiveness of antibacterial agents. J Endod 2005;31:124-129.
- Schmidlin PR, Zehnder M, Gohring TN, Waltimo TM. Glutaraldehyde in bonding systems disinfects dentin in vitro. J Adhes Dent 2004;6:61-64.
- Felton D, Bergenholtz G, Cox CF. Inhibition of bacterial growth under composite restorations following GLUMA pretreatment. J Dent Res 1989;68:491-495.
- Manabe A, Hasegawa T, Chigira H, Itoh K, Wakumoto S, Nakayama S, Tachikawa T. Morphological changes of rabbit skin by application of dentin primer. *Dent Mater J* 1990;9:147-152.
- 38. Schweikl H, Schmalz G, Göttke C. Mutagenic activity of various dentine

bonding agents. Biomaterials 1996;17:1451-1456.

- Fardal O, Turnbull RS. A review of the literature on use of chlorhexidine in dentistry. J Am Dent Assoc 1986;112:863-869.
- Löe H, Rindom Schiøtt C. The effect of mouthrinses and topical application of chlorhexidine on the development of dental plaque and gingivitis in man. J Periodontal Res 1970;5:79-83.
- Siqueira JF, Paiva SSM, Rocas IN. Reduction in the cultivable bacterial populations in infected root canals by a chlorhexidine-based antimicrobial protocol. *J Endod* 2007;33:541-547.
- 42. Brambilla E, Ionescu AC, Cazzaniga G, Ottobelli M, Mazzoni A, Cadenaro M, Gagliani M, Tay FR, Pashley DH, Breschi L. In vitro Streptococcus mutans biofilm formation on surfaces of chlorhexidine-containing dentin bonding systems. *Int J Adhes Adhes* 2017;75:23-30.
- Atalayin C, Turkun LS, Ates M, Kemaloglu H, Turkun M. Are antibacterial component additions in etchants and adhesives effective against Streptococcus Mutans? J Adhes Sci Technol 2018;32:197-206.
- 44. Xiao J, Klein MI, Falsetta ML, Lu B, Delahunty CM, Yates JR, 3rd, Heydorn A, Koo H. The exopolysaccharide matrix modulates the interaction between 3D architecture and virulence of a mixed-species oral biofilm. *PLoS Pathog* 2012;8:e1002623.
- Loesche WJ. Role of Streptococcus mutans in human dental decay. Microbiol Rev 1986;50:353.
- Koo H, Falsetta ML, Klein MI. The exopolysaccharide matrix: A virulence determinant of cariogenic biofilm. J Dent Res 2013;92:1065-1073.
- 47. Galvao LC, Rosalen PL, Rivera-Ramos I, Franco GC, Kajfasz JK, Abranches J, Bueno-Silva B, Koo H, Lemos JA. Inactivation of the spxA1 or spxA2 gene of Streptococcus mutans decreases virulence in the rat caries model. *Mol Oral Microbiol* 2016;32:142-153.
- Lemos JA, Abranches J, Burne RA. Responses of cariogenic streptococci to environmental stresses. *Curr Issues Mol Biol* 2005;7:95-107.
- van Houte J. Role of micro-organisms in caries etiology. J Dent Res 1994;73:672-681.
- Borgstrom MK, Sullivan A, Granath L, Nilsson G. On the pH-lowering potential of lactobacilli and mutans streptococci from dental plaque related to the prevalence of caries. *Community Dent Oral Epidemiol* 1997;25:165-169.
- Fritschi BZ, Albert-Kiszely A, Persson GR. Staphylococcus aureus and other bacteria in untreated periodontitis. J Dent Res 2008;87:589-593.
- Dahlen G, Samuelsson W, Molander A, Reit C. Identification and antimicrobial susceptibility of enterococci isolated from the root canal. *Oral Microbiol Immunol* 2000;15:309-312.
- Souto R, Colombo AP. Prevalence of Enterococcus faecalis in subgingival biofilm and saliva of subjects with chronic periodontal infection. *Arch Oral Biol* 2008;53:155-160.
- Koo H, Gomes BP, Rosalen PL, Ambrosano GM, Park YK, Cury JA. In vitro antimicrobial activity of propolis and Arnica montana against oral pathogens. *Arch Oral Biol* 2000;45:141-148.
- Schupbach P, Osterwalder V, Guggenheim B. Human root caries: Microbiota in plaque covering sound, carious and arrested carious root surfaces. *Caries Res* 1995;29:382-395.
- Dorn BR, Leung KL, Progulske-Fox A. Invasion of human oral epithelial cells by Prevotella intermedia. *Infect Immun* 1998;66:6054-6057.
- Marcotte H, Lavoie MC. Oral microbial ecology and the role of salivary immunoglobulin A. *Microbiol Mol Biol Rev* 1998;62:71-109.
- Yakob M, Soder B, Meurman JH, Jogestrand T, Nowak J, Soder PO. Prevotella nigrescens and Porphyromonas gingivalis are associated with signs of carotid atherosclerosis in subjects with and without periodontitis. J Periodontal Res 2011;46:749-755.
- Stingu CS, Schaumann R, Jentsch H, Eschrich K, Brosteanu O, Rodloff AC. Association of periodontitis with increased colonization by Prevotella nigrescens. J Investig Clin Dent 2013;4:20-25.

- Diaz PI, Zilm PS, Rogers AH. Fusobacterium nucleatum supports the growth of Porphyromonas gingivalis in oxygenated and carbon-dioxidedepleted environments. *Microbiology* 2002;148:467-472.
- Zilm PS, Rogers AH. Co-adhesion and biofilm formation by Fusobacterium nucleatum in response to growth pH. Anaerobe 2007;13:146-152.
- Signat B, Roques C, Poulet P, Duffaut D. Fusobacterium nucleatum in periodontal health and disease. *Curr Issues Mol Biol* 2011;13:25-36.
- Mah T-FC, O'Toole GA. Mechanisms of biofilm resistance to antimicrobial agents *Trends Microbiol* 2001;9:34-39.
- Flemming H-C, Wingender. J The biofilm matrix. Nat Rev Microbiol 2010;8:623.
- Imazato S, Ebi N, Takahashi Y, Kaneko T, Ebisu S, Russell RR. Antibacterial activity of bactericide-immobilized filler for resin-based restoratives. *Biomaterials* 2003;24:3605-3609.
- Müller R, Eidt A, Hiller K-A, Katzur V, Subat M, Schweikl H, Imazato S, Ruhl S, Schmalz G. Influences of protein films on antibacterial or bacteriarepellent surface coatings in a model system using silicon wafers. *Biomaterials* 2009;30:4921-4929.
- André CB, Rosalen PL, Galvao LCC, Fronza BM, Ambrosano GMB, Ferracane JL, Giannini M. Modulation of Streptococcus mutans virulence by dental adhesives containing anti-caries agents. *Dent Mater* 2017;33:1084-1092.
- Koo H, Schobel B, Scott-Anne K, Watson G, Bowen WH, Cury JA, Rosalen PL, Park YK. Apigenin and tt-farnesol with fluoride effects on S. mutans biofilms and dental caries. *J Dent Res* 2005;84:1016-1020.
- Mashwani ZUR, Khan T, Khan MA, Nadhman A. Synthesis in plants and plant extracts of silver nanoparticles with potent antimicrobial properties: Current status and future prospects. *Appl Microbiol Biotechnol* 2015;99:9923-9934.
- Park YK, Alencar SM, Aguiar CL. Botanical origin and chemical composition of Brazilian propolis. J Agric Food Chem 2002;50:2502-2506.
- Simone-Finstrom M, Spivak M. Propolis and bee health: The natural history and significance of resin use by honey bees. *Apidologie* 2010; 41:295-311.
- Koo H, Pearson SK, Scott-Anne K, Abranches J, Cury JA, Rosalen PL, Park YK, Marquis RE, Bowen WH. Effects of apigenin and tt-farnesol on glucosyltransferase activity, biofilm viability and caries development in rats. Oral Microbiol Immunol 2002;17:337-343.
- Koo H, Hayacibara MF, Schobel BD, Cury JA, Rosalen PL, Park YK, Vacca-Smith AM, Bowen WH. Inhibition of Streptococcus mutans biofilm accumulation and polysaccharide production by apigenin and tt-farnesol J Antimicrob Chemother 2003;52:782-789.
- Jeon JG, Klein MI, Xiao J, Gregoire S, Rosalen PL, Koo H. Influences of naturally occurring agents in combination with fluoride on gene expression and structural organization of Streptococcus mutans in biofilms. *BMC Microbiol* 2009;9:228.
- Kong M, Chen XG, Xing K, Park HJ. Antimicrobial properties of chitosan and mode of action: A state of the art review. *Int J Food Microbiol* 2010;144:51-63.
- Elsaka SE. Antibacterial activity and adhesive properties of a chitosancontaining dental adhesive. *Quintessence Int* 2012;43:603-613.
- Elsaka S, Elnaghy A. Effect of addition of chitosan to self-etching primer: Antibacterial activity and push-out bond strength to radicular dentin. J Biomed Res 2012;26:288-294.
- Lobato MF, Turssi CP, Amaral FL, Franca FM, Basting RT. Chitosan incorporated in a total-etch adhesive system: Antimicrobial activity against Streptococcus mutans and Lactobacillus casei. *Gen Dent* 2017; 65:62-66.
- Du X, Huang X, Huang C, Wang Y, Zhang Y. Epigallocatechin-3-gallate (EGCG) enhances the therapeutic activity of a dental adhesive. *J Dent* 2012;40:485-492.